

Abiotic stresses and vegetable production in the era of climate change: A review

Naeem M.Y.*; Selamoglu Z.

Received: November 2022 Accepted: February 2023

Abstract

Global warming has been recognised as a critical hazard to natural ecosystem existence. Changing climate is a continuous, multidimensional process of changes in climatic conditions that affect the abiotic and biotic aspects. This causes changes in climatic circumstances including extreme heat, rainfall amount, and temperatures, which leads to an increase in novel parasites, vegetation, and diseases, while vegetables crops must employ specific methods such as physiological, biochemcial and molecualar process to ensure optimal development during stress situations. The synthesis of hormones (phytohormones) and osmolytes is one of these processes. Plant hormones are natural materials which influence the physiological processes of plants such as growth, development, and cell differentiation. Under normal and stressful situations, plant hormones regulate essential physiological activities. Phytohormones, which come in a variety of forms, serve a significant role in plants, either on their own or in concert with one another. The type and concentration of such substances in plants have a significant impact on plant activities and functions. The current study describes phytohormones' possible involvement in fostering plant growth and development during stressful conditions.

Keywords: Abiotic Stress, Climate Change, Vegetable Production.

Department of Plant Production and Technologies, Faculty of Agricultural Sciences and Technologies, Nigde Omer Halisdemir University Nigde, 51200, Turkey *Corresponding author's Email: yasir.naeem91@yahoo.com

Introduction

The word "vegetable" includes any form of plant life or plant product in its widest definition. In a more limited definition, it alludes to the fresh, palatable component of an herbaceous plant that is ingested raw or processed (Ward, 2016). Minerals, carbohydrate, calcium, and proteins are abundant in vegetables. Vegetables are the greatest assets for alleviating micronutrient malnutrition, and they give substantially greater revenue as well as more opportunities per hectare to small scale farmers than agricultural foods (Kumar et al., 2011). Vegetables, on the other hand, are vulnerable to environmental fluctuations, therefore extreme temps and insufficient moisture levels are the leading reasons of low yields, as they have a significant impact on various physiological and biochemical processes.

Climate change can be defined as changes in the average of several climatic factors like temp, precipitation, humidity levels, and atmospheric gas compositions, among others, over a longer period of time and across a wider geographic region. Changes in climatic factors such air as temperature, excess precipitation patterns, UV radiation, and increased occurrence of extreme weather events such as droughts and floods are playing a dominant role risk to tropical vegetable production (Tirado et al., 2010). Vegetable crops are extremely susceptible to climate fluctuations, and rapid temperature increases and also erratic precipitation at any stage of agricultural production can disrupt normal growing, blooming, pollination, and fruit development, leading to a drop in crop production (Afroza *et al.*, 2010).

Changing climate as well as its unpredictability are key concerns impacting agricultural productivity. particularly vegetable crops. Fruit and vegetable yield is expected to be reduced resulting in a short growth period, that would have a detrimental influence on development and growth, especially due to final extreme heat and limited water accessibility (Venkateswarlu and Shanker, 2012). Global warming and unpredictability have created more ambiguities and dangers, putting additional vegetable limits on environments. Vegetable production crop prices may rise as a result of climate change (Abdelmageed et al., 2014). Furthermore, changes in climate are promoting pathogen proliferation and the emergence of new types of pest species, as well as fungal, bacterial, and viral illnesses. As a result, the aim of the article is to evaluate the consequences of climate change on various vegetable production.

Climate change and Abiotic stress

Plants are subjected to a variety of abiotic challenges as a result of global warming and changing climatic circumstances. Droughts, excessive salinity, harsh temps, micronutrient deficiencies, and metals sensitivities are among abiotic factors that affect crop growth (Newton *et al.*, 2011). Such variables might even have a detrimental impact on soil fertility and contribute to increased soil erosion (de la Pena and Hughes, 2007).

Cold stress and vegetables

Cold stress is a primary abiotic stress that reduces crop yields by reducing the quality and longevity of the crop after harvest. Cold stress has an influence on all aspects of cellular activity in plants. The metabolic process of plants is hindered by cold temperatures. Several tropical and subtropical crops, mostly vegetables, might suffer acute freezing harm and even mortality if subjected temperatures of 1–10°C. Plants physiological process such as nutrition, photosynthesis, water potential, and respiration, among others, are significantly harmed (Jouyban et al., 2013). It could harm plant cells and induce yellowing leaves, mitochondrial dysfunction, apoptosis, alterations in enzyme concentration, and variations in cytoplasmic fluidity in vegetable. As a result, cold stress has a significant influence on plants, producing chilling injury, cellular membrane degradation, and a decrease in net photosynthetic, as well as a devastating effect on vegetable crop output. Reduced plant growth, irregular fruit development, interior discoloration (vascular browning), increased sensitivity to decay, and plant mortality are all symptoms of cellular damage altered metabolism. and Freezing stress reduces photosynthesis performance and improves leaf electrolyte leakage watermelon in seedlings (Shirani Bidabadi and Mehralian, 2020).

Heat stress and vegetables

Since 2021, the globe temperature is projected to climb by 1.4-5.8°C result of climate change (Arora et al., 2005). Heat stress lowered photosystem II activity and water holding capacity (Li et al., 2014). Reactive oxygen species (ROS) are produced in cells amid heat stress, and oxidative stress, together with increased hydrogen peroxide, malondialdehyde, and superoxide anions, is a major hazard to vegetable production (Yuan et al., 2016). Even though the nutrient composition has a variable influence, a 4°C rise in temperature is anticipated to lower the production of vegetables and legumes by 31.5 percent, using 20°C as the baseline temperature (Scheelbeek et al., 2018). A large change in temperature due to global warming over time would result in decreased rainfall, air velocity, and snowpack, reducing the duration of the growth period for plants and significantly impacting crop quality and agricultural output. Increased temperature also can contribute to drought conditions by increasing water loss through transpiration and evaporation. Extreme temps also affect germination seeds, plant of development, and maturation, as well as cause permanent drought stress that can result in mortality (Takahashi et al., 2013).

Salinity stress and vegetables

In several salt-affected places, salinity is a severe concern that limits the development and yield of vegetable crops. The most vegetable crops possess lower salt tolerance, especially in the range of 1-2.5 dS/m, although this diminishes when saline water is used for irrigation (Machado and Serralheiro, 2017). Salt-tolerant vegetable plants reduce salinity stress by altering leaf growth and perspective, improving root progression to get accessibility to lower waterbodies, producing osmolytes, and triggering a range of stress genetics and antioxidant (Tuteja, 2007). Salinity has an impact on plant morphological development, physiological state, and production, and it impacts a large amount of arable land. Salinity causes tomato plants' leaf area and dry matter content to decrease. Furthermore, the leaves were shown to be significantly vulnerable to salt stress than the fruits because they had higher proline and Na+ (Babu et al., 2012). Under salt stress, there was an elevation in protein, mineral, dietary fibers, vitamin c. phenolic content, and flavaoids in the leaves (Sarker et al., 2018). Salinity increased the nutritional content of radish seedlings, and its sprouting under environmental salinity stress resulted in the development of beneficial substances in the plant food (Yuan et al., 2016). Salinity affects dry matter production, leaf area, relative growth rate, and net assimilation rate, according to Lopez et al. (2011). In bean plants, salinity stress causes growth and photosynthetic activity to be suppressed, as well as alterations in stomata conductance, quantity, and size. and also lowers transpiration and the cell water potential (Kaymakanova et al., 2008).

Flooding stress and vegetables

Flooding seems to be another significant abiotic stress that has a major impact on the operations and productivity of vegetable crops, that are often regarded as flood-prone crops (Parent et al., 2008). Flooding usually results in an oxygen (O_2) deficit, that is caused by the sluggish exchange of gases in water and O_2 use by microbes and plant roots. Certain plants, especially tomatoes, are extremely susceptible to floods, and genetic diversity in this trait is restricted. Flooding damages plants in general because it reduces oxygen in the root zone. inhibiting aerobic activities. Endogenous ethylene accumulates in flooded tomato plants, causing harm to the crops (Drew et al., 2009). Onion is also susceptible to floods at bulb growth, with production losses of up to 30%. The plant responses to environmental factors are dependent on the growing period as well as the duration and intensity of the stimuli (Kumar, 2017) These challenges are the leading cause of reduction in yield of much more than 50% globally.

The physiology of vegetable plants is affected by flooding. Reduced stomatal closure is one of the first physiological reactions of plants to soil flooding (Folzer et al., 2006). Flooding has an adverse influence on plant vegetative and reproductive development because it affects physiological functioning (Pautasso et al., 2012). Floods facilitate the spreading of water-borne diseases, severe droughts expose plants to infections, and storms aid spores' distribution by the wind (Gibbs and Greenway, 2008). Root respiration is hampered in waterlogged conditions as soil air is substituted by water, resulting in poor nutrient absorption and an increased risk of soil borne illnesses.

Drought stress and vegetables

Drought is defined as a state of stress caused by a lack of water. Drought is a climatic term that refers to a period of time when there is less rainfall or water availability. Drought stress occurs in plants if environmental circumstances cause a decline in the total of soil water, resulting in continual water loss via transpiration or evaporation (Ashkavand et al., 2018). Drought is the most significant result of global warming, and it is accountable for the world's worst mass starvation in the last century. Drought affects over % of the world's agricultural regions (Bot et al., 2000). Also has an impact on the soil ecosystem's functionality, form, and fertility (Liu et al., 2010). Due to increased evapotranspiration, reduced precipitation, and lowered soil microbial activities. significant water stress with combined an increasing temperature will affect vegetable crop productivity and quality. This will result in an increase in solute concentration and eventually a sinking of the water content, interrupting membranes and photosynthetic activity processes, and ultimately cell death (Yusuf, 2012). Numerous vegetables (cucumbers, melons, pumpkins, squashes, lima beans, snap beans, peas, chili, sweet corn, and tomato) are susceptible to water stress through blooming and fruiting. Potato tubers are extremely

drought-sensitive, and even mild water shortage has a negative impact on production capacity (Luoh et al., 2014). Under water stress, the development of vegetable seeds such as onion, okra, and potato tubers, as well as the sprouting of potato tubers, are adversely affected. Under water deficiency conditions, productivity and quality in tuber, root, and bulb vegetable crops are considerably reduced because the transfer of carbohydrates from the leaves to the storage organs is inhibited (Kusvuran et al., 2016).

Role of phytohormones for stress tolerance

Stress has a significant detrimental influence on agricultural output because it disrupts plant processes caused by hormonal and nutritional irregularities. Plants, being sessile creatures, are subjected to harsh environmental circumstances. Droughts or floods, flooding. excessive salt, rainwater higher or lower temp, toxic substances, and radiation from the sun are only a few of the environmental variables that are harmful to plant growth and development (Raza et al., 2020; Hossain et al., 2021). To deal with these harmful plants have developed pressures, effective detecting, communication, and response systems. The PHs, that could be identified as cellular signal molecules which function as signalling molecules in crops under trace levels and have crucially important activities in the regulation of plant responses to abiotic stresses, are one of the most visible examples of these response mechanisms

exerted by plants (Williams, 2011). Plant hormones are determining factors that help plants control their growth and survival functions. The proportion of hormones produced depends on a variety of biotic and abiotic conditions. Such significant hormones can have а growth influence on plant and development even at extremely low concentrations. Auxins, the very first phytohormone revealed (Darwin and Darwin, 1880). salicylates (SA). ethylene (ET), cytokinins (CKs), (GAs). brassinosteroids gibberellins (BRs), jasmonates (JA), abscisic acid (ABA), and strigolactones (SL), the last PHs to be found (Su et al., 2017), are among the nine types of PHs (Gomez-Roldan et al., 2008). ABA, SA, ET, and JA are among the PHs that were identified as having a critical role in the plant's response to environmental challenges (Peleg and Blumwald, 2011; Wasternack, 2014). Phytohormones such as ABA, BRs, CK, ET, GA, JA, and SA, according to Yadav et al. (2021), play a critical role in drought stress tolerance in plants by regulating cellular processes at the molecular level through cell signaling. According to significant participation in the responding to the challenges such as severe temperatures and drought, the SL and BR form a novel plant hormone of rising relevance (Nolan et al., 2020).

Phytohormones

Ethylene

Ethylene is a gaseous hormone that plays a role in a variety of morphophysiological plant functions, including the threefold response in germinating seeds, forming flowers, ripening fruits, and stimulating plant reactions to external stimuli (Awan et al., 2017). It's important for changing how plants react pathogens, external mechanical to injury, Ultraviolet rays, and nutritional shortage. Numerous incidences of ET accumulation have been documented in reaction to mechanochemical injury (Abeles et al., 1992; Kendrick and Chang, 2008). Ethylene is a growth regulator, the more of it is produced, the slower the plant grows and the sooner it reaches maturity (Nadeem et al., 2010a). Ethylene, which is involved in a variety of plant activities, has a deleterious influence on plant growth and development when levels are high, especially under stressful situations (Nadeem et al., 2010a). Root growth stimulation at low concentrations while the suppression of root elongation has been seen at high concentrations (Mattoo and Suttle 1991; Belimov et al., 2002). Ethylene has a considerable influence on rhizobial infection in legumes and plays a crucial role in legumerhizobia interaction (Penmetsa and Cook, 1997). Ethylene was shown to prevent nodulation either administered immediately as a gas or in the form of its precursor, such as ACC (Yuhashi et al., 2000). It also has a detrimental impact on plant growth by inducing a variety of problems including leaf abscission, senescence, epinasty, and chlorophyll degradation (Shibli et al., 2007; Nadeem et al., 2010b).

The gaseous hormone ethylene (ET) and the oxylipin-based hormone jasmonate (JA) work together to help plants respond to biotic and abiotic challenges in a variety of ways (Zhu, 2014). ET is capable of activating and has increased heat stress tolerance (Clarke et al., 2009). Crop seed development is severely threatened by extreme temps during the reproductive period. ethylene production is variably controlled in floral and fruit tissues in pea during heat stress to maximize resource allocation in reproductive tissues (Savada et al., 2017). Ethylene confers thermostability and mitigates heat-induced negative effects in rice (Wu and Yang, 2019). As a result, plant species and tissues differed in their physiological, biochemical, and molecular activities of ET in response to heat stress. Further research is needed to have a better understanding of ET's involvement in plant thermotolerance. ET is required for the initiation of the anti-necrotrophic pathogen innate immunity.

Gibberellins

The GAs is a long-standing and quite well class of regulating hormones that involved several are in plant physiological processes including as germination, inter-nodal seed elongation, induced blooming, and fruit formation (Islam et al., 2021). There are around 250 GAs representatives documented, however only a couple of them are physiologically active and play various functions in plant growth. GA's receptor protein is nucleus confined GID1, and binding of GA to receptor protein aids in interaction with the

repressor DELLA protein. GA regulates signaling by disintegrating repressor proteins, which result in a wide variety biological and physiological of responses (Lo et al., 2017). GA regulates redox homeostasis in H. vulgare by increasing electron mobility (Mark et 2016). The phytohormones al.. communicate with one another, and this association can be detrimental, as shown with gibberellins (Heinrich et al., 2013), where a high amount of jasmonic acid gibberellin inhibits production. Nicotiana attenuata stem elongation is inhibited due to a reduction in gibberellins. Researchers observed that the total quantity of jasmonic acid was responsible for the suppression of gibberellins (Fleet and Sun, 2005). Gibberellins are vital in growth and development of plants because they influence seed germination, root/shoot elongation, flowering. and fruit patterning (Fleet and Sun, 2005; Shani et al., 2013). The use of gibberellins has improve been shown to wheat production in salty conditions (Parasher and Varma, 1988). Afroz et al. (2005) demonstrated that the use of gibberellins improved the photosynthetic rate and nitrogen fixation of salt stress mustard plants. Plants subjected to extreme heat stress exhibit elevated acidification of extracellular solution and reduced proteolysis, which may be controlled by the administration of gibberellins (Aleksandrova et al., 2007).

Auxins

Auxins, like CKs, are natural plant growth hormones that are usually

engaged in root/shoot development and comparative growth (Sachs, 2005). Auxins collaborate with CKs in a variety of cellular and physiological functions, including cell cycle regulation, cell growth, apical domination, leaf growth, and embryonic development during seed maturation (Tromas et al., 2009; Jurado et al., 2010). The Auxin Response Factor (ARF) group has been discovered as transcriptional regulators which regulate auxin's effects; for instance, several ARFs have been shown to play an important role in lateral root formation (Wilmoth et al., 2005). According to research by (Marin et al., 2010), the collective activity of micro RNAs impacted the amount of lateral root miR390 TAS3development, and derived transacting short interfering RNAs, and Auxin Responsive Factors constituting an auxin-responsive regulatory network. When soil water tension is low, auxins accumulate in root system and are transferred to the leaves. When soil water tension is low, auxins accumulate in root system and are transferred to the leaves. These auxins bind to receptors on the surface of the membrane of stomatal cells, cell resulting increased stomatal in conductance (Babu et al., 2012). Auxins are well-known phytohormones that help plants cope with environmental stresses such as salt, underwatering, and soil acidity (Salama and Awadalla, 1987; Ribaut and Pilet, 1991).

Salicylic acid

Salicylic acid is among the most important phenolic natural growth

factors, and has been identified in practically all plant species belonging to diverse groupings. It has been discovered that it regulates a variety of physiological and biochemical processes in plants. It has been extensively explored in plants to aid in the development of systemic acquired resistance to a variety of diseases (Misra and Saxena, 2009; Kolaksazov et al., 2013). Nevertheless, depending on the amount and period of osmotic stress, the SA involvement in plants treated to salt and osmotic stressors remained relatively equivocal in many plant species. Similarly, SA exogenous treatment reduced salinity-related effects (Horváth et al., 2007). Cold stress causes a rise in the quantity of jasmonates and salicylic acid in wheat (Kosova et al., 2012; Kolaksazov et al., 2013). Furthermore, plants that are many developmental engaged in processes such as seed germination, root growth, and senescence have been shown to contain jasmonic acid and salicylic acid (Chen et al., 2009b; Wasternack and Hause, 2013).

Abscisic acid

The plant hormone abscisic acid was identified in the early 1960s and was determined to be capable of breaking seed dormancy (Cornforth *et al.*, 1965). It was eventually discovered that it may have a function in plant growth and stress adaptation responses (Guschina *et al.*, 2002; Iqbal, 2015a). Abscisic acid (ABA) is the most researched stressresponsive hormone in plants, and it is implicated in a variety of stressors such

9

as osmotic, drought, and cold stress and Blumwald. (Peleg 2011: Wasilewska et al.. 2008). Seed development and seed dormancy, as well as water management in the plant body via stomatal opening and closure, and plant responses to external challenges, are all affected by ABA. When there is a lack of water. ABA is mainly synthesized in the roots and transferred to the leaves through the xylem, resulting in a rise in leaf concentrations. That rise in ABA functions as a stimulus in guard cells, triggering a signalling pathway that regulates guard cell turgor (Danquah et al., 2013; Awan et al., 2017). The production and translocation of ABA in the abiotic stress response causes stomatal conductance and a reduction in transpiration rate, limiting cell development (Peleg and Blumwald, 2011). Dar et al. (2017) found that ABA upregulates hormone-responsive transcription and factors that phosphatases and kinases in ABA signaling mediate quick responses to diverse abiotic stressors. Furthermore, ABA not just to mediates physiological reactions by its own signaling, but it also regulates physiological responses in plants subjected to abiotic stressors by connecting with other PHs (He et al., 2018). It's important to note that ABA activates and galvanizes a variety of physiochemical defence mechanisms, including proline biosynthetic pathway, antioxidants, ROS detoxifying enzymes, heat shock proteins, and unsaturated fatty acids, as well as the strengthening of cuticular waxes, all of which help plants to mitigate the negative effects of abiotic stress (Liu *et al.*, 2018; Yin *et al.*, 2018.

Jasmonates

Jasmonates are a large category that includes a variety of substances such as jasmonic acids (JAs), jasmonic acid methyl ester (JAME), and others. Jasmonates are distributed all through plant, but levels are particularly high at the stem apical, root tips, immature fruits, and young leaves, which are fragile growth portions (Wasternack, 2007). JAs are thought to play a role in embryo and reproductive organ development, sex determination, seed germination and seedling development, growth, root fruit ripening, leaf movements and senescence, gravitropism, trichome formation, and tuber development (Wasternack and Hause. 2013; Wasternack, 2014). Because of their strong protective plants against stress potential on (Takeuchi et al., 2011), research have focused on JAs, which play an important role in abiotic stress tolerance. Furthermore, plant motions associated to adapting seasonal and circadian cycles influenced by JAs-mediated are secondary metabolites synthesis (Chen et al., 2006). It is well-established that jasmonates have a role in abiotic stress control in Carica papaya (Mahouachi et al., 2007), citrus, and Arabidopsis thaliana (A. thaliana) (Arbona et al., 2010; Brossa et al., 2011).

Strigolactones

Strigolactones (SLs) are carotenoidderived PHs that are released by 80% of plants that claim to have a symbiotic interaction with arbuscular soil mycorrhizae (Akiyama and Hayashi, 2006). Such PHs were first recognized as an "ecological signal" for parasite seed development and a symbiotic link between helpful bacteria and plants. Those substances were generally dubbed strigolactones because this obligatory hemiparasite plant requires a live host for germination and initial growth (Cook et al., 1972). Numerous strigolactone analogs, including GR5, GR7, and GR24 (the most active), were produced synthetically in several plants, and subsequent descriptions also had characterized one 's fully functioning roles in a variety developmental stage, as root development, such shoot branching, leaf senescence, reproductive development. and controlling the architectural style of plant organs (Gomez-Roldan et al., 2008; Kapulnik et al., 2011). Additionally, SLs have an effect on the quantitative growth of the symbiotic nitrogen-fixing bacteria in root nodules (Rochange et al., 2019). SLs are important in controlling many physiological and morphological activities throughout plant adaptation to abiotic stressors such as salt, drought, nutritional deficiency, temperature, and pathogenic assail, according to several studies (Marzec et al., 2013). SLs were also shown to have an important function in nitrogen and phosphorus deprivation, causing changes in root and shoot architecture and encouraging rhizobial bacteria and AM fungus symbiosis (Marzec, 2016).

Cytokinin

The CKs are important plant PHs that act as ABA opponents and auxin antagonists/synergists in a range of methods that affect plant growth, development, and tolerance to abiotic stressors (Pospilová, 2003; Danilova et al., 2016). The usage of cytokinins has also been found to be beneficial in modulating the plant's sensitivity to external stress (Ha et al., 2014). Whenever a plant is subjected to salt stress, CKs are triggered. CKs can improve salt stress resistance by associating with other plant hormones like auxins and ABA (Iqbal et al., 2006). Because the photosynthetic system is destroyed in suspectable plants under low photosynthesis settings, there is a higher drop in photosynthesis in susceptible plants than in stress tolerant transgenic plants during water stress. As result. CKs may а preserve photosynthetic systems in transgenic plants, adding to stress tolerance (Rivero et al., 2010). Plant intrinsic CKs are nucleotide derivative with isoprenoid or aromatic side chains that are present in lesser amounts in plants (Sakakibara, 2006). Cytokinin is thought to improve wheat salt tolerance by interacting with other plant hormones and regulating and detoxifying harmful ions and reactive oxygen species. It is also shown that tobacco can withstand dryness by accumulating cytokinin Rivero et al. (2007). Cytokinin has been found to alter nitrogen metabolism in plants by increasing nitrate reductase activity (Sakakibara., 2006; Sykorova et al., 2008). Plant resistance to salt and

drought was also found by Barciszewski et al. (2000) owing to cytokinin buildup. Other authors have found on the role of exogenous cytokinin application in plant stress tolerance (Gupta et al., 2003; Yang et al., 2005), which is regarded as a cost-effective and simple strategy for inducing stress tolerance in a variety of crops (Torres- Garcia et al., 2009). According to Ketabchi and Shahrtash (2011), cytokinins can help protect plants against infections. The use of cytokinin lowers the deleterious effects of *Fusarium moniliforme* on maize seedlings significantly.

Melatonin

Melatonin is a differentially expressed chemical that has a wide range of effects on plants. For the first time in 1995, two of researchers groups discovered melatonin (N-acetyl-5methoxytrytamine) vascular in plants (Wang et al., 2017). It is regarded as an antioxidant with considerable antioxidant properties in the management of reactive oxygen species (ROS) (Raza et al., 2020). Plant melatonin is also engaged in a variety of physiological functions, including growth, roots, seed germination, photosynthesis, and stress resistance (Arnao and Hernández-Ruiz, 2015). The discovery of the first plant melatonin receptor has paved the way for this regulating molecule to be classified as a novel plant hormone. Melatonin has also been considered as a plant master regulator due to the range of its activities. Additional activities regulated by it include ripening or senescence, the internal biological clock,

and parthenocarpy. Lastly, melatonin is an endogenous plant bio-stimulator that protects plants from abiotic and biotic stresses (Sun et al., 2015). The absence of a recognized sensor is among the most significant constraints in research on melatonin in plants. Growth stimulation, induction. tropism, roots seed germination promotion, photosynthesis, efficiency optimization, and leaf water/CO₂ exchange are all affected by melatonin (Sharif et al.. 2018). Phytomelatonin, like other plant hormones, regulates H₂O₂ synthesis by controlling stomatal closure via the CAND2/PMTR1-mediated signaling pathway (Arnao and Hernández-Ruiz, 2019).

Brassinosteroids

Brassinosteroids, a novel class of plant hormones, affect plant development processes such as seed germination, blooming, and senescence (Rao et al., 2002). Brassinosteroids are found in plants at extremely low concentrations, and their concentration fluctuates in plant tissues, with young tissues containing more brassinosteroids than mature tissues. Chlorella vulgaris, a unicellular green alga, has also been found contain brassinosteroids to (Bajguz, 2009; Stirk et al., 2013). Over than 50 naturally occurring brassinosteroids were discovered by Kim et al. (2005) across the whole plant kingdom. Brassinosteroids are essential for a wide range of plant developmental activities; include shoot and root elongation, seed development and germination, and the formation of vascular tissue. according to Schumacher and Chory (2000). When green algae are subjected to heat stress in the presence of brassinosteroids, the level of ABA increases (Bajguz, 2009). Cell elongation, cell division. photomorphogenesis, xvlem differentiation. growth, and reproduction are the among physiological and biochemical processes regulated by them in plants (Nolan et al., 2020). Auxins are intimately connected to BRs because they modulate their coordinating transport, the tropic responses of plant organs and encouraging lateral root primordial initiation during lateral root growth (Li et al., 2005; Bao et al., 2004). BRs, unlike other PHs, are employed near synthesizing cells rather than for longdistance transport. BRs do, however, have a long-distance effect when they interact with PHs like auxin (Symons et al., 2008; Vriet et al., 2013).

Polyamines

Small aliphatic nitrogenous bases generated by cellular metabolism are known as polyamines (PAs). The PAs are not hormones, but they have been proposed as a novel category of plant growth regulators because of their role in regulating many growth and development processes as well as plant responses to abiotic stress (Liu et al., 2007; Chen et al., 2019). PAs also have role in plant embryogenesis, a organogenesis, reproductive development, leaf senescence, and fruit maturity, among other physiological processes (Igarashi and Kashiwagi,

2010; Childs *et al.*, 2017). Additionally, PAs have been shown to defend against environmental stressors in a number of investigations (Minocha *et al.*, 2014; Chen *et al.*, 2019).

The H₂O₂ generated by PA oxidation is implicated in signal transduction and biotic plant responses to and environmental stressors (Freitas et al., 2017; Mellidou et al., 2017; Sariyev et al., 2020). The PA biosynthesis process intertwined with the is ethylene biosynthetic pathway, since they both use the same precursor (Sadenosylmethionine) and compete for it. Furthermore, PA metabolism is linked to nitric oxide production (Pál et al., 2015).

Impact of abiotic stresses on vegetable production

Abiotic challenges are currently a key block for worldwide stumbling agricultural crops. A large portion of the community in developing nations whose agricultural production still persists is continually endangered by biotic and abiotic stressors. The problem is projected to worsen as a due to global warming. With the expected increase in global population and food supply, this will be vital to identify strategy to enhance crop resistance to abiotic stress conditions in order to increase productivity agricultural and food security (Calanca, 2017). Drought, waterlogging, heat, cold, and salt challenges are reviewed, as well as their negative effects vegetable on production.

Abiotic stress and Onion

Salinity Stress

Around the year 2050, it is projected that significant soil salinity would have damaged 50% of cultivated land (Wang et al., 2003). One of the primary abiotic factors that restricts agricultural growth and results in economic loss is salinity, which is induced by excessive NaCl concentrations (Capiati et al., 2006). In comparison to other vegetables, onions are the most vulnerable to salinity, as per United States Department of the Agriculture (USDA). Onion growth and photosynthetic metabolism are affected by soil salinity (Beinsan et al., 2015). Salt stress has a number of effects on plant development and output. Salt stress has two major impacts on crops: osmotic stress and ionic toxicity. Assimilate synthesis, decreased cell growth and membranes functionality, and reduced cytosolic metabolism are all secondary effects generated by the fundamental effects of salt stress (Gull et al., 2019). High salinity generates a scarcity of water in the rhizosphere, resulting in a greater salt content, as measured by the concentration of Na⁺ and Cl⁻ ions (Tuteja et al., 2012). The number of bulbs per unit area, size, and fresh weight of onion bulbs are all affected by salinity in irrigation water. It has an impact on bulbing and the quality of harvested bulbs. The fresh weight of onion bulbs at harvest is also affected by salt at various development stages (Sta-Baba et al., 2010). The flavor production and minerals content of onion bulbs are also affected by salinity. It is yet to be revealed whether or not utilizing NaCl at

specific developmental stages will impact taste accumulation and growth (Chang and Randle, 2004). H_2O_2 application increases chlorophyll content, which helps in reduction of salt stress in onions (Semida, 2016).

Drought Stress

Onion is an irrigated crop that requires a significant quantity of irrigation water to grow. Low productivity is caused by a lack of water. Onions require a lot of soil moisture to generate a lot of yield (Kadayifci et al., 2005). Onions are a shallow-rooted crop, with most penetration up to 18 cm and only a few roots extending up to 31 cm, extracting relatively little water from depths more than 60 cm. According to Srinivasa (Rao et al., 2016), the top 30 cm of soil absorbs the majority of the water, thus it's critical to maintain the soil wet to ensure the plant gets adequate water.

Onion production is increased by a constant supply of water in the needed amount. Drip irrigation is the most effective way to distribute water in drought-prone locations. Bagali et al. (2012) discovered that drip watering at shorter intervals resulted in a much larger bulb production. Onion growth phases that are subjected to soil drought stress have a significant influence on bulb output and quality. Onion bulbs' weight and size are also affected by drought. At the first stage, non-stressed and water-stressed plants generated the greatest average bulb weights of 102g and 91g, respectively, with significant differences at the 0.05 level of probability (Zayton, 2007).

Heat stress

global Rising temperatures are becoming a big worry, hurting not just plant development but also crop yield. The severity, duration, and rate of temperature rise characterize high temperature stress. The amplitude and scope of stress increase fast when the temperature rises over a specific threshold, resulting in complicated acclimation effects that rely on temperature and other environmental parameters (Wollenweber et al., 2003). When a plant is stressed by heat, the rate of seed germination decreases, lowering photosynthetic efficiency and performance. Incredibly high temperatures during the reproductive period might impact pollen viability, fertilization, and grain or fruit production (Hatfield et al., 2011). Heatrelated impairment to crop generative tissues is a major source of yield reduction in agriculture across the world (Suzuki et al., 2003). Tapetal cells lose its functionality and become dysplastic under heat stress during reproductive growth. Due to a fast temp spike during the winter period, the crop matures early, resulting in smaller bulbs. Temperature increase has been associated to lower net photosynthetic and, as a result, lower agricultural output (Mathur et al., 2011).

Cold stress

Abiotic stressors such as cold and freezing are prevalent environmental abiotic stresses that impact plant growth and development. Crops in various parts of the world are subjected to periods of exceptionally low temperatures. Plant development is hampered and production is harmed when plants are exposed to freezing and drought at the same time (Wang et al., 2016). Cold stress, that comprises temperatures as low as 20°C and as high as 0°C, impairs plant development and output while also limiting agricultural productivity. Plants are prevented from developing its full genetic potential by cold stress, which slows metabolic activities both directly indirectly through and osmotic, oxidative. other and stressors (Tommasini et al, 2008). Cold stress can result in poor germination, stunted seedlings, chlorosis, decreased leaf growth, and wilting, as well as tissue death (necrosis). Plants' reproductive development is also harmed by cold stress. Cold stress causes significant membrane damage, which is mostly produced by the severe dehydration that occurs as a result of freezing during the cold stress (Yadav, 2010). Plants undergo a variety of physiological and biochemical changes as a defense strategy against cold (0-15°C) and freezing $(0^{\circ}C)$ conditions. The average bulb weight per plant increased after being exposed to low temperatures for brief periods of time (Khokhar et al., 2007).

Waterlogging stress

Waterlogging is a significant abiotic factor that crops experience. Heavy and irregular rainfalls, as well as poor soil permeability produced by granular soil from the use of heavy agricultural machinery, are to blame (Hirabayashi *et al.*, 2013). Waterlogging reduces the

oxygen supply to submerged tissues, affecting plant growth, development, and yield (Barickman et al., 2019). Because of their shallow roots, onions are very vulnerable to waterlogging stress. In a pot experiment, the response of onion genotypes to waterlogging stress was investigated (Dubey et al, 2020). Waterlogging stress can prevent assimilates from moving from the source the sink, lowering bulb yield to (Prasanna and Ramarao. 2014). Waterlogging has a substantial influence on bulb output and yield at various development stages. Waterlogging at early development phases following transplanting and bulb initiation affects bulb quality and marketable bulb size, according to Ghodke et al. (2018).

Abiotic stress and potato

Water stress

Water stress is the most significant factor affecting agricultural output and quality. Potatoes are known to be extremely water-stressed. Water stress has different impacts on plants depending on when it occurs, how long it lasts. and how severe it is. Modifications to a lack of water cause changes in a variety of morphological, biochemical, and physiological features, the majority of which are detrimental to plant production (Minhas et al., 2018). Shoot height, root depth, leaf area, and other morphological characteristics show the impact of water stress on potato crop growth and development (Kumar et al., 2015). Nilsen and Orcutt (1996) state that one of the strongest markers for evaluating plant responses to environmental or abiotic stress is shoot growth. Water stress produces a substantial decrease in shoot height at all growth stages, according to Kumar *et al*. (2017), although it is most noticeable at the tuber start stage. Reduced cell size in shoots is caused by a decrease in cell expansion rate. Water shortages have a significant impact on leaf production, reducing the number, size, and length of leaves (Kumar *et al.*, 2015).

Under water stress, the tuber initiation stage of the potato plant is the most vulnerable, while the tuber maturity stage is the least sensitive, resulting in a drop in overall biomass output (Kumar *et al.*, 2017). Resulting from water stress, the quantity of nonmarketable tubers rises at the expense of marketable tubers (Kumar *et al.*, 2015).

Drought

Soil dehydration changes the morphology of the potato root system, leading to increased adventitious root proliferation, decreased root elongation, increased root thickness, and the production of root hairs that maintain root-soil contact in soil that shrinks away from roots during drying (Wishart et al., 2013, 2014). Drying the soil enhances the flowability of the roots at first. However, prolonged water withdrawal reduces root hydraulic conductivity (to avoid soil losses), and these changes are attributed to aquaporin activity and control (Maurel et al., 2010). Xylem embolism, a phenomenon in which air enters the xylem vessels and interrupts the sap flow, can significantly diminish root hydraulic conductivity in the plant

during extended durations of dryness (Cruiziat and Cochard, 2002). Plant roots are also thought to sense soil dryness biochemically, causing the phytohormone ABA to be synthesized and transferred to the shoot via the xylem, inhibiting leaf growth and inducing stomatal closure (Dodd, 2005). Reduced stomatal density may also cause lower transpiration rates in response to drying soil (Yan *et al.*, 2012; Sun *et al.*, 2014).

Salinity

Salinity raises the osmotic pressure of soil pore water, limiting root water intake, and has a negative impact on soil structure, lowering water infiltration, soil aeration, and oxygen delivery to the roots. Some components that are connected with salt have a higher absorption. Toxicity is caused by elements such as sodium, chlorine, and boron (Levy and Veilleux, 2007). Significant osmotic stress in the tissues is among the main effects of salinity, and it causes adverse alterations in cellular function that impede amino acid synthesis, protein metabolism, respiration, and photosynthesis (Koyro et al., 2012). Lower CO₂ absorption is another result of the combined effect of salt and drought, which increases the generation of reactive oxygen species (ROS), which are particularly destructive to nucleic acids, lipids, and proteins (Koyro et al., 2012).

Heat stress

The first factor that defines a location's potential for potato growing is

temperature. Potato tuber output is lowered by high temperature stress because assimilate availability is reduced and assimilate partitioning to tubers is reduced (Joseph et al., 2006). High night temperatures have a large impact on carbon partitioning across plant parts, with temperatures above 18°C favoring carbon diversion to above-ground portions at the expense of tubers, causing plants to grow very tall and lowering harvest index (Minhas et al., 2018). Tuberization in potatoes is a morphogenetic shift from stolon development to tuber production that is influenced by both external and endogenous factors (Singh et al., 2015). In warm areas, heat tolerance in potatoes entails strong growth, tuber initiation development, concurrent and and balanced glucose partitioning to the haulm and tubers, and starch buildup in the tubers (Kumar et al., 2007). Under heat stress. several secondary metabolites, including antioxidants with nutritional benefit. known alter dramatically (Kumar et al., 2013). Without adaptation, climate change is expected to reduce world potato crop by 18% to 32%, or by 9% to 18% with adaptation (Hijmans, 2003).

Abiotic stress and Tomato

Heat stress

Tomatoes are susceptible to high temps, and extreme heat can cause floral stomatal closure and reduce fruit output (Camejo *et al.*, 2005; Driedonks, 2018). Increased temperature had a deleterious impact on pollen grain growth, especially in the early stages, resulting in poor pollen germination and pollen tube formation (Raja al., et 2019). Photosynthesis is similarly affected by high temperatures (Nankishore and Farrell, 2016). According to Pressman et (2002), heat stress in tomato al. produced male sterility, although male sterile plants grown at 29°C were able to yield fruits after receiving pollen created at 25°C. Temperatures above 30°C limit pollen germination and pollen tube formation in tomato (Vasil, 1987). According to Kakani et al. (2005), the ideal temperature for pollen germination in vitro was 15–22°C, while the optimal temperature for pollen germination in vivo was 25°C (Dempsey, 1970).

High temperatures have an impact on the physiological activities of roots as well as the development of aboveground plant components like fruits. The heat stress changed the architecture and physiological metabolism of tomato blossoms, as well as the synthesis of substances including carbohydrates, polyamines, and proline (Alsamir et al., 2017). Increased temperature to inadequate temperature circumstances has a considerable impact on tomato reproductive development, producing proportionally more harm to anthers than to female organs (Xu et al., 2017).

Drought

According to Rao *et al.* (2016) abiotic stressors have been associated to altered shoot, root, and leaf development, as well as crop-season life cycle and duration. Khan *et al.* (2017) investigated morpho-physiological variations in tomato crop metrics during drought stress. During drought stress, the plant body's relative water content drops due to a decrease in water supply. According to the findings, reduced water inhibited photosynthetic capability, impact on energy output and, as a consequence, poor growth. For up to 6 days, moisture stress reduced the weight of tomato seedlings' shoots and roots (Zhou et al., 2019). Drought stress causes the amount of solute in the soil solution to rise, allowing water to move osmotically from plant cells. This causes a rise in solute concentrations in plant cells, lowering water potential and disrupting membranes and cell functions including photosynthesis (Rao et al., 2016). Stomatal closure in tomato cultivated under non-irrigated settings ranges from 14 percent to 73 percent, depending on the climatic conditions and cultivars (Nemeskeri and Helves, 2019).

Salinity

Salts dissolved in soil solution come into intimate contact with roots and may impair plant development because the osmotic action reduces water intake from plants, lowering the water potential of leaves and tissues. In many species, salt has the impact of not only decreasing the proportion of seeds that germinate, but also prolonging the time it takes to complete germination. At low NaCl concentrations, tomato seed germination is inhibited (Petretto et al., 2019). Plant roots are subjected to fast osmotic stress as a result of salinity stress, which results in a decrease in leaf and shoot growth. Salinity stress causes an ion imbalance, which causes necrosis and early

mortality of older leaves (Munns and Tester, 2008). Because of the reduction in plant water potential and chlorophyll production, salinity has an impact on photosynthesis (Parihar et al., 2015). Roots are unable to ingest water directly from the soil under high salinity stress conditions, resulting in hazardous salt and chloride concentrations. plant nutritional problems, and oxidative damage (Rouphael et al., 2018). Under salt stress, the tomato variety Shed displayed impacted stomatal features such stomatal density, stomatal breadth, leaf stomatal area, and leaf stomatal area index, but the Alam variety remained unharmed (Guo et al., 2018).

Conclusion

In the last few years, there has been a significant shift in the climate, that has an immediate impact on the production, productivity, and quality of vegetable crops. As per studies, it appears that it will have a significant influence on global food and nutrition security in the coming years. As such, to ensure that the quality and quantity of vegetables, the effects of climate change can be mitigated by adopting appropriate approaches/strategies such as developing improved cultivars tolerance to biotic and abiotic stresses; conserving forests. soil. and water; utilizing renewables, and protected cultivation.

To adapt to the changing environment, plants evolve many techniques, one of which is the synthesis of hormones. Such plant hormone contributes in growth and development of plants by speeding up processes in plants. Also, it is clear from the foregoing section that all these plant hormones communicate with others, and so this engagement can be good or detrimental. The majority of unfavorable reactions to plant hormones are caused by large concentrations, which directly impact the specific plant process. It had an influence on both the vegetative and reproductive stages of plants, and its influence varied depending on the species. Furthermore, consistent with earlier study findings, it is discovered that extreme drought stress encountered by diverse plant species impeded the photosynthetic process in plants. resulting in lower plant growth and output. Under various abiotic conditions, several species of plants showed a rise in ROS, RNS, and proline content, as well as a decline in development and a drop in photosynthetic pigments. Plants have been observed to lose dry matter, pigmentation intensity, and phenolics as a result of saline condition. Phyto regulators such as plant hormones, osmolytes, bio stimulants, and others have been found to give plant tolerance to various abiotic stimuli by boosting protein, proline, and antioxidant metabolism in many plant species.

The research from a community of scholars mentioned that the presence of a hormone, its use for a clear objective, through the use of a phytohormone producing strain are some of the most important aspects to be considered when trying to improve plant growth and development under normal and stressful conditions. There is a large deficit in the knowledge of such features that has to be covered by finding out information at the vast system level. The quantity and frequency of plant hormones administration must be studied.

References

- Abdelmageed, A.H., Gruda, N. and Geyer, B., 2014. Effects of temperature and grafting on the growth and development of tomato plants under controlled conditions. Rural Poverty Reduction through Research for Development and Transformation.
- Abeles, F.B., Morgan, P.W. and Saltveit, M.E.Jr., 1992. Ethylene in Plant Biology, 2nd Edn. New York, NY: Academic.
- Afroz, S., Mohammad, F., Hayat, S. and Siddiqui, M.H., 2005. Exogenous application of gibberellic acid counteracts the ill effect of sodium chloride in mustard. *Turkish Journal of Botany*, 29, 233–236
- Afroza, B., Wani, K.P., Khan, S.H., Jabeen, N., Hussain, K., Mufti, S. and Amin, A., 2010. Various technological interventions to meet vegetable production challenges in view of climate change. Asian Journal of Horticulture, 5, 523-529.
- Akiyama, K. and Hayashi, H., 2006. Strigolactones: chemical signals for fungal symbionts and parasitic weeds in plant roots. *Annals of Botany*, 97, 925–931. DOI: 10.1093/aob/mcl063
- Aleksandrova, I.F., Lebedeva, A.S.
 and Petrunina, N.A., 2007.
 Modulating infl uence of gibberellic acid in hyperthermia in wheat grains.
 2nd International symposium on

plant growth substances: intracellular hormonal signaling and applying in agriculture, 8–12 October, 2007, Kyiv, Ukraine

- Alsamir,M.,Ahmad,N.M.,Mahmood,T. and Trethowan,R.,2017.Morpho-PhysiologicalTraitsLinked to High TemperatureStressToleranceinTomatoIycopersicumL.).American.Journalof Plant Sciences,8(11),2681.
- Arbona, V., Argamasilla, R. and Gomez-Cadenas. 2010. A., Common and divergent physiological, hormonal and metabolic responses of Arabidopsis thaliana and Thellungiella halophilato water and salt stress. Journal of Plant Physiology, 167, 1342-1135. DOI: 10.1016/j.jplph.2010.05.012

Arora, M., Goel, N.K. and Singh, P., 2005. Evaluation of temperature trends over India/Evaluation de tendances de température en Inde. *Hydrological Sciences Journal*, 50, 81–93.

- Arnao, M.B. and Hernández-Ruiz, J.,
 2015. "Melatonin: synthesis from tryptophan and its role in higher plants," in Amino Acids in Higher Plants, ed J.P. F. D'Mello (Boston, MA: CAB International), 390–435. DOI: 10.1079/9781780642635.0390
- Arnao, M.B. and Hernández-Ruiz, J.,
 2019. Melatonin and reactive oxygen and nitrogen species: a model for the plant redox network. *Melatonin Research*, 2, 152–168. DOI: 10.32794/11250036

- **P.**. Zarafshar, Ashkavand, **M.**. Tabari, M., Mirzaie, J., Nikpour, A., Bordbar, S.K., Struve, D. and Striker G.G., 2018. Application of SIO2 nanoparticles AS pretreatment alleviates the impact of drought on the physiological performance of Prunus mahaleb (Rosaceae). Boletin de la Sociedad Argentina de Botanica, 53, 207
- Awan, F.K., Khurshid, M.Y. and Mehmood, A.J.I.J.I.R.B., 2017.
 Plant growth regulators and their role in abiotic stress management. *International Journal of Innovative Research in Biosciences*, 1, 9–21.
- Babu, M.A., Singh, D., Gothandam,
 K.M., 2012. The effect of salinity on growth, hormones and mineral elements in leaf and fruit of tomato cultivar PKM1. Journal of Animal and Plant Sciences, 22, 159–164
- Bagali, A.N., Patil, H.B., Guled, M.B., and Patil, R.V., 2012. Effect of scheduling of drip irrigation on growth, yield and water use efficiency of onion (Allium cepa L.). *Karnataka Journal of Agricultural Sciences*, 25(1).
- Bajguz, A., 2009. Isolation and characterization of brassinosteroids from algal cultures of *Chlorella vulgaris* Beijerinck (Trebouxiophyceae).

Journal of Plant Physiology, 166, 1946–1949

Bao, L., Locovei, S. and Dahl, G.P., 2004. Annexin membrane channels are mechanosensitive conduits for ATP. *FEBS Letters*, 572, 65–68. DOI: 10.1016/j.febslet.2004.07.009

- Barciszewski, J., Siboska, G., Rattan, S.I.S. and Clark, B.F.C., 2000. Occurrence, biosynthesis and properties of kinetin (N6furfuryladenine). *Plant Growth Regulation*, 32, 257–265
- Barickman, T.C., Simpson, C.R. and Sams, C.E., 2019. Waterlogging causes early modification in the physiological performance, carotenoids, chlorophylls, proline, and soluble sugars of cucumber plants. *Plants*, 8(6), 160.
- Beinşan, C., Sumalan, R. and Vâtcă,
 S., 2015. Influence of Salt Stress on
 Quality of some Onion (*Allium cepa*L.) Local Landraces. Bulletin
 USAMV series Agriculture, 72, 2.
- Belimov AA, Safranova VI, Mimura
 T., 2002. Response of spring rape (Brassica napus) to inoculation with
 PGPR containing ACC-deaminase
 depends on nutrient status of plant.
 Can J Microbiol 48:189–199.
- Bot, A.J., Nachtergaele, F.O. and Young, A., 2000. Land Resource Potential and Constraints at Regional and Country Levels; World Soil Resources Reports 90; Land and Water Development Division, FAO: Rome, Italy.
- Brossa, R., López-Carbonell, M., Jubany-Mar,í T. and Alegre, L., 2011. Interplay between abscisic acid and jasmonic acid and its role in water-oxidative stress in wild-type, ABA-deficient, JA-deficient, and ascorbate-deficient Arabidopsis plants. *Plant Growth Regulation*, 30, 322–333. DOI: 10.1007/s00344-011-9194-z

- **Calanca, P.P., 2017.** Effects of abiotic stress in crop production. In Quantification of climate variability, adaptation and mitigation for agricultural sustainability (pp. 165-180). Springer, Cham.
- Camejo, D., Rodríguez, P., Morales, M.A., Dell'Amico, J.M., Torrecillas, A. and Alarcón, J.J., 2005. High temperature effects on photosynthetic activity of two tomato cultivars with different heat susceptibility. *Journal of Plant Physiology*, 162(3),281–289.
- Capiati, D.A., País, S.M. and Téllez-Iñón, M.T., 2006. Wounding increases salt tolerance in tomato plants: evidence on the participation of calmodulin-like activities in crosstolerance signalling. *Journal of Experimental Botany*, 57(10), 2391-2400.
- Chang, P.T. and Randle, W.M., 2004. Sodium chloride in nutrient solutions can affect onion growth and flavor development. *HortScience*, 39(6), 1416-1420.
- Chen, Y.H., Aires, F., Francis, J.A. andMiller, J.R., 2006. Observed relationships between Arctic longwave cloud forcing and cloud parameters using a neural network. *Journal of Climate*, 19, 4087–4104. DOI: 10.1175/JCLI3839.1
- Chen, Z., Zheng, Z., Huang, J., Lai, Z. and Fan, B., 2009b. Biosynthesis of salicylic acid in plants. *Plant Signaling & Behavior*, 4, 493–496
- Chen, D., Shao, Q., Yin, L., Younis, A. and Zheng, B., 2019. Polyamine function in plants: development, and

roles in abiotic stress responses. *Frontiers in Plant Science*, 9, 1945. DOI: 10.3389/fpls.2018.01945

- Childs, C., Holdsworth, R.E.,
 Christopher, A.L., Jackson,
 Manzocchi, T., Walsh, J.J. and
 Yielding, G., 2017. Introduction to
 the geometry and growth of normal faults. *Geological Society*, 439, 1–9.
 DOI: 10.1144/SP 439.24
- Clarke, S.M., Cristescu, S.M., Miersch. 0., Harren, **F.J.M.** Wasternack, C. and Mur, L.A.J., 2009. Jasmonates act with salicylic acid to confer basal thermotolerance in Arabidopsis thaliana. New Phytologist, 182, 175–187. DOI:10.1111/j.1469-8137.2008.02735.x
- Cook, C.E., Whichard, L.P., Wall, M.E., Egley, G.H., Coggon, P., Luhan, P.A. and Mcphail, A.T., 1972. Germination stimulants. II. The structure of strigol-a potent seed germination stimulant for witchweed (Striga lutea Lour.). *Journal of the American Chemical Society*, 94, 6198–6199. DOI: 10.1021/ja00772a048
- Cornforth, J.W., Milborow, B.V., Ryback, G. and Wareing, P.F., 1965. Identity of sycamore 'dormin' with abscisin II. *Nature*, 205, 1269– 1270. DOI: 10.1038/2051269b0
- Cruiziat, P. and Cochard, H., 2002. Hydraulic architecture of trees: main concepts and results. *Annals of Forest Science*, 59, 723–752
- Danilova, M.N., Kudryakova, N.V., Doroshenko, A.S., Zabrodin, D.A., Vinogradov, N.S. and Kuznetsov,

V.V. 2016. Molecular and physiological responses of Arabidopsis thaliana plants deficient in the genes responsible for ABA and cytokinin reception and metabolism shock. Russian to heat Journal of Plant Physiology, 63. 308-318. DOI: 10.1134/S1021443716030043

- Danquah, A., Colcombet, J. and Hirt,
 H., 2013. The role of ABA and
 MAPK signaling pathways in plant
 abiotic stress responses. *Biotechnology Advances*, 32, 6. DOI: 10.1016/j.biotechadv.2013.09.006
- Dar, N.A., Amin, I., Wani,W.,Wani, S.A., Shikari, A.B.,Wani, S.H. and Masoodi, K., 2017. Abscisic acid: a key regulator of abiotic stress tolerance in plants. *Plant Gene*, 11, 106–111. DOI: 10.1016/j.plgene.2017.07.003
- Darwin, C. and Darwin, F., 1880. The power of movement in plants. London: John Murray. DOI: 10.5962/bhl.title.102319
- **De la Pena, R. and Hughes, J., 2007.** Improving Vegetable Productivity in a Variable and Changing Climate. *Journal of SAT Agricultural Research*, 4:1-22.
- Dempsey, W., 1970. Effects of temperature on pollen germination and tube growth. *Report of the Tomato Genetics Cooperative*, 20, 15–16.
- **Dodd, I.C., 2005.** Root-to-shoot signalling: assessing the roles of Bup^ in the up and down world of long-distance signalling in planta. *Plant Soil*, 274, 251–270

- Drew, M.C., 2009. Plant responses to anaerobic conditions in soil and solution culture. *Current Advances in Plant Science*, 36, 1-14.
- **Driedonks, N.J.W., 2018.** From flower to fruit in the heat-Reproductive thermotolerance in tomato and its wild relatives (Doctoral dissertation. [*Sl.* 2018;sn])
- Dubey, S., Kuruwanshi, V. **B..** Ghodke, P.H. and Mahajan, V., 2020. **Biochemical** and vield evaluation of onion (Allium cepa L.) waterlogging genotypes under condition. International Journal of Chemical Studies, 8, 2036-2040.
- Fleet, C.M. and Sun, T.P., 2005. A delicate balance the role of gibberellin in plant morphogenesis. *Current Opinion in Plant Biology*, 8, 77–85
- Folzer, H., Dat, J.F., Capelli, N., Rieffel, D. and Badot, P.M., 2006. Response of sessile oak seedlings (*Quercus petraea*) to flooding: an integrated study. *Tree Physiology*, 26, 759-766.
- Freitas, A.M., Nair, V.D., Harris, W.G., Mosquera-Losada, M.R. and Nair, P.K.R., 2017. Phosphorus Release Behavior of Biosolids and Corresponding Biochars in Abstract Retrieved From ASA, CSSA and SSSA International AnnualMeetings. Available online at: https://scisoc.confex.com/~ crops/2017am/webprogram/Paper10 6996.html (accessed August 20, 2021).

- Ghodke, P.H., Shirsat, D.V., Thangasamy, A., Mahajan, V., Salunkhe, V.N., Khade, Y. and Singh, M., 2018. Effect of water logging stress at specific growth stages in onion crop. International Journal of Current Microbiology and Applied Sciences, 7, 3438-48.
- Gibbs, J., Greenway, H., 2008. Mechanisms of anoxia tolerance in plants. I. Growth, survival and anaerobic catabolism. *Functional Plant Biology*, 30: 1-47.
- Gomez-Roldan, M.V., Fermas, S., Brewer, P.B., Puech-Pagès, V., Dun, E.A., Pillot, J.P., Letisse, F., Matusova, R., Danoun, S., Portais, J.C., Bouwmeester, H.J., Becard, G. Beveridge, C.A., Rameau, C. and Rochange, S.F., 2008. Strigolactone inhibition of shoot branching. *Nature*, 455, 189–194. DOI: 10.1038/nature07271
- Gull, A., Lone, A.A. and Wani, N.U.I., 2019. Biotic and abiotic stresses in plants. *Abiotic and Biotic Stress in Plants*, 1-19.
- Guo, L.L., Hao, L.H., Jia, H.H., Li, F. , Zhang, X.X., Cao, X., Xu, M. and Zheng, Y.P., 2018. Effects of NaCl stress on stomatal traits, leaf gas exchange parameters, and biomass of two tomato cultivars. *Ying Yong Sheng Tai Xue Bao*, 29, 3949– 3958. [PubMed], [Google Scholar]).
- Gupta, N.K., Gupta, S., Shukla, D.S.
 and Deshmukh, P.S., 2003.
 Differential response of BA injection on yield and specifi c grain weight in wheat genotypes recommenced for

normal and late sown conditions. *Plant Growth Regulation*, 40, 201–205.

- Guschina, I.A., Harwood, J.L., Smith, M. and Beckett, R.P., 2002. Abscisic acid modifies the changes in lipid brought about by water stress in the moss (Atrichum androgynum). New Phytologist, 156, 255–264. DOI: 10.1046/j.1469-8137.2002.00517.x
- Ha, C.V., Leyva-Gonzalez, M.A., Osakabe, Y., Tran, U.T., Nishiyama, R., Watanabe, Y., et al. 2014. Positive regulatory role of strigolactone in plant responses to drought and salt stress. Proceedings of the National Academy of Sciences U.S.A., 111, 851–856. DOI: 10.1073/pnas.1322135111.
- Hatfield, J.L., Boote, K.J., Kimball,
 B.A., Ziska, L.H., Izaurralde, R.C.,
 Ort, D.R., Thomson, A.,M. and
 Wolfe, D., 2011. Climate impacts on agriculture: implications for crop production. *Agronomy*

Journal, 103(2), 351-370.

- He, M., He, C.Q., and Ding, N.Z., 2018. Abiotic stresses: general defenses of land plants and chances for engineering multistress tolerance. *Frontiers in Plant Science*, 9, 1771. DOI: 10.3389/fpls.2018.01771
- Heinrich, M., Hettenhausen, C., Lange, T., Wunsche, H., Fang, J.,
 Baldwin, I.T. and Wu, J., 2013.
 High levels of jasmonic acid antagonize the biosynthesis of gibberellins and inhibit the growth of Nicotiana attenuata stems. *Plant Journal*, 73, 591–606.

- Hijmans, R.J., 2003. The effect of climate change on global potato production. American Journal of Potato Research, 80:271–280
- Hirabayashi, Y., Mahendran, R.,
 Koirala, S., Konoshima, L.,
 Yamazaki, D., Watanabe, S., Kim,
 H. and Kanae, S., 2013. Global flood
 risk under climate change. *Nature Climate Change*, 3(9), 816-821.
- Horváth, E., Pál, M., Szalai, G., Páldi,
 E. and Janda, T., 2007. Exogenous
 4- hydroxybenzoic acid and salicylic acid modulate the effect of short-term drought and freezing stress on wheat plants. *Biologia plantarum*, 51, 480–487. DOI: 10.1007/s10535-007-0101-1
- Hossain, A., Raza, A. and Maitra, S., Asaduzzaman M., Islam, M.R., Hossain, M.J., Sabagh, E.L., Garai, S., Mondal, M., Abdel-Latif, A.H. and Aftab, T., 2021. "Strigolactones: carotenoid-derived а novel biosynthesis, phytohormone transporters, signalling, and mechanisms in abiotic stress," in Plant Growth Regulators, eds T. Aftab, and K.R. Hakeem (Cham: Springer), 275-303. DOI: 10.1007/978-3-030-61153-8 13
- Igarashi, K. and Kashiwagi, K., 2010. Modulation of cellular function by polyamines. *International Journal of Biochemistry Cell Biology*, 42, 39– 51. DOI:
 - 10.1016/j.biocel.2009.07.009
- Iqbal, M., Basra, S.M.A. and Jamil, A., 2006. Seed enhancement with cytokinins: changes in growth and grain yield in salt stressed wheat

plants. *Plant Growth Regulation*, 50, 29–39. DOI: 10.1007/s10725-006-9123-5

- Iqbal, M.A., 2015a. Improving the growth and yield of canola (Brassica napus L.) with seed treatment and foliar sprays of brassica (*Brassica* naups L.) and moringa (*Moringa* oleifera L.) leaf extracts. American-Eurasian Journal of Agricultural & Environmental Scien ces, 14, 1067–1073. DOI: 10.5829/idosi.aejaes.2014.14.10.124 29
- Islam, M.S., Hasan, M.K., Islam, B., Renu, N.A., Hakim, M.A., Islam, M.R., Chowdhury, M.K., Ueda, A., Saneoka, H., Reza, M.A., Fahad, S., barutçular, C., Çığ, F., Erman, M., Sabagh, E.L., 2021. Responses of water pigments status, drv and matter partitioning, seed production, and traits of yield and quality to foliar application of GA3 in mugnbean (Vigna radiata L.). Frontiers in Agronomy, 2, 596850. DOI: 10.3389/fagro.2020.596850
- Joseph, T.A., Krishna, K.S., Kumar, P.D., Rajkumar J., Aipe, K.C., Pradeep, Kumar, T. and Minhas, J.S., 2006. Evaluation of heat tolerant hybrids under tropical conditions in Kerala. *Potato Jouranl*, 33 (3-4), 151-153
- Jouyban, Z., Hasanzade, R. and Sharafi, S., 2013. Chilling stress in plants. International Journal of Agriculture and Crop Sciences (IJACS), 5, 2961–2968.
- Jurado, S., Abraham, Z., Manzano, C., López-Torrejón, G., Pacios, L. F. and Del Pozo, J.C., 2010. The

Arabidopsis cell cycle F-box protein SKP2A binds to auxin. *Plant Cell*, 22, 3891–3904. DOI:

10.1105/tpc.110.078972

- Kadayifci, A., Tuylu, G.İ., Ucar, Y. and Cakmak, B., 2005. Crop water use of onion (*Allium cepa* L.) in Turkey. *Agricultural Water Management*, 72(1), 59-68.
- Kakani, V.G., Reddy, K.R., Koti, S.,
 Wallace, T.P., Prasad, P.V.V.,
 Reddy, V.R. and Zhao, D., 2005.
 Differences in in vitro pollen germination and pollen tube growth of cotton cultivars in response to high temperature. *Annals of Botany*, 96(1),59–67.
- Kapulnik, Y., Resnick, N., Mayzlish-Gati, E., Kaplan, Y., Wininger, S., Hershenhorn, J., Koltai, H., 2011.
 Strigolactones interact with ethylene and auxin in regulating root-hair elongation in Arabidopsis. *Journal of Experimental Botany*, 62, 2915–2924. DOI: 10.1093/jxb/erq464
- Kaymakanova, M., Stoeva, N. and Mincheva, T., 2008. Salinity and its effects on the physiological response of bean (*Phaseolus vulgaris L.*). *Journal of Central European Agriculture*, 9: 749-756.
- Kendrick, M.D. and Chang, C., 2008.
 Ethylene signaling: new levels of complexity and regulation. *Current Opinion in Plant Biology*, 11, 479–485. DOI: 10.1016/j.pbi.2008.06.011
- Ketabchi, S. and Shahrtash, M., 2011. Effects of methyl jasmonate and cytokinin on biochemical responses of maize seedlings infected by

Fusarium moniliforme . *South Asian Journal of Experimental Biology*, 2, 299–305

- Khan, S., Rahman, J., Khan, N., Hussian, I., Izharullah and Khan,
 F., 2017. Morphological variation among tomato germplasams. *International Journal* of Agriculture and Environmental Research, 3(2), 240-244.
- Khokhar, K.M., Hadley, P. and Pearson, S., 2007. Effect of cold temperature durations of onion sets in store on the incidence of bolting, bulbing and seed yield. *Scientia Horticulturae*, 112(1), 16-22.
- Kim, T.W., Hwang, J.Y., Kim, Y.S., Joo, S.H., Chang, S.C., Lee, J.S., Takatsuto, S. F. Kim, S.K., 2005. CYP85A2, Arabidopsis a cytochrome P450, mediates the baever-villiger oxidation of brassinolide castasterone to in brassinosteroid biosynthesis. Plant Cell, 17, 2397-2412
- Kolaksazov, M., Laporte, F.,
 Ananieva, K., Dobrev, P., Herzog,
 M. F. Ananiev, E., 2013. Effect of chilling and freezing stresses on jasmonate content in arabis alpina.
 Bulgarian Journal of Agricultural Science, 19, 15–17
- Kosova, K., Prasil, I.T., Vitamvas, P.,
 Dobrev, P., Motyka, V., Flokova,
 K., Novak, O., Tureckova, V.,
 Rolcik, J. and Pesek, B., 2012.
 Complex phytohormone responses
 during the cold acclimation of two
 wheat cultivars differing in cold
 tolerance, winter Samanta and spring

Sandra. *Journal of Plant Physiology*, 169, :567–576

- Koyro. H.W., Ahmad, P. and Geissler,
 N., 2012. Abiotic stress responses in plants: an overview. In: Ahmad P,
 Prasad MNV (eds) Environmental adaptations and stress tolerance of plants in the era of climate change.
 Springer, New York, pp. 1–28
- Kumar S., Kumar, D. and Minhas, J.S., 2005. Varietal differences in response of potatoes to repeated periods of water stress in winter crop. *Potato Journal*, 32 (3-4),197-198
- Kumar, D., Chalkoo, S., Singh, B.P. and Minhas, J.S., 2007. Heat tolerant variety Kufri Surya has higher antioxidant content in tubers exposed to high temperature. *Potato Journal*, 34 (1-2), 63-64
- Kumar, B., Mistry, N.C., Chander,B.S. and Gandhi, P., 2011. Indian horticulture production at a glance. Indian horticulture database 2011. National horticulture Board, Ministry of Agriculture, Government of India.
- Kumar D. and. Minhas, J.S., 2013.
 Evaluation of indigenous potato varieties, advanced clones and exotic genotypes against water deficit stress under sub-tropical environment. *Indian Journal Plant Physiology*, 18(3), 240-249
- Kumar, D. and Minhas, J.S., 2015.
 Can water deficit be useful in potato?
 Some issues. In Recent Advances in Crop Physiology (Singh AL Ed.)
 Daya Publishing House A Division of Astral International Pvt. Ltd. New Delhi, Vol 2, pp. 225-236.

- Kumar, S.N., 2017. Climate change and its impacts on food and nutritional security in India. In: Belavadi VV, Karaba NN, Gangadharappa NR (eds) Agriculture under Climate Change: Threats, Strategies and Policies, 48 P.
- Kumar, S., Kumar, P., Kumar, D. and. Malik, P.S., 2017. Effect of Water Stress on Haulm Yield, Total Biomass and Harvest Index of Potato Cultivars. *Plant Archives*, 17(1), 623-626
- Kusvuran, S., Kiran, S., Ellialtioglu,
 S.S., 2016. Antioxidant enzyme activities and abiotic stress tolerance relationship in vegetable crops. In: Abiotic and Biotic Stress in Plants Recent Advances and Future Perspectives, InTech.
- Levy, D. and Veilleux, R.E., 2007. Adaptation of potato to high temperatures and salinity—a review. *American Journal of Potato Research*, 84, 487–506
- Li, Z., Palmer, W.M., Martin, A.P., Wang, R., Rainsford, F., Jin, Y. and Ruan, Y.L., 2014. High invertase activity in tomato reproductive organs correlates with enhanced sucrose import into, and heat tolerance of, young fruit. *Journal of Experimental Botany*, 63(3),1155–1166.
- Li, L., Xu, J., Xu, Z.H. and Xue, H.W.,
 2005. Brassinosteroids stimulate plant tropisms through modulation of polar auxin transport in Brassica and Arabidopsis. *Plant Cell*, 17, 2738–2753. DOI: 10.1105/tpc.105.034397

- Liu, H. Y., Yu, X., Cui, D. Y., Sun, M.H., Sun, W.N., Tang, Z.C., Kwak, S.S. and Sun, W., 2007. The role of water channel proteins and nitric oxide signaling in rice seed germination. *Cell Reseach*, 17, 638– 649. DOI: 10.1038/cr.2007.34
- Liu Z.F., Fu B.J. and Zheng, X.X., 2010. Plant biomass, soil water content and soil N:P ratio regulating soil microbial functional diversity in a temperate steppe: A regional scale study. *Soil Biology and Biochemistry*, 42, 445-450.
- Liu, Q., Yan, S., Huang, W., Yang, J.Y., Dong, J., Zhang, S., Zhao, J., Yang, T., Mao, X., Zhu, X. and Liu,
 B., 2018. NAC transcription factor ONAC066 positively regulates disease resistance by suppressing the ABA signaling pathway in rice. *Plant Molecular Biology*, 98, 289–302. DOI: 10.1007/s11103-018-0768-z
- Lo, S.F., Ho, T.H.D., Liu, Y.L., Jiang, M.J., Hsieh, K.T., Chen, K.T., Yu, L.C., Lee, M.H., Chen, C.Y., Huang, T.P., Koima. М., Sakakibara, H., Chen, L.J. and Yu, S.M., 2017. Ectopic expression of specific GA 2 oxidase mutants promotes yield and stress tolerance in rice. Plant Biotechnology Journal, 850-864. 15, DOI: 10.1111/pbi.12681
- Lopez, M.A.H., Ulery, A.L., Samani,
 Z., Picchioni, G. and Flynn, R.P.,
 2011. Response of chile pepper (capsicum annuum L.) to salt stress and organic and inorganic nitrogen sources: i. growth and yield. *Tropical*

and Subtropical Agroecosystems, 14: 137-147.

- Luoh, J.W., Begg, C.B., Symonds, R.C., Ledesma, D. and Yang, R.Y., 2014. Nutritional yield of African indigenous vegetables in waterdeficient and water-sufficient conditions. *Food and Nutrition Sciences*, 05, 812–822. https://doi.org/10.4236/ fns.2014.59091.
- Machado, R.M.A. and Serralheiro, R.P., 2017. Soil Salinity: Effect on vegetable crop growth. Management practices to prevent and mitigate soil salinization. *Horticulturae*, 3, 1–13.
- Mahouachi, J., Arbona, V. and Gómez-Cadenas, A., 2007. Hormonal changes in papaya seedlings subjected to prograssive water stress in this halophyte. *Plant Growth Regulation*, 53, 43–51. DOI: 10.1007/s10725-007-9202-2
- Marin, E., Jouannet, V., Herz, A., Lokerse, A.S., Weijers, D., Н., Nussaume, Vaucheret, L., Crespi, M.D. and Maizel, A., 2010. miR390, arabidopsis TAS3 tasiRNAs, and their AUXIN **RESPONSE FACTOR targets define** autoregulatory network an quantitatively regulating lateral root growth. Plant Cell, 22, 1104-1117. DOI: 10.1105/tpc.109.072553
- Mark, C., Zór, K., Heiskanen, A., Dufva, M., Emnéus, J. and Finnie, C., 2016. Monitoring intra-and extracellular redox capacity of intact barley aleurone layers responding to phytohormones. *Analytical*

Biochemistry, 515, 1–8. DOI: 10.1016/j.ab.2016.09.011

- Marzec, M., Muszynska, A. and Gruszka, D., 2013. The role of strigolactones in nutrient-stress responses in plants. *International Journal of Molecular*, 14, 9286– 9304. DOI: 10.3390/ijms14059286
- Marzec, M., 2016. Perception and signaling of strigolactones. *Frontiers in Plant Science*, 7, 1260. DOI: 10.3389/fpls.2016.01260
- Mathur, S., Allakhverdiev, S.I. and Jajoo, A., 2011. Analysis of high temperature stress on the dynamics of antenna size and reducing side heterogeneity of Photosystem II in wheat leaves (Triticum aestivum). *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 1807(1), 22-29.
- Mattoo, A.K. and Suttle, C.S. 1991. The plant hormone ethylene. CRS Press, Boca Raton, FL Belimov AA, Safranova VI, Mimura T (2002) Response of spring rape (Brassica napus) to inoculation with PGPR containing ACC-deaminase depends on nutrient status of plant. *Canadian Journal of Microbiology*, 48, 189– 199
- Maurel, C., Simonneau, T. and Sutka, M., 2010. The significance of roots as hydraulic

rheostats. *Journal of Experimental Botany*, 61, 3191–3198

Mellidou, I., Karamanoli, K., Beris, D.and Al, E., 2017. Underexpression ofapoplasticpolyamineoxidaseimprovesthermotoleranceinNicotiana tabacum. Journal of Plant

Physiology, 218, 171–174. DOI: 10.1016/j.jplph.2017.08.006

- Minhas, J.S., Kumar, P., Kumar, D.,
 Gupta, Y.K. and Dua, V.K., 2018.
 Response of potato to elevated CO2 under short days: Growth, physiological parameters and tuber yield. *Indian Journal of Horticulture*, 75(1),82-86.
- Minocha, R., Majumdar, R. and Minocha, S.C., 2014. Polyamines and abiotic stress in plants: a complex relationship. *Frontiers in Plant Science*, 5, 175. DOI: 10.3389/fpls.2014.00175.
- Misra, N. and Saxena, P., 2009. Effect of salicylic acid on proline metabolism in lentil grown under salinity stress. *Plant Science*, 177, 181–189.
- Munns, R. and Tester, M., 2008. Mechanisms of salinity tolerance. *Annual review of Plant Biology*, 59, 651.
- Nadeem, S.M., Zahir, Z.A., Naveed,
 M., Asghar, H.N. and Arshad, M.,
 2010a Rhizobacteria capable of producing ACC– deaminase may mitigate salt stress in wheat. Soil Science Society of America Journal, 74, 533–542.
- Nadeem, S.M., Zahir, Z.A., Naveed, M. and Ashraf, M., 2010b. Microbial ACC-deaminase: prospects and applications for inducing salt tolerance in plants. *Critical* Reviews *in Plant Sciences*, 29, 360–393.
- Nankishore, A. and Farrell, A.D.,2016. The response of contrasting tomato genotypes to combined heat

and drought stress. *Journal of Plant Physiology*, 202, 75–82.

- Nemeskéri, E. and Helyes, L., 2019. Physiological responses of selected vegetable crop species to water stress. *Agronomy*, 9(8), 447.
- Newton, A.C., Johnson, S.N. and Gregory, P.J., 2011. Implications of climate change for diseases, crop yields and food security. *Euphytica*, 179, 3-18.
- Nilsen Erik T. and Orcutt, D.M., 1996. *Physiology of plants stress* (abiotic factors). John Wiley and Sons.Inc. New York, 696P.
- Nolan, T., Vukasinovi, N., Liu, D., Russinova, E. and Yina, Y., 2020. Brassinosteroids: multidimensional regulators of plant growth, development, and stress responses. *Plant Cell*, 32, 295–318. DOI: 10.1105/tpc.19.00335.
- Pál, M., Szalai, G. and Janda, T., 2015.
 Speculation: polyamines are important in abiotic stress signaling. *Plant Science*, 237, 16–23. DOI: 10.1016/j.plantsci.2015.05.003.
- Parasher, A. and Varma, S.K., 1988. Effect of pre-sowing seed soaking in gibberellic acid on growth of wheat (Triticum aestivum L.) under different saline conditions. *Indian Journal of Biological* Science, 26, 473–475.
- Parent, C., Capelli, N., Berger, A., Crèvecoeur, M. and Dat, J.F., 2008. An overview of plant responses to soil waterlogging. *Plant Stress*, 2, 20-27.
- Parihar, P., Singh, S., Singh, R., Vijay Pratap Singh, V.P. and Prasad,

S.M., 2015. Effect of salinity stress on plants and its tolerance strategies: A review. *Environmental Science and Pollution Research*, 22, 4056–4075.

- Pautasso, M., Doring, T.F., Garbelotto, M., Pellis, L. and Jeger, M.J., 2012. Impacts of climate change on plant diseases-opinions and trends. *European Journal of Plant Pathology*, 133, 295-313.
- Peleg, Z. and Blumwald, E., 2011. Hormone balance and abiotic stress tolerance in crop plants. *Current Opinion in Plant Biology*, 14, 290– 295. DOI: 10.1016/j.pbi.2011.02.001
- Penmetsa, R.V. and Cook, D.R., 1997. A legume ethylene-insensitive mutant hyperinfected by its rhizobial symbiont. *Science*, 275, 527–530
- Petretto, G.L., Urgeghe, P.P., Massa, D. and Melito, A., 2019. Effect of salinity (NaCl) on plant growth, nutrient content, and glucosinolate hydrolysis products trends in rocket genotypes. *Plant*

Physiology and Biochemistry, 141, 30–39.

- Pospíšilová, J., 2003. Participation of phytohormones in the stomatal regulation of gas exchange during water stress. *Biologia Plantarum*, 46(4), 491-506.
- Prasanna, Y.L. and Ramarao, G., 2014. Effect of waterlogging on physiological and biochemical parameters and seed yield in greengram genotypes. *International Journal of Food, Agriculture and Veterinary Sciences*, 4(2), 176-183.

- Pressman, E., Peet, M.M. and Pharr, D.M., 2002. The effect of heat stress on tomato pollen characteristics is associated with changes in carbohydrate concentration in the developing anthers. *Annals of Botany*, 90(5),631–636.
- Raja, M.M., Vijayalakshmi, G., Naik,
 M.L., Basha, P.O., Sergeant, K.,
 Hausman, J.F. and Khan, P.S.S.V.,
 2019. Pollen development and function under heat stress: from effects to responses. *Acta Physiologiae Plantarum*, 41(4),47.
- Rao, S.S.R., Vardhini, B.V., Sujatha,
 E., Anuradha, S., 2002.
 Brassionsteroids-a new class of phytohormones. *Current Science*, 82, 1239–1245.
- Rao, N.S., Shivashankara, K.S. and Laxman, R.H., (Eds.) 2016. *Abiotic* stress physiology of horticultural crops (Vol. 311). India: Springer.
- Raza, M.A., van der Werf, W. and Ahmed M., Yang, W., 2020. Removing top leaves increases yield and nutrient uptake in maize plants. *Nutrient Cycling in Agroecosystems*, 118, 57–73. DOI: 10.1007/s10705-020-10082-w
- **Ribaut, J.M. and Pilet, P.E., 1991.** Effect of water stress on growth, osmotic potential and abscisic acid content of maize roots. *Physiologia Plantarum*, 81, 156–162.
- Rivero, R.M., Kojima, M., Gepstein,
 A., Sakakibara, H., Mittler, R.,
 Gepstein, S. and Blumwald, E.,
 2007. Delayed leaf senescence induces extreme drought tolerance in a fl owering plant. *Proceedings of the*

National Academy of Sciences USA, 104, 19631–19636

- Rivero, R.M., Gimeno, J., Van Devnze, A., Walia, H. and Blumwald, E., 2010. Enhanced cytokinin synthesis in tobacco plants expressing PSARK:IPT prevents the degradation of photosynthetic protein complexes during drought. Plant Cell Physiology, 51, 1929–1941. DOI: 10.1093/pcp/pcq143
- Rochange, **S.**. Goormachtig, S.. Lopez-Raez, J.A. and Gutjahr, C., 2019. "The role of strigolactones in plant-microbe interactions," in Strigolactones-Biology and Applications, eds H. Koltai and C. Prandi (Cham: Springer). pp. 121-142. DOI: 10.1007/978-3-030-12153-2 4
- Rouphael, Y., Raimondi, C., Lucini, L., Carillo, P., Kyriacou, M.C., Col la, G., Cirillo, V., Pannico, A., El-Nakhel, C. and De-Pascale, S., 2018. Physiological and metabolic responses triggered by omeprazole improve tomato plant tolerance to NaCl stress. *Frontiers in Plant Science*, DOI: 10.3389/fpls.2018.00249
- Sachs, T., 2005. Auxins role as an example of the mechanisms of shoot/root relations. *Plant Soil*, 268, 13–19. DOI: 10.1007/s11104-004-0173-z
- Sakakibara, H., 2006. Cytokinins: Activity, biosynthesis, and translocation. *Annual Review of Plant Biology*, 57, 431–49. DOI:

10.1146/annurev.arplant.57.032905. 105231

- Salama, F.M. and Awadalla, A.A., 1987. The effects of different kinetin application methods on some chlorophyll parameters of two crop plants grown under salinity stress. *Phyton*, 21, 181–193
- Sariyev, A., Barutcular, C., Acar, M., Hossain, A.E.L. and Sabagh, A.,
 2020. Sub-surface drip irrigation in associated with H2O2 improved the productivity of maize under clay-rich soil of Adana, Turkey. *International Journal of Experimental Botany*, 89, 519–528. DOI: 10.32604/phyton.2020.09142
- Sarker, U., Islam, M.T. and Oba, S., 2018. Salinity stress accelerates nutrients, dietary fiber, minerals, phytochemicals and antioxidant activity in Amaranthus tricolor leaves. PLoS 13. One. https://doi.org/10.1371/journal.pone. 0206388. e0206388-e0206388.
- Savada,R.P.,Ozga,J.A.,Jayasinghege,C.P.A.,Waduthanthri, K.D. and Reinecke,D.M., 2017. Heat stress differentiallymodifies ethylene biosynthesis andsignaling in pea floral and fruittissues. PlantMolecularBiology, 95, 313–331.

DOI:10.1007/s11103-017-0653-1

Scheelbeek, P.F.D., Bird, F.A.,
Tuomisto, H.L., Green, R., Harris,
F.B., Joy, E.J.M., Chalabi, Z.,
Allen, E., Haines, A. and Dangour,
A.D., 2018. Effect of environmental changes on vegetable and legume yields and nutritional quality.

Proceedings of the National Academy of Sciences U.S.A., 115, 6804–6809.

https://doi.org/10.1073/pnas.180044 2115

- Schumacher, K. and Chory, J., 2000. Brassinosteroid signal transduction: still casting the actors. *Current Opinion in Plant Biology*, 3, 79–84
- Semida, W.M., 2016. Hydrogen peroxide alleviates salt-stress in two onion (Allium cepa L.) cultivars. *Management*, 2, 2.
- Shani, E., Weinstain, R., Zhanga, Y., Castillejo, C., Kaiserli, E., Chory, J., Tsienb, R.Y. and Estelle, M., 2013. Gibberellins accumulate in the elongating endodermal cells of Arabidopsis root. *PNAS*, 110, 4834– 4839
- Sharif, R., Xie, C., Zhang, H., Arnao, M. B., Ali, M., Ali, Q., Muhammad,
 I., Shalmani, A., Nawaz, M.A.,
 Chen P. And Li, Y., 2018. Melatonin and its effects on plant systems. *Molecules*, 23, 2352. DOI: 10.3390/molecules23092352
- Shibli, R.A., Kushad, M., Yousef, G.G. and Lina, M.A., 2007. Physiological and biochemical responses of tomato microshoots to induced salinity stress with associated ethylene accumulation. *Plant Growth Regulation*, 51, 159– 169.
- Shirani Bidabadi, S. and Mehralian, M., 2020. Arbuscular mycorrhizal fungi inoculation to enhance chilling stress tolerance of watermelon. *Gesunde Pflanzen*, 72, 171–179.

https://doi.org/10.1007/s10343-020-00499-2.

- Singh, A., Siddappa, S., Bhardwaj, V.,
 Singh, B., Kumar, D. and Singh,
 B.P., 2015. Expression profiling of potato cultivars with contrasting tuberization at elevated temperature using microarray analysis. *Plant Physiology and Biochemistry*, 97, 108-116
- Sta-Baba, R., Hachicha, M., Mansour, M., Nahdi, H. and Kheder, M.B.,
 2010. Response of onion to salinity. African Journal of Plant Science and Biotechnology, 4(2), 7-12.
- Stirk, W.A., Bálint, P., Tarkowská, D., Novák, O., Strnad, M., Ördög, V. and van Staden, J., 2013. Hormone profi les in microalgae: gibberellins and brassinosteroids. *Plant Physiology and Biochemistry*, 70, 348–353
- Su, Y.i, Xia, S., Wang, R. and Xiao, L.,
 2017. Phytohormonal quantification based on biological principles. *Hormone Metabolic Signaling in Plants*, 431–470. DOI: 10.1016/B978-0-12-811562-6.00013-X
- Sun, Q., Zhang, N., Wang, J., Zhang,
 H., Li, D., Shi, J., Li, R., Weeda, S.,
 Zhao, B. Ren, S. and Guo, Y.D.,
 2014. Melatonin promotes ripening and improves quality of tomato fruit during postharvest life. *Journal of Experimental Botany*, 66,
 657–668. DOI: 10.1093/jxb/eru332
- Sun, Y., Cui, X. and Liu, F., 2015. Effect of irrigation regimes and phosphorus rates on water and

phosphorus use efficiencies in potato. *Scientia Horticulturae*, 190, 64–69.

- Suzuki, N., Miller, G., Sejima, H., Harper, J. and Mittler, R., 2013. Enhanced seed production under prolonged heat stress conditions in Arabidopsis thaliana plants deficient in cytosolic ascorbate peroxidase 2. Journal of Experimental Botany, 64(1), 253-263.
- Sykorova. Kurešova, **B.**, G., Daskalova, S., Trckova, M., K., Raimanova, Hoverova, I., Motyka, V., Travnickova, A., Elliott, M.C. and Kaminek, M., 2008. Senescence-induced ectopic expression of the A. tumefaciens ipt gene in wheat delays leaf senescence, increases cytokinin content, nitrate infl ux, and nitrate reductase activity, but does not affect grain yield. Journal of Experimental Botany, 59. 377-387.
- Symons, G.M., Ross, J.J., Jager, C.E. and Reid, J.B., 2008. Brassinosteroid transport. *Journal of Experimental*, 59, 17–24. DOI: 10.1093/jxb/erm098
- Takahashi, D., Li, B., Nakayama, T.,
 Kawamura, Y. and Uemura, M.,
 2013. Plant plasma membrane proteomics for improving cold tolerance. *Frontiers in Plant Science*, 4, 90.
- Takeuchi, K., Gyohda, A., Tominaga,
 M., Kawakatsu, M., Hatakeyama,
 A., Ishii, N., Shimaya, K.,
 Nishimura, T., Riemann, M., Nick,
 P., Hoshimoto, M., Komano, T.,
 Endo, A., Okamoto, T., Jikumaru,
 Y., Kamiya, Y., Terakawa, T. and

Koshiba, T., 2011. RSOsPR10 expression in response to environmental stresses is regulated antagonistically by jasmonate/ethylene and salicylic acid signaling pathways in rice roots. *Plant Cell Physiology*, 52, 1686– 1696. DOI: 10.1093/pcp/pcr105

- Tirado, M.C., Clarke, R., Jaykus, L.A., McQuatters Gollop, A. and Frank, J.M., 2010. Climate change and food safety: A review. *Food Research International*, 43, 1745-1765.
- Tommasini, L., Svensson, J.T., Rodriguez, E.M., Wahid, A., Malatrasi, M., Kato, K., Wanamaker, S., Resnik, J. and Close, T.J., 2008. Dehydrin gene expression provides an indicator of low temperature and drought stress: transcriptome-based analysis of barley (Hordeum vulgare L.). Functional and *Integrative* Genomics, 8(4), 387-405.
- Torres-Garcia, J.R., Estradaa, J.A.E. and Gonzólez, M.T.R., 2009. Exogenous application of growth regulators in snap bean under water and salinity stress. *Journal of Stress Physiology & Biochemistry*, 5, 13– 21.
- Tromas, A., Braun, N., Muller, P., Khodus, T., Paponov, I.A., Plame, K., Ljun, K., Lee, J.Y., Benfey, P.
 Murray, J.A.H., Scheres, B. and Perrot-Rechemnamm, C., 2009. The auxin binding protein 1 is required for differential auxin responses mediating root growth.

PLoS ONE, 4, e6648. DOI: 10.1371/journal.pone.0006648

- Tuteja, N., 2007. Mechanisms of high salinity tolerance in plants. In: Methods Enzymol., Elsevier, pp. 419–438.
- Tuteja, N., Gill, S.S., Tiburcio, A.F.andTuteja,R.(Eds.),2012. Improving crop resistance to
abiotic stress. John Wiley and Sons.
- Vasil, I.K., 1987. Physiology and culture of pollen. *International Review of Cytology*, 107, 127–174.
- Venkateswarlu, B. and Shanker, A.K., 2012. Dryland agriculture bringing resilience to crop production under changing climate. In: Crop stress and its management: Perspectives and strategies. Springer, Netherlands, pp. 19-44.
- Vriet, C., Russinova, E. and Reuzeau, C., 2013. Fromsqualene to brassinolide: the steroid metabolic and signaling pathways across the plant kingdom. *Molecular Plant*, 6, 1738–1757. DOI: 10.1093/mp/sst096
- Wang, W., Vinocur, B. and Altman, A., 2003. "Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance." *Planta*, 218, 1-14.
- Wang, W., Chen, Q., Hussain, S., Mei,
 J., Dong, H., Peng, S., Huang, J.,
 Cui, K. and Nie, L., 2016. Presowing seed treatments in direct-seeded early rice: consequences for emergence, seedling growth and associated metabolic events under chilling stress. Scientific reports, 6(1), 1-10.

- Wang, Y., Reiter, R. J. and Chan, Z., 2017. Phytomelatonin: a universal abiotic stress regulator Journal of Experimenta, 1 69, 963– 974. DOI: 10.1093/jxb/erx473
- Ward, A.W., 2016. Encyclopedia britannica.
- Wasternack, C., 2007. Jasmonates: An Update on Biosynthesis, Signal Transduction and Action in Plant Stress Response, Growth and Development. *Annals of Botany*, 100, 681–697. DOI: 10.1093/aob/mcm079
- Wasilewska, A., Vlad, F., Sirichandra, C., Redko, Y., Jammes, F., Valon,
 C., Frey, N.F.D. and Leung, J.,
 2008. An update on abscisic acid signaling in plants and more. *Molecular Plant*, 1, 198–217.
- Wasternack, C. and Hause, B., 2013. Jasmonates: biosynthesis, perception, signal transduction and action in plant stress response, growth and development. An update to the 2007 review in Annals of Botany. *Annals of Botany*, 111, 1021–1058.
- Wasternack, C., 2014. Action of jasmonates in plant stress responses and development-applied aspects. *Biotechnology Advances*, 32, 31–39. DOI:

10.1016/j.biotechadv.2013.09.009

- Williams, M.E., 2011. Introduction to phytohormones. Teaching tools in plant biology: lecture notes. *Plant Cell*, 22:1. DOI: 10.1105/tpc.118.tt0718
- Wilmoth, J.C., Wang, S., Tiwari, S.B., Joshi, A.D., Hagen, G., Guilfoyle, T.J., Alonsa, J.M., Ecker, J.R. and Reed, J.W., 2005. NPH4/ARF7 and

ARF19 promote leaf expansion and auxin-induced lateral root formation. *Plant Journal*, 43, 118–130. DOI: 10.1111/j.1365-313X.2005.02432.x

- Wishart, J., George, T.S., Brown,
 L.K., Ramsay, G., Bradshaw, J.E.,
 White, P.J. and Gregory, P.J.,
 2013. Measuring variation in potato roots in both field and glasshouse: the search for useful yield predictors and a simple screen for root traits. *Plant Soil*, 368, 231–249.
- Wishart, J., George, T.S., Brown, L.K., White, P.J., Ramsay, G., Jones, H. and Gregory, P.J., 2014. Field phenotyping of potato to assess root and shoot characteristics associated with drought tolerance. *Plant Soil*, 378:351–363.
- Wollenweber, B., Porter, J.R. and Schellberg, J., 2003. Lack of interaction between extreme hightemperature events at vegetative and reproductive growth stages in wheat. *Journal of Agronomy and Crop Science*, 189(3), 142-150.
- Wu, Y.S. and Yang, C.Y., 2019. Ethylene-mediated signaling confers thermotolerance and regulates transcript levels of heat shock factors in rice seedlings under heat stress. *Plant Journal*, 60, 23. DOI:10.1186/s40529-019-0272-z
- Xu. J., Wolters-Arts. M., Mariani. C.,
 Huber. H. and Rieu. I., 2017. Heat stress affects vegetative and reproductive performance and trait correlations in tomato (*Solanum lycopersicum*). *Euphytica*, 213(7),15
 6.

- Yadav, S.K., 2010. Cold stress tolerance mechanisms in plants. A review. Agronomy for Sustainable Development, 30(3), 515-527.
- Yadav, B., Jogawat, A., Gnanasekaran, P., Kumari, P., Lakra, N., Lal, S.K., Pawar, J.S. and Narayan, O.P., 2021. An overview of recent advancement in phytohormones-mediated stress management and drought tolerance in crop plants. *Plant Gene*, 25, 100264. DOI: 10.1016/j.plgene.2020.100264
- Yan, F., Sun, Y.Q., Song, F.B. and Liu, F.L., 2012. Differential responses of stomatal morphology to partial rootzone drying and deficit irrigation in potato leaves under varied nitrogen rates. *Scientia Horticulturae*, 145, 76–83.
- Yang, X. and Lu, C., 2005. Photosynthesis is improved by exogenous glycinebetaine in saltstressed maize plants. *Physiologia Plantarum*, 124, 343–352.
- Yin, Y., Jiang, X., Ren, M., Xue, M., Nan, D., Wang, Z., Xing, Y. and Wang, M., 2018. AmDREB2C, from Ammopiptanthus mongolicus, enhances abiotic stress tolerance and regulates fatty acid composition in transgenic Arabidopsis. *Plant Physiology and Biochemistry*, 130, 517–528. DOI: 10.1016/j.plaphy.2018.08.002
- Yuan, L., Liu, S., Zhu, S., Chen, G.,Liu, F., Zou, M. and Wang, C.,2016. Comparative response of two

wucai (Brassica campestris *L*.) genotypes to heat stress on antioxidative system and cell ultrastructure in root. Acta **Physiologiae** Plantarum, 38. https://doi.org/10.1007/s11738-016-2246-z.

- Yuhashi, K.I., Ichikawa, N., Ezuura, H., Akao, S., Minakawa, Y., Nukui, T. and Minamisawa, K., 2000. Rhizobitoxine production by Bradyrhizobium elkanii enhances nodulation and competitiveness on Macroptilium atropurpureum. Applied and Environmental Microbiology, 66, 2658–2663
- Yusuf. R.O., 2012. Coping with Environmentally Induced Change in Tomato Production in Rural Settlement of Zuru Local Government Area of Kebbi State. *Environmental Issues*, 5(1), 47-54.
- Zayton, A.M., 2007. Effect of soilwater stress on onion yield and quality in sandy soil. *Misr Journal of Agricultural Engineering*, 24(1), 141-160.
- Zhou, L., Tian, S. and Qin, G., 2019. RNA methylomes reveal the m6Amediated regulation of DNA demethylase gene SIDML2 in tomato fruit ripening. *Genome Biology*, 20(1), 1-23.
- Zhu, Z., 2014. Molecular basis for jasmonate and ethylene signal interactions
 in Arabidopsis. Journal of Experime ntal Botany, 65, 5743–5748.