

Review Article:**Advances in Probiotic-Assisted Nutritional Strategies for Sustainable Aquaculture: Mechanisms, Applications, and Future Perspectives****Faseeha I.¹; khaliq A.¹; Laraib D.²; Bhatti A.¹; Naveed U.³**

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Abstract

Aquaculture has become one of the fastest-growing food-producing sectors worldwide and plays a critical role in ensuring global food security by supplying high-quality animal protein. As production intensifies, sustainable nutritional strategies that enhance animal health while reducing environmental impacts have become increasingly important. Among these strategies, probiotics have emerged as promising functional feed additives due to their ability to improve growth performance, feed utilization, gut health, immune competence, and disease resistance in cultured aquatic organisms. Probiotics promote a balanced intestinal microbiota through competitive exclusion of pathogenic microorganisms, production of antimicrobial compounds, and stimulation of digestive enzyme activity, thereby enhancing nutrient digestion and absorption. Their efficacy is influenced by multiple factors, including microbial strain, host species, physiological status, dietary formulation, and farming conditions. In addition to improving growth and survival, selected probiotic strains have demonstrated beneficial effects on reproductive performance, including enhanced fecundity, gamete quality, hatchability, and larval viability. Recent advances also highlight the role of probiotics in reducing antibiotic dependence and supporting environmentally responsible aquaculture practices. Overall, probiotic-assisted nutritional strategies represent an effective and sustainable approach to improving aquaculture productivity, animal welfare, and production resilience, while contributing to the long-term sustainability of the aquaculture industry.

Keywords: Probiotics; Sustainable aquaculture; Functional feed additives; Fish nutrition; Gut microbiota; Disease resistance.

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Introduction

In developing countries aquaculture is the main source for production of animal protein as compared to the other sources of food production. In Asia, fish is the dominant source of protein so now recently the aquaculture industries are increasing tremendously to fulfill the need of fish growth for using as food source (FAO, 2019). In coming world for compensation of animal protein shortage, aquaculture plays an important role in their expansion. In the field of aquaculture, feed plays essential role for production outcomes. Sectors of feed are expanding day by day to meet their level of good health and growth of shrimp and fish also to reach the culturists potential requirements. Success in animals and humans feeding proved the efficiency of probiotics and it gains attention in aquaculture, in culturing of organism's probiotics show good effect on reducing stress, promoting growth and controlling disease spread (Ibrahim *et al.*, 2013).

Probiotics has now been used as a nutritional factor in aquaculture industry for enhancing function and physiology of gastrointestinal (Diplock *et al.*, 1999). This enhancement face many hurdles and also creates opportunities for scientists to develop new products, improve food quality and nutritional units for good health of host organisms. In aquaculture, probiotics administration seems to be advantageous for disease prevention, nutritional factors and biological control (Balcazar *et al.*, 2006) (Table 1).

History

This term was first used by Parker (1974) to describe it as a microbial food supplement.

He defines that it is an organism or substances that play role in microbial balance in intestine.

The modified term of probiotics was given by Lilly and Stillwell in 1965 from the word probiotika. The term was applied to explain that microorganisms produced substance that enhances the logarithmic growth phase of host animal. The use of probiotics was first applied in aquaculture by Kozasa (1986), and give information about advantageous role of probiotics on poultry and humans.

The first experimental attempt of the probiotic application in aquaculture was made by considering the beneficial effects of probiotics on humans and poultry (Kozasa, 1986) the swift development of probiotic use in aquaculture has been driven by the harmful consequences of widely applied antibiotics and broad-spectrum chemicals that destroy beneficial microbes along with disease causing bacteria in aquatic organism (Lakshmi *et al.*, 2013). Additionally, probiotics also work through different mechanisms in aquaculture system to eliminate the organic wastes and pollutants, as a result of incorporation of bioremediation and biocontrol when dealing with the environmental problems. In this context, probiotics can play an effective role in aquaculture production by providing greater non-specific disease protection as well as pollution free water sources (Panigrahi *et al.*, 2010; Nandi *et al.*, 2018).

In 1986, first time the probiotics were applied to test their ability to enhance the hydrobionts growth. Further probiotics were used for controlling infections of bacteria and improving water quality. Nowadays it is documented that probiotics

can improve reproductive efficacy, stress tolerance and nutrients digestibility. Recently new probiotics are producing commercially from different species of bacteria such as *Lactobacillus* sp., *Bacillus*

sp., *Carnobacterium* sp., *Enterococcus* sp., and the yeast *Saccharomyces cerevisiae* and it is recommended that their use is regulated carefully (Martínez *et al.*, 2012).

Table 1: Probiotics and its utilization.

Probiotics	Fish	Immune system	Nutrient utilization	References
<i>Bacillus subtilis</i>	Asian seabass (<i>Lates calcarifer</i>)	Upregulation of immune-related genes (IL-1b, TNF-a)	Enhanced apparent digestibility of dry matter and protein	(Adorian <i>et al.</i> , 2019)
	Nile tilapia (<i>Oreochromis niloticus</i>)	Increased lysozyme and phagocytic activity	Improved protein efficiency ratio and feed conversion ratio	(Abou-El-Atta <i>et al.</i> , 2019)
<i>Lactobacillus plantarum</i>	Common carp (<i>Cyprinus carpio</i>)	Increased lysozyme and phagocytic activity	Increased weight gain and performance, enhanced digestibility, improved feed conversion ratio	(Liu <i>et al.</i> , 2021)
<i>Lactobacillus rhamnosus</i>	Rainbowtrout (<i>Oncorhynchus mykiss</i>)	Enhanced immune cell activity, upregulation of immune-related genes, increased cytokine production, and improved lysozyme and complement activity	Enhanced protein utilization, improved liver function and detoxification, and improved feed conversion ratio	(Hajirezaee and Khan jani, 2023)
<i>Lactobacillus acidophilus</i>	Zebra fish (<i>Danio rerio</i>)	Increased serum lysozyme and complement activity	Improved growth performance and feed efficiency	(Hoseinifar <i>et al.</i> , 2015a)
Heat-killed <i>Lactobacillus plantarum</i>	Nile tilapia (<i>Oreochromis niloticus</i>)	Increased lysozyme activity, enhanced complement activity, and a boost in respiratory burst activity	Increased protein efficiency ratio and improved apparent digestibility of dry matter.	(Van Doan <i>et al.</i> , 2020)
Heat inactivated <i>Bacillus coagulans</i>	Common carp (<i>Cyprinus carpio</i>)	Increased serum lysozyme activity, higher complement C3 levels, and enhanced superoxide dismutase activity	Increased apparent digestibility of dry matter and crude protein, as well as enhanced intestinal protease and lipase activities.	(Xu <i>et al.</i> , 2016)

Probiotics application is a vast concept used for the enhancement of species such as aquatic organisms, pet and farm animals guided by the success of probiotics in human's medicine. It is noticed that antimicrobial production and attachment by lactic acid bacteria are the important factor in removing pathogens (Munoz-Atienza *et al.*, 2013). Lactic acid bacteria attachment to the mucus layer may be the first defensive barrier against the bacteria that cause disease or infections bacteria (Ringo *et al.*, 1998).

The ability of pathogenic bacteria attachment is related to causing virulence which is considered as its first step (Bengmark, 1998). The capacity of probiotics attachment to the mucosal layer intestine of fish is reported in the study (Vine *et al.*, 2006). Rinkinen *et al.* (2003) determined that the efficiency or capacity of attachment depend on bacterial strain, they are not host specific.

Types of probiotics (Table 2; Fig. 1)

Table 2: Types of probiotics (Rao *et al.*, 2016).

No.	Types	Description
1	Viable probiotics	Live form with guaranteed number of organisms has a protocol for counting and to be very steady and effective
2	Non-Viable Probiotics	Dead form of organism.
3	Fermentation probiotics	These are produced through fermentation.
4	Freeze-Dried Probiotics	These will expire speedily upon departure refrigeration

Selection criteria for probiotics

Numerous authors have described the necessary characteristics to qualify a microorganism as a probiotic. Necessary requirements for a probiotic to be effective and qualified as such are listed as follows (Kiron, 2015):

- 1- A suitable microorganism should be able to colonize the host and survive harsh conditions like bile, gastric juice, and host pH.
- 2- The probiotic candidate must be free of antibiotic-resistant genes and must not modify heritable traits of the host organism.
- 3- The microbe should benefit the host system by enhancing the growth and development of the immune system against pathogens. It also should have antimicrobial properties.

- 4- The probiotic candidate should not have harmful effects on the host. The evaluation of probiotics involves experiments conducted both outside and inside the living system in vitro or in vivo tests focus on resistance to bile and pH, adherence factors, anti-pathogenic effect, and non-antibiotic resistance. On the other hand, the in vivo evaluation of the probiotic candidate must show beneficial effects in the host (increasing the immune response, growth and absorption and utilization of food, modulation of intestinal microbiota, and reducing stress), not have harmful effects assessed by using a biosafety assay and improve the diseases resistance with an experimental challenge.

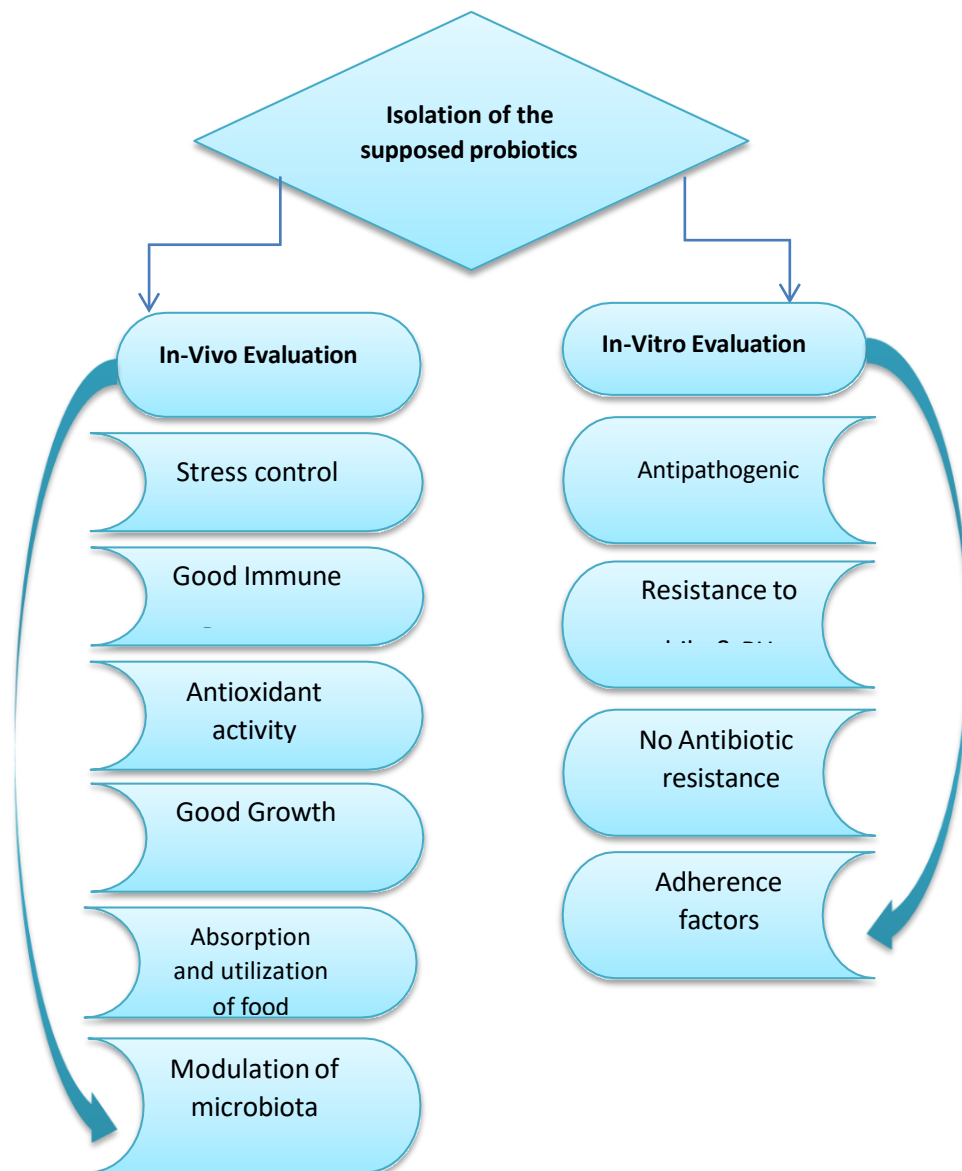


Figure 1: Probiotics types of flow-chart representation.

Application methods of probiotics

Application of probiotics based on its category, it is divided into two broad forms

- 1- Water probiotics are the bacterial species that proliferate in medium of water and reduce the growth of pathogenic bacteria by consuming all available nutrients and leaving no source of food for pathogenic bacteria
- 2- Gut probiotics are applied orally along with other feed ingredients to enhance gut microbial flora (Hasan and Banerjee, 2020) (Fig. 2).

Probiotics can be categories in several forms according to their mode of action (Hasan and Banerjee, 2020).

1. The type of probiotics called gut probiotics that can be applied orally with food of fish to enhance the efficiency of gut.
2. Type of probiotics that can be added in water medium, eliminate the pathogenic bacteria called water probiotics.

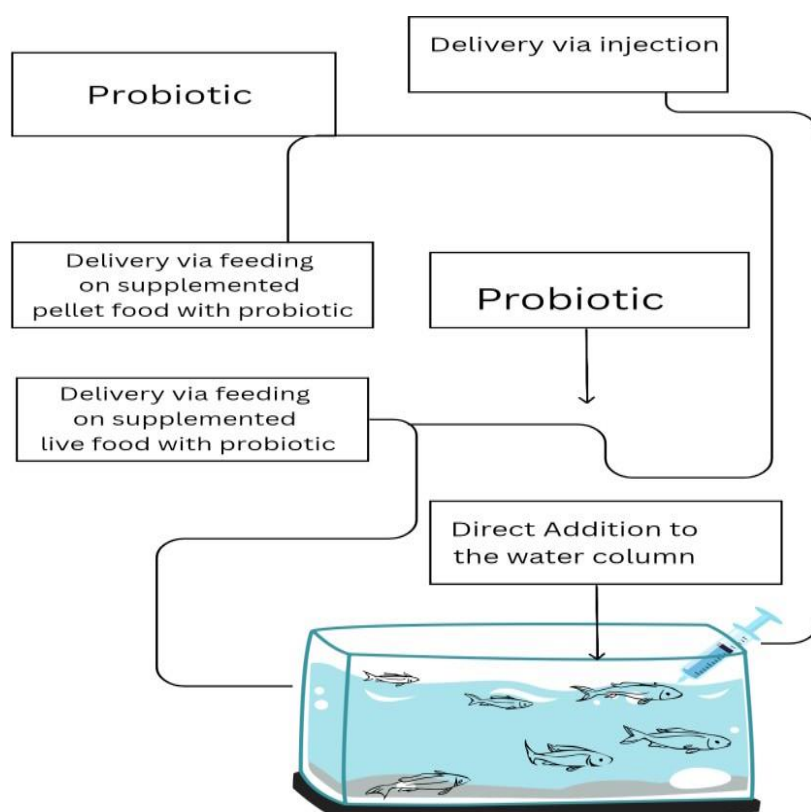


Figure 2: Probiotics application.

It is accepted widely that probiotics contain some important characteristics that increase the efficacy of agent at which it applied (Thakur *et al.*, 2016; Priyodip *et al.*, 2017).

Probiotic application

Probiotics can be applied in a several ways which was described (Ringo *et al.*, 2018; Banerjee and Ray, 2017).

1. Add in the culture water and artificial diet.
2. Addition of probiotics through live feed.

At first, you need to understand the mode of action and proper method of application for taking good results in aquaculture practices. Gram-negative facultative symbiotic anaerobes such as *Vibrio*, *Pseudomonas*, *Plesiomonas*, and *Aeromonas* were also reported to be potential probiotic candidates present in the gastro-intestinal tract (GIT) of fish and shellfish (Lakshmi *et al.*, 2013).

According to the food and agriculture organization [FAO] probiotics are recommended for improving aquatic environmental conditions by reducing morality and enhancing resistance to harmful pathogens furthermore probiotics candidates play an important role in enhancing the nutritional status of the host. The beneficial effects are temporal on occasion, depending on the time of application. There is a strong tendency to combine different photosynthetic bacteria, *Bacillus nitrifiers*, and denitrifiers together therefore, probiotics are often labeled as multifunctional and can be applied to various species under diverse culture conditions (Wang and Wang, 2008).

Probiotics are added constantly to fish cultures to maintain the health by enhancing the expression of several immunological factors, and to reduce the pathogen load to the gut mucus layer by

occupying the physical space (Banerjee and Ray, 2017). Moreover, probiotic candidate also plays a crucial role in nutrient enhancement in host. Hamdan *et al.* (2016) have reported the enhancement of crude lipid, total protein, and body weight in Nile tilapia (*Oreochromis niloticus*) fed with probiotic strain of *Lactobacillus* sp. The most commonly used probiotic preparations in this purpose are the lactic acid bacteria (Ringø *et al.*, 2018). These microorganisms are typically found in large population within the gut of healthy animals and are classified by the food and drugs administration It (FDA), is generally regarded as safe (GRAS status) (Giri *et al.*, 2013). Additionally, *Lactobacillus brevis* and *Bacillus subtilis* have been reported to generate significant quantity of *phytase subtilis* are capable of producing higher amount of enzyme phytase (up to 1,354,906.6 U/mL) facilitating the breakdown and use of plants phytase, chemically known as myo-inositol hexaphosphate (Priyodip *et al.*, 2017). Till date, several bacterial candidates (*Pseudomonas* sp., *Brevibacterium* sp., *Microbacterium* sp., *Agrobacterium* sp., and *Staphylococcus* sp.) have been reported to contribute in nutritional and metabolism physiology in Arctic charr (*Salvelinus alpinus*) (Ringø *et al.*, 2014).

Single and combination

Probiotics can be applied singly or in combination in different forms such as multistrain probiotics, probiotics with plant extract and probiotics with yeast extract (Shefat, 2018). The combination of *Bacillus megaterium* PTB 1.4 and *Pediococcus pentosaceus* E2211 administrated in feed showed best result than the single

application of either probiotic in catfish *Clarias* sp. (Hamka *et al.*, 2020). Combining probiotics often produces better outcomes than single-strain use, including higher survival rates in fish species such as Nile tilapia. (54.17%) and *Bacillus velezensis* (41.67%) (Van Doan *et al.*, 2018). The combination of probiotic *Bacillus coagulans* and plant extract *Mentha piperita* showed the higher growth performance, nutrient retention and immunity in *Catla catla* than their single application (Bhatnagar *et al.*, 2019).

Dose

The appropriate number of probiotics is influenced by the strain used, species of fish, their physiological state, farming conditions, and the objective of use. *Bifidobacterium* strains @ 1×10^7 CFU/g but not with 3×10^7 CFU/g (Sahandi *et al.*, 2019). This suggests that more probiotics in feed does not necessarily result in more increase in growth. Dose is an important consideration while supplementing functional nutrients in aquafeeds., While a specific dose enhanced growth in rainbow trout, higher doses did not help and even decreased weight gain in pabda catfish *Ompok pabda* (Chowdhury *et al.*, 2020) (Table 3).

Stress tolerance

Adding probiotics greatly improves the ability of aquaculture species to tolerate stress. Fish given *Lactobacillus plantarum* exhibited a smaller rise in cortisol under ammonia stress than those on a control diet, demonstrating decreased stress levels. The use of *Aspergillus oryzae* as a probiotic in Nile tilapia significantly improves the defense against hypoxia stress (Dawood *et*

al., 2019). Supplementation of dietary commercial probiotics at 0.2% has significantly higher saline water stress

tolerance than higher and lower doses in pabda catfish (Chowdhury *et al.*, 2020).

Table: 3 Dosage of probiotics.

Fish Specie	Probiotics	Dose	Duration	Beneficial effects	References
<i>O.niloticus</i>	<i>B. subtilis</i> and <i>B. licheniformis</i> (Biogen)	3g Kg ⁻¹	14 weeks	Significant improvement in growth parameters (weight gain, AWG, ADG, SGR, and FCR) and haematological parameters (haemoglobin, RBC count, PCV, blood platelets, and WBC count, plasma proteins)	(Mehrim, 2009)
<i>O.niloticus</i>	<i>B. subtilis</i> and <i>S. cerevisiae</i>	1.5gkg ⁻¹	6 weeks	Improved growth performance (weight gain, feed conversion ratio, and protein efficiency ratio)	(Marzouk <i>et al.</i> , 2008)
<i>Oncorhynchus mykiss</i>	<i>Bacillus species</i> and <i>Aeromonas sobria</i>	2×10 ⁸	2 weeks	Enhanced protection against multiple pathogens (<i>Aeromonas salmonicida</i> , <i>Streptococcus iniae</i> , <i>Yersinia ruckeri</i> , <i>Vibrio ordalii</i> , <i>Vibrio anguillarum</i> , and <i>Lactococcus garvieae</i>)	(Brunt <i>et al.</i> , 2007)
<i>O.niloticus</i>	<i>B. subtilis</i> and <i>L.plantarum</i>	10 ⁷ CFUg ⁻¹	60 days	Improved immunity (phagocytic, lysozyme, phenol oxidase, phosphatase activities and immunoglobulin)	(El-Ezabi <i>et al.</i> , 2011)
<i>Labeo rohita</i>	<i>B. subtilis</i> and <i>Lactobacillus lactis</i> / <i>B. subtilis</i> , <i>L. lactis</i> and <i>S. cerevisiae</i>	10 ¹¹ CFUg ⁻¹	60 days	Improved growth parameters (growth, protein efficiency ratio, nutrient retention and digestibility, lower feed conversion ratio), and micro ecology of gut	(Mohapatra <i>et al.</i> , 2014)
<i>O.niloticus</i>	<i>B. subtilis</i> and <i>B. licheniformis</i>	3g kg ⁻¹ , 5g kg ⁻¹ , 7g kg ⁻¹ and 10g kg ⁻¹	4 weeks	Enhanced immunity (lysozyme, protease, antiprotease, SOD, immunoglobulin M level in both serum and skin mucus, upregulation of C-lysozyme, HSP-70, b-defensin, transforming growth factor-β in mid-intestines and head kidney)	(Abarike <i>et al.</i> , 2018)

Probiotics and different types of food in aquaculture

Probiotic enriched formulated artificial balanced diet is beneficial for the health of fish furthermore its application is easy. Additionally, farmers have to be attentive of three main constraints (Vine *et al.*, 2006; Vilchez *et al.*, 2015) viz.,

- 1- Leaching of feed which reduces the availability of probiotic to the host. Thus, dose standardization and regular monitoring is required.
- 2- Probiotic candidates confer beneficial effects to the host only when they are alive and functional under different appropriate environmental conditions, which farmers must carefully consider.
- 3- Nature of various ponds differs depending on the physicochemical parameters and natural feeds (zooplankton and phytoplankton). So, application, types, and dose of probiotics will be varied accordingly.

As a result of all of this, a microorganism must meet the following criteria before it can be considered as a probiotic, being nonpathogenic, both for the host species and aquatic animals as a whole, as well as for human consumers; lacking antibiotic resistance genes encoded by plasmids; and tolerant of low to high pH as well as bile salt concentrations (>2.5%). Alternatively, key advantageous features include strong adherence successful colonization of epithelial surface and efficient growth. They should be appropriate for feed supplementation demonstrate favorable growth characteristics and process varying degree of anti-pathogenic activity. additionally this may produce beneficial enzymes or vitamins originates from the

host and its environment being native to both the host and the culturing environment; being robust enough to survive industrial processes and being viable under normal storage conditions; possessing better sensorial properties, possessing action of fermentation, a tolerance to freeze-drying and viability in feed during packaging and storage; promoting fish growth, boosting immunity and protecting fish from disease-causing microorganisms (Simon *et al.*, 2021). or live systems. Moreover, they must tolerate bile, gastric fluids, and host pH while maintaining the ability to grow and attach. Probiotic assessment is conducted through both in vitro and in vivo A probiotic is more likely to be effective if it meets several of these characteristics, though rarely all. Candidates are typically selected based on their ability to inhibit pathogens in laboratory methods (Amenyogbe *et al.*, 2022).

Nutrient competition

Nutrients are essential for bacterial growth. The use of similar nutrients gives rise to hostile competition among species (Wuertz *et al.*, 2021). The utilizing nutrients present in the environment probiotics reduces accessibility to pathogenic microorganism (Kuebutornye *et al.*, 2020). In fact, this restriction resulting from competition for nutrients is one of the main mechanisms used by probiotics to inhibit pathogens (Hoseinifar *et al.*, 2018). Iron is essential for harmful bacteria, as it contributes not only to their growth but also to their virulence. For instance, *Bacillus spp.* can synthesize siderophores and show greater carbon utilization (Khan *et al.*, 2017). The absence of iron and carbon limits microbes'

pathogenic functions. Furthermore, probiotics have the capacity to release a wide range of digestive enzymes. Thus, an increase in digestive enzymes can lead to the degradation of nutrients (Assan *et al.*, 2022).

Dawood *et al.* (2019) Research indicates that *Lactobacillus plantarum* significantly boosts digestive enzymes like amylase, lipase, and protease, with similar benefits observed from other probiotic strains and mixtures (Jang *et al.*, 2019). *Lactobacillus casei* combined with b-glucan and mannan oligosaccharide significantly increased the production of these enzymes and proteases in carp (*Cyprinus carpio*), *Citrobacter*, *Lactobacillus bulgaricus*, and *Lactobacillus acidophilus* also increase rainbow trout amylase, trypsin, and alkaline phosphatase (Mohammadian *et al.*, 2019). A study by Tarkhani *et al.* (2020) reported an increase in intestinal digestive enzyme activity in Caspian roaches (*Rutilus caspicus*) following the administration of *E. faecium*. Amenyogbe *et al.* (2022) also reported increased intestinal digestive enzyme activity in juvenile cobia (*Rachycentron canadum*) after supplementing indigenous isolates *Pantoea agglomerans* RCS2, indigenous isolates *Bacillus sp.* RCS1 and *Bacillus cereus* RCS3, and a mixture of *P. agglomerans* RCS2, *Bacillus sp.* RCS1 and *Bacillus cereus* RCS3. Even though these enzymes are linked to fish metabolism, their exact contribution remains unclear.

Prompt of decolonization (pathogenic bacteria)

For preventing colonization by pathogen, the possible mechanism is to compete physically for sites of attachment on host's

gut mucosal layer. It is called as ability to adhere to mucus and surface of wall is important for bacteria to become immersed in intestine of fish (Roeselers *et al.*, 2011; Martínez *et al.*, 2012). Adhesion of bacteria to the surface of tissue is effective in the early stage of infection of pathogens, the first effect of probiotics might be the competition for adhesion receptors with infectious pathogens (Chabrillón *et al.*, 2006).

Uses

Probiotics minimize, partially or completely, the growth of pathogenic bacteria (Akayli *et al.*, 2016). Xia *et al.* (2020) determined that it, also increase the response of immune system and enhance production of mucosal enzymes to promote the growth of host animal without causing any secondary pollution problem. Blood parameter variations of fish are a good indicator of water quality, health and nutrition (Ahmed *et al.*, 2020). El-Haroun *et al.* (2006) observed that probiotics start consumption of carbohydrates after their settlement in intestine and then produce wide range of digestive enzymes as protease, lipase and amylase which increase digestibility that directly influence the growth rate due to the free intestinal disorder and increase of pre-digestion secondary compounds. The commonly used probiotic in aquaculture belongs to yeast (*Debaryomyces*, *Phafia* and *Saccharomyces*), microalgae (*Tetraselmis*) and gram-negative bacteria (*Aeromonas*, *Al-teromonas*, *Photorhodobacterium*, *Pseudomonas* and *Vibrio*), gram positive (*Bacillus*, *Lactococcus*, *Micrococcus*, *Carnobacterium*, *Enterococcus*, *Lactobacillus*, *Streptococcus*, *Weissella*)

(Nayak 2013; Pandiyan *et al.*, 2013).

Bacillus subtilis and *Saccharomyces cerevisiae* are widely used probiotics in aquaculture due to their low cost and efficiency that can increase the feed utilization, and growth also they survive in the intestinal tract by colonizing in the gut of fish (Van, 2015; Allameh *et al.*, 2016; Hassaan *et al.*, 2018).

Probiotics enhance the larval nutrition and broodstock by establishing their presence in the gut of fish and providing enzymes (Protease, Amalyse, and lipase) and essential nutrients (fatty acids and protein). Fatty acids and protein are essential constituents of yolk that are considered as diet for maturation, development of oocytes and good rate of vitellogenesis (Dahlgren, 1980).

Lysozyme is considered as essential bactericidal enzymes having indispensable tool of innate immunity for fighting against pathogenic agents in fish. In serum, mucosal membrane of intestine and skin lysozyme is present. Single or combined probiotics are used to enhance the level of lysozyme in teleost (Adel *et al.*, 2017).

Intestine health

As a feed supplement probiotics play a key role in improving enzymatic contribution in intestine, value of feed intake, enhance anticarcinogenic and antimutagenic activity, reduce the growth of infectious microorganisms, increase immunity and growth promoting factor (Harikrishnan *et al.*, 2010). A study revealed that using probiotics strains can produce extracellular enzymes (lipase, protease, amylase) which show efficient result on digestive mechanisms of aquatic organisms and also produce growth factors e.g. fatty acids,

amino acids and vitamins (Balcazar *et al.*, 2006). The enhancement of intestine morphology includes intestine width, intestine length, lamina propria, goblet cells and area of villi were observed in fish that was treated with probiotics containing diet. Liver of probiotics treated fish show good result in regular nuclei shape and also decrease distance between liver tissues. Number of nuclei increase, diameter of hyperplastic muscles and muscle fiber were also observed while using multi strain species of probiotics in the feed of fish in comparison to control group (Hossain *et al.*, 2022). Probiotics are live microorganisms which are when used in balanced quantity can enhance health of host organism (Giannenas *et al.*, 2015; Egerton *et al.*, 2018). They have capacity to survive in the intestinal tract of animals. The advantageous effect of probiotics on fish is in controlling disease spread (Sharifuzzaman and Austin, 2017), survival and growth promoter (Kathia *et al.*, 2018). contribution of enzymes for digestion and feed efficiency (Egerton *et al.*, 2018; Mehrabi *et al.*, 2018), enhancement of immune system (Bagherzadeh *et al.*, 2018). Due to usage of probiotics the use of chemicals and antibiotics has reduced. Many of the probiotics applied through encapsulation, feeding, injection and immersion (Chizhayeva, et al., 2019).

Digestive enzymes

Taoka *et al.* (2007) determined that applying direct probiotics (*Clostridium butyricum*, *Bacillus subtilis*, *S. cerevisiae* and *Lactobacillus acidophilus*) or using it in feed can enhance the digestive enzymes production in tilapia. Metchnikoff (2004) was the first who determined the efficient

role of some good bacteria while testing on farmers who consumed milk having pathogenic bacteria reliance on gut microbes for food makes it possible to take steps to change the flora of our bodies and to replace harmful microbes by beneficial microbes.

Increase growth and survival rates

Probiotics increase the appetite of organism which enhances survival and growth rate. It is reported that using *Bacillus spp.* single or combine with other spp. of bacteria in Tilapia can efficiently improve survival and growth performance (weight gain, fresh weight, specific growth rate, average weight gain, feed efficiency, feed conversion ratio) (Gobi *et al.*, 2018; Galagarza *et al.*, 2018). Results have been reported in other similar article on microbial species such as *Micrococcus luteus* (Abd El-Rhman *et al.*, 2009), *Psychrobacter namhaensis* SO89 (Makled *et al.*, 2017), *Lactobacillus spp.* (Vílchez *et al.*, 2014; Xia, 2018), *E. faecium* (Mehisan *et al.*, 2015), *Aspergillus oryzae* (Dawood *et al.*, 2019) and *Pedio- coccus acidilactici* (Ferguson *et al.*, 2010).

Effects of probiotics on the reproductive performance and larval quality of fish

Nutritional effects are also known for efficient reproductive performance and gonadal development of fish. It is reported that diet which have low level of protein, lipid, vitamin E and C, fatty acids then carotenoids produced poor quality of eggs as compared to the diet which have balanced level of formulated diet for fish (Watanabe *et al.*, 1995). The reproductive health of fish is important for their offsprings. Quality of gametes in fish

reproduction is crucial for intensive culturing of fish species and fish farming industries. The most effective impairment in captive fish farming is to regulate the quality of gametes for the commercial fish farming development (Aydın and Çek-Yalınz, 2019). Some probiotics (good bacteria) not only enhance fecundity and ovulation but also promote quality of gametes, survival of larva and embryo, rate of hatchability (Vílchez *et al.*, 2014; Ahmadnia- Motlang *et al.*, 2017). In aquaculture the use of antibiotics is as preventive measure against various resistant pathogens of humans e.g *Escherichia tarda*, *Vibrio vulnificus*, *Aeromonas sp.*, *Vibrio parahaemolyticus*, *Escherichia coli*, *Vibrio cholerae*, and many more (Allameh *et al.*, 2016). Sai Gopal and Lakshmi Vishwanath (2013) reported about the development of resistivity against pathogens of aquatic environment under long term antibiotic pressure. So, the application of antibiotics is banned/restricted in many countries, then use probiotics in supplementary diet as an alternate which show beneficial results to control pathogens (Bandyopadhyay *et al.*, 2015; Wu *et al.*, 2015). In the field of aquaculture, the concept of probiotics application is so different from using in terrestrial animals. Now it is well known that application of probiotics could enhance the efficiency of guts by causing variation in community of microbes (Nayak, 2010).

Effects on phytoplankton

Probiotic bacteria play vital role in controlling algal growth, particularly of red tide plankton (Qi *et al.*, 2009). Bacteria antagonistic toward algae will be undesirable in green water larval rearing

technique in hatchery where unicellular algae are cultured but will be advantageous when undesired algae species are developed in the culture pond.

Bacteriostatic effects of probiotics

Bacterial populations of probiotics are able to secrete diverse chemical substances with bactericidal or bacteriostatic activity against Gram-negative as well as Gram-positive bacteria. These inhibitory compounds arise from different categories, including proteins (like lysozyme and proteases), chemical elements such as hydrogen peroxide, and

iron-binding compounds known as siderophores (lysozyme and different kind of proteases), chemical (hydrogen peroxides), and iron-chelating compound like siderophores (Giri *et al.*, 2013).

Augmentation in the immune system

Probiotics play the beneficial role as immunostimulatory to assist in the protection of aquatic cultured species by reducing the impact of diseases and entrance of pathogens (Hai, 2015; Dawood and Koshio, 2016) (Table 4).

Table 4: Effectiveness of probiotics against pathogen.

Probiotics	Fish	Pathogenic agent	Effectiveness	References
<i>Bacillus subtilis</i>	Red hybrid tilapia	<i>Streptococcus agalactiae</i>	Reduced mortalities	(Ng <i>et al.</i> , 2014)
<i>Lactobacillus plantarum</i>	Catfish	Viral hemorrhagic septicemia virus	Improves survival rate	(Hai, 2015)
<i>Bacillus subtilis</i> ,	Nile tilapia	<i>Streptococcus</i>	Competitive exclusion, immune enhancement	(Aly <i>et al.</i> , 2008)
<i>Flavobacterium sasangense</i>	Common carp	<i>A. hydrophila</i>	Enhance immune response and disease resistance	(Chi <i>et al.</i> , 2014)
<i>Bacillus subtilis</i>	Tilapia	Spring viremia of carp virus	Enhance immune response	(Tang <i>et al.</i> , 2021)
<i>Bacillus licheniformis</i>	Tilapia	<i>Streptococcus iniae</i>	Improved the disease resistance	(Han <i>et al.</i> , 2015)
<i>Lactobacillus acidophilus</i>	Nile tilapia	<i>Pseudomonas fluorescens</i> , <i>Streptococcus iniae</i>	Improve immune function and disease resistance	(Al-Dohail <i>et al.</i> , 2011)

Probiotic candidates, including *Pseudomonas* and *Vibrio* species, have been proven effective in combating infectious *hematopoietic necrosis virus* (IHNV) (Sahu *et al.*, 2008). Furthermore, *Paralichthys olivaceus* fed with *Sporolac* (*Lactobacillus* sp.) supplemented food develop resistance against lymphocystis disease virus (LCDV) (Harikrishnan *et al.*,

2010). Similar experiments have also proved the enhanced virus resistance power in grouper fish fed with probiotic strain of *Bacillus subtilis* E20 (Liu *et al.*, 2012)

Effect on reproduction

Few studies have examined how probiotics affect reproduction in aquaculture, although strains such as *Bacillus subtilis* and *Lactobacillus* species are commonly used

Lactobacillus casei. It is well researched that reproduction in different factors like fertilization, gonadosomatic index, fecundity, and production fry from the females is influenced by probiotics (Abasali and Mohammad, 2011). Recent study also reported that probiotics increase the daily numbers of ovulated eggs compared to control levels with higher hatching rate and faster embryonic development in zebrafish (Gioacchini *et al.*, 2013). Additionally, it is now profoundly accepted that probiotics reduce the concentration of the stress hormone cortisol and activate the expression of antioxidative enzymes (superoxide dismutase, catalase, and glutathione peroxidase) to increase the stress tolerance (Zolotukhin *et al.*, 2018). Probiotics are now gaining considerable global attention for both scientific research and commercial applications in aquaculture (Hoseinifar *et al.*, 2016; Banerjee and Ray, 2017)

Conclusion

Probiotics represent a well-established and sustainable strategy for enhancing aquaculture productivity and the health of cultured species, particularly in light of the rapid expansion of the industry and the increasing demand for environmentally responsible disease management practices. Accumulating evidence indicates that the efficacy of probiotics is contingent upon factors such as strain specificity, dosage, mode of administration, and host species, with multi-strain formulations frequently demonstrating synergistic interactions. Both experimental and field studies have consistently shown that probiotics contribute to the maintenance of gut microbial homeostasis, suppression of

pathogenic microorganisms, and modulation of host immune responses, thereby enhancing disease resistance. In addition, probiotic application has been associated with improved water quality and a reduced reliance on antibiotics, which is critical for limiting the development and spread of antimicrobial resistance in aquaculture systems. Overall, current findings support the strategic and sustained use of probiotics as an effective and eco-friendly approach for improving growth performance, physiological health, and disease resilience in aquaculture species.

References

- Abarike, E.D., Cai, J., Lu, Y., Yu, H., Chen, L., Jian, J. and Kuebutornye, F.K., 2018.** Effects of a commercial probiotic BS containing *Bacillus subtilis* and *Bacillus licheniformis* on growth, immune response and disease resistance in Nile tilapia, *Oreochromis niloticus*. *Fish and shellfish immunology*, 82, 229-238.
<https://doi.org/10.1016/j.fsi.2018.08.037>
- Abasali, H. and Mohamad, S., 2011.** Dietary prebiotic immunogen supplementation in reproductive performance of platy (*Xiphophorus maculatus*). *Veterinary research* (Pakistan), 2011, 4, 3, 66-70 ref. 27.
<https://doi.org/10.36478/vr.2011.66.70>
- Abd El-Rhman, A.M., Khattab, Y.A. and Shalaby, A.M., 2009.** *Micrococcus luteus* and *Pseudomonas* species as probiotics for promoting the growth performance and health of Nile tilapia, *Oreochromis niloticus*. *Fish and Shellfish Immunology*, 27(2), 175-180.
<https://doi.org/10.1016/j.fsi.2009.03.020>
- Abou-El-Atta, M.E., Abdel-Tawwab, M.,**

- Abdel-Razek, N. and Abdelhakim, T.M., 2019.** Effects of dietary probiotic *Lactobacillus plantarum* and whey protein concentrate on the productive parameters, immunity response and susceptibility of Nile tilapia, *Oreochromis niloticus* (L.), to *Aeromonas sobria* infection. *Aquaculture Nutrition*, 25(6), 1367-1377. <https://doi.org/10.1111/anu.12957>
- Adel, C.C. Lazado, R. Safari, S. and Yeganeh, M.J., 2017.** Zorriehzakra, Aqualase®, a yeast-based in-feed probiotic, modulates intestinal microbiota, immunity and growth of rainbow trout *Oncorhynchus mykiss*. *Aquaculture Research*, 48, 1815–1826, <https://doi.org/10.1111/aqr.13019>.
- Adorian, T.J., Jamali, H., Farsani, H.G., Darvishi, P., Hasanpour, S., Bagheri, T. and Roozbehfar, R., 2019.** Effects of probiotic bacteria *Bacillus* on growth performance, digestive enzyme activity, and hematological parameters of Asian sea bass, *Lates calcarifer* (Bloch). *Probiotics and antimicrobial proteins*, 11(1), 248-255. <https://doi.org/10.1007/s12602-018-9393-z>
- Ahmadnia-Motlagh, H., Hajimoradlo, A., Gorbani, R., AGH, N., Safari, O. and Lashkarizadeh-Bami, M., 2017.** Reproductive performance and intestinal bacterial changes of *Carassius auratus* fed supplemented lactoferrin and *Lactobacillus rhamnosus* PTCC 1637 diet. *Iranian Journal of Ichthyology*, 4(2), 150-161. <https://doi.org/10.22034/iji.v4i2.184>
- Ahmed, I., Reshi, Q.M. and Fazio, F., 2020.** The influence of the endogenous and exogenous factors on hematological parameters in different fish species: a review. *Aquaculture international*, 28(3), 869-899. <https://doi.org/10.1007/s10499-019-00501-3>
- Akayli, T., Albayrak, G., Ürkü Atanasov, Ç., Canak, Ö. and YÖRÜK, E., 2016.** Characterization of *Micrococcus luteus* and *Bacillus marisflavi* recovered from common dentex (*Dentex dentex*) larviculture system. *Mediterranean Marine Science*, 17(1). <https://doi.org/10.12681/mms.1322>
- Al-Dohail, M.A., Hashim, R. and Aliyu-Paiko, M., 2011.** Evaluating the use of *Lactobacillus acidophilus* as a biocontrol agent against common pathogenic bacteria and the effects on the haematology parameters and histopathology in African catfish, *Clarias gariepinus*, juveniles. *Aquaculture Research*, 42(2), 196-209. <https://doi.org/10.1111/j.1365-2109.2010.02606.x>
- Allameh, S.K., Yusoff, F. M., Ringø, E., Daud, H. M., Saad, C.R. and Ideris, A., 2016.** Effects of dietary mono-and multiprobiotic strains on growth performance, gut bacteria and body composition of Javanese carp (*Puntius gonionotus*, B leeker 1850). *Aquaculture nutrition*, 22(2), 367-373. <https://doi.org/10.1111/anu.12265>
- Aly, S.M., Abd-El-Rahman, A.M., John, G. and Mohamed, M.F., 2008.** Characterization of some bacteria isolated from *Oreochromis niloticus* and their potential use as probiotics. *Aquaculture*, 277(1-2), 1-6. <https://doi.org/10.1016/j.aquaculture.2008.02.021>
- Amenyogbe, E., Luo, J., Fu, W.J., Abarike, E. D., Wang, Z.L., Huang, J. S., and Chen, G., 2022.** Effects of autochthonous strains mixture on gut microbiota and metabolic profile in cobia (*Rachycentron canadum*). *Scientific Reports*, 12(1), 17410. <https://doi.org/10.1038/s41598-022-00501-3>

- 19663-x
- Assan, D., Kuebutornye, F.K.A., Hlordzi, V., Chen, H., Mraz, J., Mustapha, U.F. and Abarike, E.D., 2022.** Effects of probiotics on digestive enzymes of fish (finfish and shellfish); status and prospects: a mini review. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 257, 110653. <https://doi.org/10.1016/j.cbpb.2021.110653>
- Aydın, F. and Çek-Yalınz, Ş., 2019.** Effect of probiotics on reproductive performance of fish. *Natural and Engineering Sciences*, 4(2), 153-162. <https://doi.org/10.28978/nesciences.586541>
- Bagherzadeh Kasmani, F., Torshizi, K. and Mehri, M., 2018.** Effect of *Brevibacillus laterosporus* probiotic on hematology, internal organs, meat peroxidation and ileal microflora in *Japanese quails* fed aflatoxin B1. *Journal of Agricultural Science and Technology*, 20(3), 459-468.
- Balcazar, J.L., De Blas, I., Ruiz-Zarzuela, I., Cunningham, D., Vendrell, D. and Múzquiz, J.L., 2006.** The role of probiotics in aquaculture. *Veterinary microbiology*, 114(3-4), 173-186., Review. <https://doi.org/10.1016/j.vetmic.2006.01.009>
- Bandyopadhyay, P., Mishra, S., Sarkar, B., Swain, S.K., Pal, A., Tripathy, P.P. and Ojha, S.K., 2015.** Dietary *Saccharomyces cerevisiae* boosts growth and immunity of IMC *Labeo rohita* (Ham.) juveniles. *Indian Journal of Microbiology*, 55(1), 81-87. <https://doi.org/10.1007/s12088-014-0500-x>
- Banerjee, G. and Ray, A.K., 2017.** The advancement of probiotics research and its application in fish farming industries. *Research in veterinary science*, 115, 66-77. <https://doi.org/10.1016/j.rvsc.2017.01.016>
- Bengmark, S., 1998.** Ecological control of the gastrointestinal tract. The role of probiotic flora. *Gut*, 42(1), 2-7. <https://doi.org/10.1136/gut.42.1.2>
- Bhatnagar, A. and Saluja, S., 2019.** Synergistic effects of autochthonous probiotic bacterium and *Mentha piperita* diets in *Catla catla* (Hamilton, 1822) for enhanced growth and immune response. *Fisheries and Aquatic Sciences*, 22(1), 16. <https://doi.org/10.1186/s41240-019-0130-7>
- Brunt, J., Newaj-Fyzul, A. and Austin, B., 2007.** The development of probiotics for the control of multiple bacterial diseases of rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of Fish Diseases*, 30(10), 573-579. <https://doi.org/10.1111/j.1365-2761.2007.00836.x>
- Chabrillón, M., Arijo, S., Díaz-Rosales, P., Balebona, M.C. and Moriñigo, M.A., 2006.** Interference of *Listonella anguillarum* with potential probiotic microorganisms isolated from farmed gilthead seabream (*Sparus aurata*, L.). *Aquaculture Research*, 37(1), 78-86. <https://doi.org/10.1111/j.1365-2109.2005.01400>
- Chi, C., Jiang, B., Yu, X.B., Liu, T.Q., Xia, L. and Wang, G.X., 2014.** Effects of three strains of intestinal autochthonous bacteria and their extracellular products on the immune response and disease resistance of common carp, *Cyprinus carpio*. *Fish and shellfish immunology*, 36(1), 9-18. <https://doi.org/10.1016/j.fsi.2013.10.003>
- Chizhayeva, A., Amangeldi, A., Oleinikova, Y., Alybaeva, A., & Sadanov, A. (2022).** Lactic acid bacteria

- as probiotics in sustainable development of aquaculture. *Aquatic Living Resources*, 35, 10. <https://doi.org/10.1051/alr/2022011>
- Chowdhury, M.A. and Roy, N.C., 2020.** Probiotic supplementation for enhanced growth of striped catfish (*Pangasianodon hypophthalmus*) in cages. *Aquaculture Reports*, 18, 100504. <https://doi.org/10.1016/j.aqrep.2020.100504>
- Dahlgren, B.T., 1980.** The effects of three different dietary protein levels on the fecundity in the guppy, *Poecilia reticulata* (Peters). *Journal of Fish Biology*, 16(1), 83-97.
- Dawood, M.A.O., Eweedah, N.M., Moustafa Moustafa, E. and Shahin, M.G., 2019** Effects of feeding regimen of dietary *Aspergillus oryzae* on the growth performance, intestinal morphometry and blood profile of Nile tilapia (*Oreochromis niloticus*). *Aquaculture Nutrition*, 25(5), 1063–1072. <https://doi.org/10.1111/anu.12923>
- Dawood, M.A. and Koshio, S., 2016.** Recent advances in the role of probiotics and prebiotics in carp aquaculture: a review. *Aquaculture*, 454, 243-251. <https://doi.org/10.1016/j.aquaculture.2015.12.033>.
- Dawood, M.A., Magouz, F.I., Salem, M.F. and Abdel-Daim, H.A., 2019.** Modulation of digestive enzyme activity, blood health, oxidative responses and growth-related gene expression in GIFT by heat-killed *Lactobacillus plantarum* (L-137). *Aquaculture*, 505, 127-136. <https://doi.org/10.1016/j.aquaculture.2019.02.053>
- Diplock, A.T., Aggett, P., Ashwell, M., Bornet, F., Fern, E. and Roberfroid, M., 1999.** Scientific concepts of functional food science in Europe: consensus document. *British Journal of Nutrition*, 81, S1-S27. <https://doi.org/10.1079/BJN19990003>
- Egerton, S., Culloty, S., Whooley, J., Stanton, C. and Ross, R.P., 2018.** The gut microbiota of marine fish. *Frontiers in Microbiology*, 9, 873. <https://doi.org/10.3389/fmicb.2018.00873>
- El-Ezabi, M., El-Serafy, S., Essa, M., Daboor, S. and Esmael, N., 2011.** The viability of probiotics as a factor influencing the immune response in the Nile tilapia, *Oreochromis niloticus*. *Egyptian Journal of Aquatic Biology and Fisheries*, 15(1), 105-124. <https://doi.org/10.21608/ejabf.2011.2081>
- El-Haroun, E.R., Goda, A.S. and Kabir Chowdhury, M.A., 2006.** Effect of dietary probiotic Biogen® supplementation as a growth promoter on growth performance and feed utilization of Nile tilapia (*Oreochromis niloticus* L.). *Aquaculture research*, 37(14), 1473-1480. <https://doi.org/10.1111/j.1365-2109.2006.01584.x>.
- FAO, 2019.** FAO Yearbook. Fishery and Aquaculture Statistics 2017/FAO Annuaire. FAO, Rome.
- Ferguson, R.M., Merrifield, D.L., Harper, G.M., Rawling, M.D., Mustafa, S., Picchietti, S. and Davies, S. J., 2010.** The effect of *Pediococcus acidilactici* on the gut microbiota and immune status of on-growing red tilapia (*Oreochromis niloticus*). *Journal of applied microbiology*, 109(3), 851-862. <https://doi.org/10.1111/j.1365-2672.2010.04713.x>
- Galagarza, O.A., Smith, S.A., Drahos, D.J., Eifert, J.D., Williams, R.C. and Kuhn, D.D., 2018.** Modulation of innate

- immunity in Nile tilapia (*Oreochromis niloticus*) by dietary supplementation of *Bacillus subtilis* endospores. *Fish and shellfish immunology*, 83, 171-179. <https://doi.org/10.1016/j.fsi.2018.08.062>
- Giannenas, I., Karamaligas, I., Margaroni, M., Pappas, I., Mayer, E., Encarnação, P. and Karagouni, E., 2015.** Effect of dietary incorporation of a multi-strain probiotic on growth performance and health status in rainbow trout (*Oncorhynchus mykiss*). *Fish Physiology and Biochemistry*, 41(1), 119-128. <https://doi.org/10.1007/s10695-014-0010-0>
- Gioacchini, G., Dalla Valle, L., Benato, F., Fimia, G.M., Nardacci, R., Ciccocanti, F. and Carnevali, O., 2013.** Interplay between autophagy and apoptosis in the development of *Danio rerio* follicles and the effects of a probiotic. *Reproduction, fertility and development*, 25(8), 1115-1125. <https://doi.org/10.1071/RD12187>
- Giri, S.S., Sukumaran, V. and Oviya, M., 2013.** Potential probiotic *Lactobacillus plantarum* VSG3 improves the growth, immunity, and disease resistance of tropical freshwater fish, *Labeo rohita*. *Fish and shellfish immunology*, 34(2), 660-666. <https://doi.org/10.1016/j.fsi.2012.12.008>
- Gobi, N., Vaseeharan, B., Chen, J. C., Rekha, R., Vijayakumar, S., Anjugam, M. and Iswarya, A., 2018.** Dietary supplementation of probiotic *Bacillus licheniformis* Dahb1 improves growth performance, mucus and serum immune parameters, antioxidant enzyme activity as well as resistance against *Aeromonas hydrophila* in tilapia *Oreochromis mossambicus*. *Fish and shellfish immunology*, 74, 501-508. <https://doi.org/10.1016/j.fsi.2017.12.066>
- Hai, N.V., 2015.** The use of probiotics in aquaculture. *Journal of applied microbiology*, 119(4), 917-935. <https://doi.org/10.1111/jam.12886>
- Hajirezaee, S. and Khanjani, M.H., 2023.** Rosmarinic acid alone or in combination with *Lactobacillus rhamnosus* ameliorated resistance to ammonia stress in the rainbow trout, *Oncorhynchus mykiss*: growth, immunity, antioxidant defense and liver functions. *Annals of Animal Science*, 23(3), 819-831. <https://doi.org/10.2478/aoas-2023-0024>
- Hamdan, A.M., El-Sayed, A.F.M. and Mahmoud, M.M., 2016.** Effects of a novel marine probiotic, *Lactobacillus plantarum* AH 78, on growth performance and immune response of Nile tilapia (*Oreochromis niloticus*). *Journal of applied microbiology*, 120(4), 1061-1073. <https://doi.org/10.1111/jam.13099>
- Hamka, M.S., Meryandini, A. and Widanarni., 2020.** Growth performance and immune response of catfish *Clarias* sp. given probiotics *Bacillus megaterium* PTB 1.4 and *Pediococcus pentosaceus* E2211. <https://doi.org/10.19027/jai.19.1.50-60>
- Han, B., Long, W.Q., He, J.Y., Liu, Y.J., Si, Y.Q. and Tian, L.X., 2015.** Effects of dietary *Bacillus licheniformis* on growth performance, immunological parameters, intestinal morphology and resistance of juvenile Nile tilapia (*Oreochromis niloticus*) to challenge infections. *Fish and shellfish immunology*, 46(2), 225-231. <https://doi.org/10.1016/j.fsi.2015.06.018>
- Harikrishnan, R., Balasundaram, C. and Heo, M.S., 2010.** Effect of probiotics enriched diet on *Paralichthys olivaceus* infected with lymphocystis disease virus

- (LCDV). *Fish and Shellfish Immunology*, 29(5), 868-874. <https://doi.org/10.1016/j.fsi.2010.07.031>
- Hasan, K.N. and Banerjee, G., 2020.** Recent studies on probiotics as beneficial mediator in aquaculture: a review. *The Journal of Basic and Applied Zoology*, 81(1), 53. <https://doi.org/10.1186/s41936-020-00190-y>
- Hassaan, M.S., Soltan, M.A., Jarmolowicz, S. and Abdo, H.S., 2018.** Combined effects of dietary malic acid and *Bacillus subtilis* on growth, gut microbiota and blood parameters of Nile tilapia (*Oreochromis niloticus*). *Aquaculture Nutrition*, 24(1), 83-93. <https://doi.org/10.1111/anu.12536>
- Hossain, M.K., Hossain, M.M., Mim, Z.T., Khatun, H., Hossain, M.T. and Shahjahan, M., 2022.** Multi-species probiotics improve growth, intestinal microbiota and morphology of Indian major carp mrigal *Cirrhinus cirrhosus*. *Saudi Journal of Biological Sciences*, 29(9), 103399. <https://doi.org/10.1016/j.sjbs.2022.103399>
- Hoseinifar, S.H., Esteban, M.Á., Cuesta, A. and Sun, Y.Z., 2015a.** Prebiotics and fish immune response: a review of current knowledge and future perspectives. *Reviews in Fisheries Science and Aquaculture*, 23(4), 315-328. <https://doi.org/10.1080/23308249.2015.1052365>
- Hoseinifar, S.H., Ringø, E., Shenavar Masouleh, A. and Esteban, M.Á., 2016.** Probiotic, prebiotic and synbiotic supplements in sturgeon aquaculture: a review. *Reviews in Aquaculture*, 8(1), 89-
<https://doi.org/10.1111/raq.12082>
- Hoseinifar, S.H., Sun, Y.Z., Wang, A. and Zhou, Z., 2018.** Probiotics as means of diseases control in aquaculture, a review of current knowledge and future perspectives. *Frontiers in Microbiology*, 9, 2429. <https://doi.org/10.3389/fmicb.2018.02429>
- Ibrahim, M.D., Mohamed, M.F. and Ibrahim, M.A., 2013.** The role of *Spirulina platensis* (*Arthrospira platensis*) in growth and immunity of Nile tilapia (*Oreochromis niloticus*) and its resistance to bacterial infection. *Journal of Agricultural Science*, 5(6), 109-117. <https://doi.org/10.5539/jas.v5n6p109>
- Jang, W.J., Lee, J.M., Hasan, M.T., Lee, B.J., Lim, S.G. and Kong, I.S., 2019.** Effects of probiotic supplementation of a plant-based protein diet on intestinal microbial diversity, digestive enzyme activity, intestinal structure, and immunity in olive flounder (*Paralichthys olivaceus*). *Fish and Shellfish Immunology*, 92, 719-727. <https://doi.org/10.1016/j.fsi.2019.06.056>
- Kathia, C.M., del Carmen, M.D.M., Aida, H.P., Jorge, C.M., Félix, A.G. J. and Amadeo, B.M.J., 2018.** Effect of two probiotics on bacterial community composition from biofloc system and their impact on survival and growth of tilapia (*Oreochromis niloticus*). *International Journal of Fisheries and Aquatic Studies*, 6(2), 525-533.
- Khan, A., Doshi, H.V. and Thakur, M.C., 2017.** *Bacillus* spp.: a prolific siderophore producer. In *Bacilli and agrobiotechnology* (pp. 309-323). Cham: Springer International Publishing. https://doi.org/10.1007/978-3-319-44409-3_15
- Kiron, V., 2015.** Gastrointestinal microorganisms of fish and probiotics.

- Dietary Nutrients, Additives, and Fish Health*, 283-303. <https://doi.org/10.1002/9781119005568.ch15>
- Kozasa, M., 1986.** Toyocerin (*Bacillus toyoi*) as growth promotor for animal feeding. *Microbiol. Aliment. Nutrition*, 4, 121-135.
- Kuebutornye, F.K., Abarike, E.D., Lu, Y., Hlordzi, V., Sakyi, M.E., Afriye, G. and Xie, C.X., 2020.** Mechanisms and the role of probiotic *Bacillus* in mitigating fish pathogens in aquaculture. *Fish physiology and biochemistry*, 46(3), 819-841. <https://doi.org/10.1007/s10695-019-00754-y>
- Lakshmi, B., Viswanath, B. and Sai Gopal, D.V.R., 2013.** Probiotics as antiviral agents in shrimp aquaculture. *Journal of Pathogens*, 1, 424123. <https://doi.org/10.1155/2013/424123>.
- Lilly, D.M. and Stillwell, R.H., 1965.** Probiotics: growth-promoting factors produced by microorganisms. *Science*, 147(3659), 747-748. <https://doi.org/10.1126/science.147.3659.747>.
- Liu, C.H., Chiu, C.H., Wang, S.W. and Cheng, W., 2012.** Dietary administration of the probiotic, *Bacillus subtilis* E20, enhances the growth, innate immune responses, and disease resistance of the grouper, *Epinephelus coioides*. *Fish and Shellfish Immunology*, 33(4), 699-706. <https://doi.org/10.1016/j.fsi.2012.06.012>.
- Liu, Z.Y., Yang, H.L., Hu, L.H., Yang, W., Ai, C.X. and Sun, Y.Z., 2021.** Autochthonous probiotics alleviate the adverse effects of dietary histamine in juvenile grouper (*Epinephelus coioides*). *Frontiers in Microbiology*, 12, 792718. <https://doi.org/10.3389/fmicb.2021.792718>
- 18
- Makled, S.O., Hamdan, A.M., El-Sayed, A.F.M. and Hafez, E.E., 2017.** Evaluation of marine psychrophile, *Psychrobacter namhaensis* SO89, as a probiotic in Nile tilapia (*Oreochromis niloticus*) diets. *Fish and Shellfish Immunology*, 61, 194-200. <https://doi.org/10.1016/j.fsi.2017.01.001>
- Martínez Cruz, P., Ibáñez, A.L., Monroy Hermosillo, O.A. and Ramírez Saad, H.C., 2012.** Use of probiotics in aquaculture. *International scholarly Research Notices*, 1, 916845. <https://doi.org/10.5402/2012/916845>
- Marzouk, M.S., Moustafa, M.M. and Mohamed, N., 2008.** The influence of some probiotics on the growth performance and intestinal microbial flora of *O. niloticus*. In *Proceeding of the 8th International Symposium on Tilapia in Aquaculture* (pp. 1059-1071).
- Mehisan, M., El-moghazy, G., El-Sayed, A.E.K., Iraqi, M. and Soltan, M., 2015.** Role of probiotics in improving growth performance, immunity and controlling *Aeromonas hydrophila* in the Nile tilapia (*Oreochromis niloticus*). *Egyptian Journal of Aquatic Biology and Fisheries*, 19(3), 55-70. <https://doi.org/10.21608/EJABF.2015.2271>.
- Mehrabi, F., Khalesi, M. and Hazaie, K., 2018.** Effects of pre-and probiotics on growth, survival, body composition, and hematology of common carp (*Cyprinus carpio* L.) fry from the Caspian Sea. *Turkish Journal of Fisheries and Aquatic Sciences*, 18(4), 597-602. <http://doi.org/10.4194/1303-2712-v18i411>
- Mehrim, A.I., 2009.** Effect of dietary supplementation of Biogen® (Commercial probiotic) on mono-sex Nile tilapia (*Oreochromis niloticus*)

- under different stocking densities. *Journal of Fisheries and Aquatic Sciences*, 4(6), 261-273. <https://doi.org/10.3923/jfas.2009.261.273>
- Metchnikoff, I.I., 2004.** *The prolongation of life: Optimistic studies*. Springer Publishing Company.
- Mohammadian, T., Nasirpour, M., Tabandeh, M.R. and Mesbah, M., 2019.** Synbiotic effects of b-glucan, mannan oligosaccharide and *Lactobacillus casei* on growth performance, intestine enzymes activities, immune-hematological parameters and immune-related gene expression in common carp, *Cyprinus carpio*: An experimental infection with *Aeromonas hydrophila*. *Aquaculture*, 511, 634-197. <https://doi.org/10.1016/j.aquaculture.2019.06.005>
- Mohapatra, S., Chakraborty, T., Prusty, A.K., PaniPrasad, K. and Mohanta, K.N., 2014.** Beneficial effects of dietary probiotics mixture on hemato-immunology and cell apoptosis of *Labeo rohita* fingerlings reared at higher water temperatures. *PloS one*, 9(6), e100929. <https://doi.org/10.1371/journal.pone.0100929>
- Muñoz-Atienza, E., Gómez-Sala, B., Araújo, C., Campanero, C., Del Campo, R., Hernández, P.E. and Cintas, L.M., 2013.** Antimicrobial activity, antibiotic susceptibility and virulence factors of lactic acid bacteria of aquatic origin intended for use as probiotics in aquaculture. *BMC Microbiology*, 13(1), 15. <https://doi.org/10.1186/1471-2180-13-15>
- Nandi, A., Banerjee, G., Dan, S.K., Ghosh, K. and Ray, A.K., 2018.** Evaluation of in vivo probiotic efficiency of *Bacillus amyloliquefaciens* in *Labeo rohita* challenged by pathogenic strain of *Aeromonas hydrophila* MTCC 1739. *Probiotics and Antimicrobial Proteins*, 10(2), 391-398. <https://doi.org/10.1007/s12602-017-9317-6>
- Nayak, S.K., 2010.** Probiotics and immunity: a fish perspective. *Fish and Shellfish Immunology*, 29(1), 2-14. <http://doi.org/10.1016/j.fsi.2010.02.017>
- Nayak, S.K., 2013.** Use of lactic acid bacteria in fish farming.
- Ng, W.K., Kim, Y.C., Romano, N., Koh, C.B. and Yang, S.Y., 2014.** Effects of dietary probiotics on the growth and feeding efficiency of red hybrid tilapia, *Oreochromis* sp., and subsequent resistance to *Streptococcus agalactiae*. *Journal of Applied Aquaculture*, 26(1), 22-31. <https://doi.org/10.1080/10454438.2013.874961>
- Pandiyan, P., Balaraman, D., Thirunavukkarasu, R., George, E.G.J., Subaramaniyan, K., Manikkam, S. and Sadayappan, B., 2013.** Probiotics in aquaculture. *Drug invention today*, 5(1), 55-59. <http://doi.org/10.1016/j.dit.2013.03.003>
- Panigrahi, A., Kiron, V., Satoh, S. and Watanabe, T., 2010.** Probiotic bacteria *Lactobacillus rhamnosus* influences the blood profile in rainbow trout *Oncorhynchus mykiss* (Walbaum). *Fish Physiology and Biochemistry*, 36(4), 969-977. <http://doi.org/10.1007/s10695-010-9369-6>
- Parker, R.Á., 1974.** Probiotics, the other half of the antibiotics story. *Animal Nutrition Health*, 29, 4-8.
- Priyodip, P., Prakash, P.Y. and Balaji, S., 2017.** Phytases of probiotic bacteria: characteristics and beneficial aspects. *Indian Journal of Microbiology*, 57(2), 148-154. <http://doi.org/10.1007/s12088-016-0637-2>
- Qi, Z., Zhang, X.H., Boon, N. and**

- Bossier, P., 2009.** Probiotics in aquaculture of China—Current state, problems and prospect. *Aquaculture*, 290(1-2), 15-21. <http://doi.org/10.1016/j.aquaculture.2009.02.012>
- Rao, S.C., Athalye-Jape, G.K., Deshpande, G.C., Simmer, K.N. and Patole, S.K., 2016.** Probiotic supplementation and late-onset sepsis in preterm infants: A meta-analysis. *Pediatrics*, 137(3), e20153684. <http://doi.org/10.1542/peds.2015-3684>
- Ringo, E., Bendiksen, H.R., Gausen, S.J., Sundsfjord, A. and Olsen, R.E., 1998.** The effect of dietary fatty acids on lactic acid bacteria associated with the epithelial mucosa and from faecalia of *Arctic charr*, *Salvelinus alpinus* (L.). *Journal of Applied Microbiology*, 85(5), 855-864. <http://doi.org/10.1111/j.1365-2672.1998.00593.x>
- Ringo, E., Hoseinifar, S.H., Ghosh, K., Van Doan, H., Beck, B.R. and Song, S.K., 2018.** Lactic acid bacteria in finfish—an update. *Frontiers in Microbiology*, 9, 1818. <http://doi.org/10.3389/fmicb.2018.01818>
- Rinkinen, M., Jalava, K., Westermarck, E., Salminen, S. and Ouwehand, A.C., 2003.** Interaction between probiotic lactic acid bacteria and canine enteric pathogens: a risk factor for intestinal *Enterococcus faecium* colonization. *Veterinary Microbiology*, 92(1-2), 111-119. [http://doi.org/10.1016/S0378-1135\(02\)00356-5](http://doi.org/10.1016/S0378-1135(02)00356-5)
- Roeselers, G., Mittge, E.K., Stephens, W.Z., Parichy, D.M., Cavanaugh, C.M., Guillemin, K. and Rawls, J.F., 2011.** Evidence for a core gut microbiota in the zebrafish. *The ISME journal*, 5(10), 1595-1608. <http://doi.org/10.1038/ismej.2011.38>
- Sahandi, J., Jafaryan, H., Soltani, M. and Ebrahimi, P., 2019.** The use of two *Bifidobacterium* strains enhanced growth performance and nutrient utilization of Rainbow Trout (*Oncorhynchus mykiss*) fry. *Probiotics and Antimicrobial Proteins*, 11(3), 966-972. <http://doi.org/10.1007/s12602-018-9455-2>
- Sahu, M.K., Swarnakumar, N.S., Sivakumar, K., Thangaradjou, T. and Kannan, L., 2008.** Probiotics in aquaculture: importance and future perspectives. *Indian journal of microbiology*, 48(3), 299-308. <http://doi.org/10.1007/s12088-008-0024-3>
- Sharifuzzaman, S. Á. and Austin, B., 2017.** Probiotics for disease control in aquaculture. *Diagnosis and control of Diseases of Fish and Shellfish*, 189-222. <http://doi.org/10.1002/9781119152125.ch8>
- Shefat, S.H.T., 2018.** Probiotic strains used in aquaculture. *International Research Journal of Microbiology*, 7(2), 43-55. <http://doi.org/10.14303/irjm.2018.023>
- Simon, R., Docando, F., Nuñez-Ortiz, N., Tafalla, C. and Díaz-Rosales, P., 2021.** Mechanisms used by probiotics to confer pathogen resistance to teleost fish. *Frontiers in Immunology*, 12, 653025. <http://doi.org/10.3389/fimmu.2021.653025>
- Tang, K.F., Bondad-Reantaso, M.G., Surachetpong, W., Dong, H.T., Fejzic, N., Wang, Q., and Hao, B., 2021.** *Tilapia lake virus disease strategy manual* (Vol. 1220. Food and Agriculture Org. <https://doi.org/10.4060/cb7293en>
- Taoka, Y., Maeda, H. and Jo, J., Sakata T., 2007** Influence of commercial probiotics on the digestive enzyme activities of *Tilapia*, *Oreochromis niloticus*. *Aquaculture Science*,

- 55(2),183–189.
<https://doi.org/10.11233/aquaculturesci1953.55.183>
- Tarkhani, R., Imani, A., Hoseinifar, S.H., Moghanlou, K.S. and Manaffar, R., 2020.** The effects of host-associated *Enterococcus faecium* CGMCC1. 2136 on serum immune parameters, digestive enzymes activity and growth performance of the Caspian roach (*Rutilus rutilus caspicus*) fingerlings. *Aquaculture*, 519,734741. <https://doi.org/10.1016/j.aquaculture.2019.734741>
- Thakur, N., Rokana, N. and Panwar, H., 2016.** Probiotics, Selection criteria, safety and role in health and. *Journal of Innovative Biology January*, 3(1), 259-270.
- Van Doan, H., Hoseinifar, S.H., Khanongnuch, C., Kanpiengjai, A., Unban, K., Van Kim, V. and Srichaiyo, S., 2018.** Host-associated probiotics boosted mucosal and serum immunity, disease resistance and growth performance of Nile tilapia (*Oreochromis niloticus*). *Aquaculture*, 491, 94-100. <https://doi.org/10.1016/j.aquaculture.2018.03.019>
- Van Doan, H., Hoseinifar, S.H., Ringø, E., Ángeles Esteban, M., Dadar, M., Dawood, M.A. and Faggio, C., 2020.** Host-associated probiotics: a key factor in sustainable aquaculture. *Reviews in Fisheries Science and Aquaculture*, 28(1), 16-42. <https://doi.org/10.1080/23308249.2019.1643288>
- Van Hai, N., 2015.** Research findings from the use of probiotics in tilapia aquaculture: a review, *Fish Shellfish Immunol.*, 45, 592–597, <https://doi.org/10.1016/j.fsi.2015.05.026>
- Vílchez, M.C., Santangeli, S., Maradonna, F., Gioacchini, G., Verdenelli, C., Gallego, V., and Villamil, L., Reyes, C., Martínez-Silva, M.A., 2014** *In vivo* and *in vitro* assessment of *Lactobacillus acidophilus* as probiotic for tilapia (*Oreochromis niloticus*), Perciformes: Cichlidae) culture improvement. *Aquacult Res*, 45(7),1116–1125. <https://doi.org/10.1111/are.12051>
- Vine, N.G., Leukes, W.D. and Kaiser, H., 2006.** Probiotics in marine larviculture. *FEMS Microbiology Reviews*, 30(3), 404-427. <https://doi.org/10.1016/j.1574-9762.2006.00017.x>
- Wang, Y.M. and Wang, Y.G., 2008.** Advance in the mechanisms and application of microecologies in aquaculture. *Progress in Veterinary Medicine*, 29(6), 72-75.
- Watanabe, T. and Kiron, V., 1995.** Broodstock management and nutritional approaches for quality offsprings in the Red Sea Bream. *Broodstock Management and Egg and Larval Quality*, 154-195. https://doi.org/10.1007/978-1-4615-1867-2_9
- Wu, Z.Q., Jiang, C., Ling, F. and Wang, G.X., 2015.** Effects of dietary supplementation of intestinal autochthonous bacteria on the innate immunity and disease resistance of grass carp (*Ctenopharyngodon idellus*). *Aquaculture*, 438, 105-114. <https://doi.org/10.1016/j.aquaculture.2015.01.011>
- Wuertz, S., Schroeder, A. and Wanka, K.M., 2021.** Probiotics in fish nutrition—long-standing household remedy or native nutraceuticals? *Water*, 13(10), 1348. <https://doi.org/10.1039/w13101348>
- Xia, Y., Lu, M., Chen, G., Cao, J., Gao,**

- F., Wang, M., and Yi, M., 2018.** Effects of dietary *Lactobacillus rhamnosus* JCM1136 and *Lactococcus lactis* subsp. *lactis* JCM5805 on the growth, intestinal microbiota, morphology, immune response and disease resistance of juvenile Nile tilapia, *Oreochromis niloticus*. *Fish and shellfish immunology*, 76, 368-379. <https://doi.org/10.1016/j.fsi.2018.03.020>
- Xia, Y., Wang, M., Gao, F., Lu, M. and Chen, G., 2020.** Effects of dietary probiotic supplementation on the growth, gut health and disease resistance of juvenile Nile tilapia (*Oreochromis niloticus*). *Animal Nutrition*, 6(1), 69-79. <https://doi.org/10.1016/j.aninu.2019.07.002>
- Xu, H.J., Jiang, W.D., Feng, L., Liu, Y., Wu, P., Jiang, J., Kuang, S.Y., Tang, L., Tang, W.N., Zhan, Y.A. and Zhou, X.Q., 2016.** Dietary vitamin C deficiency depressed the gill physical barriers and immune barriers referring to Nrf2, apoptosis, MLCK, NF- κ B and TOR signaling in grass carp (*Ctenopharyngodon idella*) under infection of *Flavobacterium columnare*. *Fish and shellfish immunology*, 58, 177-192. <https://doi.org/10.1016/j.fsi.2016.09.029>
- Zolotukhin, P.V., Prazdnova, E.V. and Chistyakov, V.A., 2018.** Methods to assess the antioxidative properties of probiotics. *Probiotics and Antimicrobial proteins*, 10(3), 589-599. <https://doi.org/10.1007/s12602-017-9375-6>