



Harnessing the environmental and economic potential of seaweed to create a resilient future of aquaculture

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Abstract

From the past 25 years the demand of macro-algae increase globally and its culture grown rapidly due to their environmental, nutritional and industrial strengths. It is also known for their rich content of lipids, fibers, protein, minerals and bioactive compound that make it priceless for the consumption of human animals, and aquatic organisms also use in pharmaceutical, nutraceutical and in cosmetics. Due to their rich content of polyunsaturated fatty acids and vital amino acids, macro-algae are developing as a sustainable substitute for fish oil in aquaculture feed. Historically employed in agriculture for their plant-growth-promoting properties, seaweeds continue to provide to enhancing crop yield. Furthermore, seaweed farming supports environmental stewardship by enhancing coastal ecosystem health and contributing a viable source of biofuel through anaerobic digestion. The diversity of seaweed species adaptable to various climates from tropical to temperate underscores the importance of increased investment in scientific exploration, technological advancement, and strategic policy-making to accelerate the development of the seaweed industry and its contribution to advancing the blue economy.

Keywords:

Seaweed, Sargassum, Macro-algae, Cultivation

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Introduction

Macroalgae, or seaweed, are photosynthetic marine organisms from the kingdoms *Chromista* or *Plantae*, serving as key primary producers in coastal ecosystems. The contribution of seaweed aquaculture to the economics of coastal communities is the employment of the millions of people; with 96% of all aquaculture engagement located primarily in Asian countries (FAO, 2020). It has been estimated that macroalgae have 9000 species worldwide (Khan *et al.*, 2009). They are plant-like in form but they lack true roots, stems, and vascular tissue (Dawes, 2016). Beyond its commercial importance, seaweed cultivation plays a key role in promoting environmental sustainability by improving diverse ecosystem functions. It supports ecological processes such as nutrient cycling, primary production, species diversity, and habitat integrity, while also aiding in the regulation of sediment flow, atmospheric interactions, and biological systems, including the reduction of eutrophication (Hasselström *et al.*, 2018).

Seaweed play an important role in maintaining the food chain in aquatic environment, and the absence of macroalgae show the disturbance in the water body (Khan *et al.*, 2009). Seaweed is considered as the living resources of marine and also known as the wealth of ocean (Paul *et al.*, 2007). The seaweed can store good amount of carbon and lessen its free level. As it is observed that global warming is intimidating remark seen on earth (Botkin *et al.*, 2007; De Schryver *et al.*, 2009). They amend the

ecosystem of marine and reserve the diversity. Various organisms of marine are seen adhered near the algal biodiversity, and play a productive role in the food cycle (Jones *et al.*, 2000). Single kelp can engage 8000 macro invertebrates organisms (Burrows *et al.*, 2014) and the individual engaged here will elevate with increase in the number of algae and provide larger habitat area (Christie *et al.*, 2003). Sea cucumbers, starfish, shrimp, snails, and crabs are examples (Burrows *et al.*, 2014).

Nutritional profile

Commonly, macro-algae comprise macronutrients including lipids (vital fatty acids, n-3 and n-6), proteins (essential amino acids), and carbohydrates (dietary fiber), micronutrients, such as vitamins and minerals. This composition has good health benefits on the health of living organism's e.g, anti-obesity, anti-oxidant, antibacterial and anti-inflammatory (Buschmann *et al.*, 2017). High amount of various vitamins (A, K and B12), present in seaweeds, minerals, protective pigments and trace elements that are important in the diet of humans, might collaborate with European union permitted nourishing entitlements (such as iron, iodine, magnesium or calcium) linked with cognitive function, bone health, muscle function, normal growth and maintainance of normal metabolism (Mabeau S and Fleurence, 1999; Macartain *et al.*, 2007; Barbier *et al.*, 2019) (Fig. 1).

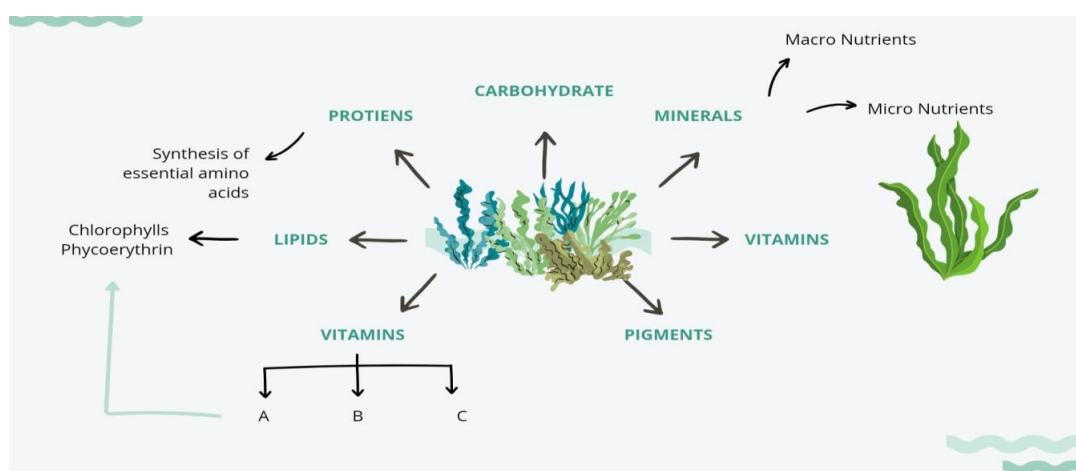


Figure 1: Nutritional components in macro-algae.

Some macro-algae are a great source of protein, carbohydrates, and vitamins A, B, B2, and C. Eatable seaweed has polysaccharides (starch, hydrocolloids, floridoside, laminarin, cellulose and hemicellulose such as agar, carrageenan and alginate) vitamins (A, B1, B2, B9 (folic acid), B12, C, D, E, and K), protein, minerals (Na, I, Mg, Fe, K, Ca, Se, Zn, and F), polyphenols (flavonols, phlorotannins, and catechins) low amount of fat which are poly and monounsaturated with little caloric value and antioxidants (polyphenols, vitamin C and E, carotenoids, sulphated polysaccharides, sterols, phlorotannins, proteins and catechins) (MacArtain *et al.*, 2007; Fernández *et al.*, 2018; Pandey *et al.*, 2020). In addition to these, there are several trace elements and minerals, with iodine being the most notable. Its low calorie content and suitability for all types of vegetarians is an added benefit. Minerals like calcium, potassium, sodium, sulphur, magnesium, phosphorus, chlorine micronutrients that are present in different seaweed are iron, copper, molybdenum, manganese, nickel, iodine, cobalt, zinc, selenium,

fluoride and boron. Normally brown seaweed has (upto 15%) protein whereas red and green seaweed have high protein (upto 30%) (Kolanjinathan *et al.*, 2014).

Many of its species include a variety of minerals; brown seaweed is typically the best provider of iodine. Although the amount of protein and calcium varies from species to species, the amount of fat is minimal. In general, the protein content of red and green seaweeds is higher (up to 30%), whereas that of brown seaweeds is lower (up to 15%) (Kolanjinathan *et al.*, 2014). Glutamic and aspartic acid collectively constitute enough portion of amino acid in various seaweed (Astorga *et al.*, 2016; Bikker *et al.*, 2020) mainly in *A. nodosum* (38.22% of total amino acids) (Kadam *et al.*, 2017), *U. rotundata* (32%), *U. rigida* (26%) and *Fucus spp.*, 22–44%, (Fleurence, 1999) *C. crispus* (38.62%), *Gracilaria spp.*, 25.82%), *Ulva spp.*, formerly *Enteromorpha spp.*) (28.11%) (Kazir *et al.*, 2019).

The minimum amount of carbohydrates is recorded in *Dictyota dichotoma* a brown seaweed (10.63%) and highest amount of carbohydrates are

seen in green macro algae *E. intestinalis* 28.58 % (Parthiban *et al.*, 2013). Chakraborty and Santra 2008, reported the content of carbohydrates in *E. intestinalis* (30.58%) and *U. lactuca* (35.27%).

Types of seaweed

Macroalgae categorize on the basis of presence of photosynthetic pigment and cell wall structure *Rhodophyta* (red), *Chlorophyta* (green), and *Phaeophyta* (brown). Their color is due the pigments fucoxanthin, chlorophyll and phycoerythrin (Khan *et al.*, 2009). Among 200 known species of seaweed, nearly 10 species of macroalgae are cultivated extensively which contain brown seaweeds (*Undaria pinnatifida*, *Saccharina japonica*, *Sargassum fusiforme*), green seaweeds (*Cauleurpa* spp., *Monostroma nitidum*, *Enteromorpha clathrata*) and red seaweeds (*Kappaphycus alvarezii*, *Eucheuma* spp., *Porphyra* spp., *Gracilaria* spp.) (FAO, 2018).

Green seaweed

Chlorella, *enteromorpha* and *ulva* are predominant seaweed. Approximately 305 species of macroalgae with blade-like or filamentous morphologies are known to exist in the genus *Ulva*. Many species of *Ulva* are edible (Pereira, 2016), and they typically have intriguing traits that make them very appealing for aquaculture, including broad distribution, rapid growth, high environmental tolerance, low susceptibility to epiphytism, and high

capacities for nutrient uptake (Carl *et al.*, 2014; Hiraoka *et al.*, 2020).

The green macroalgae *Enteromorpha intestinalis* and *Ulva lactuca* found to have enough contribution in its carbohydrate content i.e 30.58% and 35.27% (Chakraborty and Santra, 2008; Parthiban *et al.*, 2013). The content of fiber in edible seaweed varies from 33-62% of dry mass which high in amount as compare to other higher plants. In seaweed lipid content concentration varies from 1.33% in *E. intestinalis* and 4.6% in *E. clathrata*. It is reported in some studies the amount of lipid, that is 1.33% in *Kappaphycus alvarezii* and *Utricularia rigida* 12%, respectively (Satpati and Pal, 2011; Rajasulochana *et al.*, 2012) (Fig. 2).

Green seaweed has acidic nature polysaccharide e.g sulfated galactans, xylans and sulfated polysaccharides that present in the green seaweed's cell wall (Wu *et al.*, 2020; Wang *et al.*, 2020b, Wassie *et al.*, 2021; Li *et al.*, 2021a; Cao *et al.*, 2022; Chen *et al.*, 2022). β -carotene have antioxidant quality that can be formed by microalgal *Dunaliella salina* that help to control the damaging effect of free radicals, that can cause serious problems of life including coronary heart disease, arthritis, premature aging and cancer (Dembitsky and Maoka, 2007; Miyashita, 2009). Astaxanthin are produced from *haematococcus pluvialis*, it's a red component of carotenoid make astaxanthin interesting by showing antioxidant properties, anti-inflammatory, anti-diabetic and anti-cancer (Ambati *et al.*, 2014; El-Baz *et al.*, 2018) (Table 1).

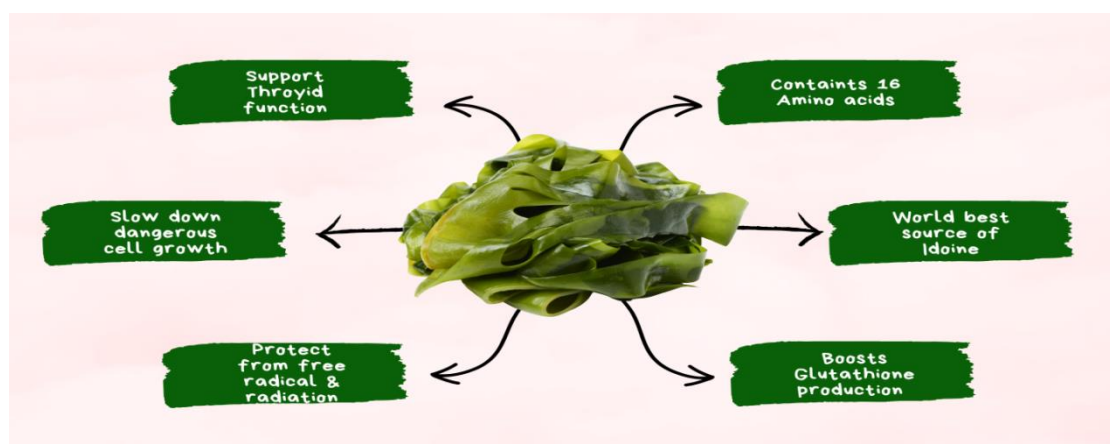


Figure 2: Functions of Green macro-algae.

Table 1: Role of green seaweed.

Green Macroalgae	Compound	Bioactivity	References
<i>Ulva fasciata</i>	Sulpholipids	Antibacterial	El Baz <i>et al.</i> , 2018
-	-	Antiviral	-
-	-	Antitumoral	-
<i>Codium vermilara</i>	Sulphated polysaccharides	Anticoagulant	Ciancia <i>et al.</i> , 2007
<i>Codium dwarkense</i>	-	-	Siddhanta <i>et al.</i> , 1999
<i>Ulva australis</i>	-	Antioxidant	Qi <i>et al.</i> , 2005
<i>Ulva rigida</i>	-	Antitumoral immunodulatory	Leiro <i>et al.</i> , 2007
<i>Ulva prolifera</i>	-	-	Kim <i>et al.</i> , 2011
<i>Monostroma latissimum</i>	-	Antiviral	Kazłowski <i>et al.</i> , 2012
<i>Codium fragile</i>	-	Antiviral	Ohta <i>et al.</i> , 2009
<i>Codium fragile</i>	Siphonaxanthin	Antitumorl	Ganesan <i>et al.</i> , 2010
-	-	Antiangiogenic	Ganesan <i>et al.</i> , 2011
<i>Ulva armoricana</i>	Glycolipids	Antitumoral	Kendel <i>et al.</i> , 2015
<i>Codium tomentosum</i>	Lipids	Antioxidants	Rey <i>et al.</i> , 2020
<i>Caulerpa racemosa</i>	Squalene	Anti-inflammatory	Fernando <i>et al.</i> , 2018
<i>Bryopsis</i> spp	Depsipeptides	Antiviral	Suárez <i>et al.</i> , 2003
-	-	Antimalarial	-

Red seaweed

The Rhodophyta (red algae) are red in color due to the pigments (Knowler *et al.*, 2020). Important pigment named Phycocyanin seen in red seaweed (Cian *et al.*, 2014). The profitable use of this compound is used as natural dyes in soft drinks, chewing gums, cosmetics and in dairy products e.g eyeliner and lipstick (Spolaore *et al.*, 2006). In all that, these compounds are good for advantageous bioactivities, so they are represented as nutraceutical products (Sekar and

Chandramohan, 2008). Component extracted from seaweed called Carrageenan. It is taken from one nominated group named carrageenophytes which is known as carrageenan producers; belong to the family Gigartinales (Gurgel *et al.*, 2007; Pereira and Mesquita, 2003) (Fig. 3).

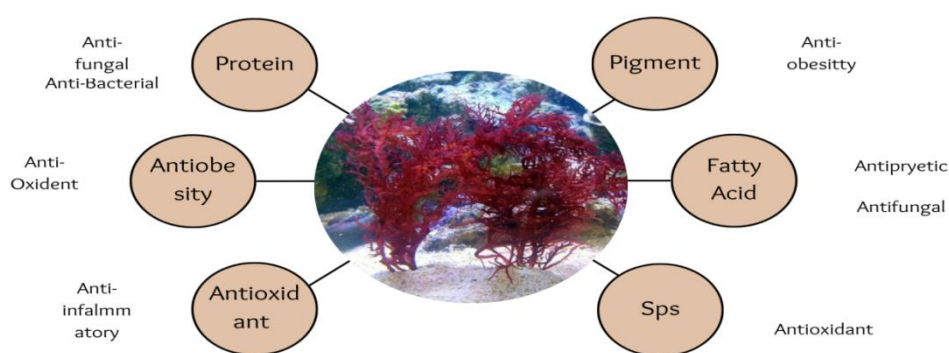


Figure 3: Role of red macro-algae.

Eucheuma and *Kappaphycus* are important red seaweed, also collectively known as eucheumatoids produced highly due to universal demand (Buschmann *et al.*, 2017). From seaweed carrageenan is produced but it is not adapted by humans, with no nutritional value and low fiber, it is just used for thicken food products (Imeson, 2009). Carrageenan is similar to the carboxymethyl cellulose (CMC) it is used as binder in toothpaste (Bixler and Porse, 2011). Among all three phyla high digestibility seen in the red seaweed (Tibbetts *et al.*, 2016), compare the values with various plants, some grains (69–84%), fruits (72–92%), vegetables (68–80%) and legumes (72–92%) (Tibbetts *et al.*, 2016; Bleakley and Hayes, 2017). 17% of digestibility recorded in *U. pinnatifida* (brown) 66.6% when using pepsin (acidic pH, 37 °C) and pancreatin (pH 7.6, 37 °C), respectively (Fujiwara *et al.*, 1984).

Red macroalgae are typically characterized by flavonoids and phenolic acids among compounds of phenols. Additionally compound of phenol (bromophenol and phlorotannins)

sole to sources of marine in less quantity and have good antioxidant activity (Olsen *et al.*, 2013; Cotas *et al.*, 2020a; Dong *et al.*, 2021). It contains anti-inflammatory anti-oxidant and anti-aids property (Choi *et al.*, 2018; Paudel *et al.*, 2019). Major terpenoids are named as carotenoids that seen in red macroalgae that contribute to distinct pigmentation, that characterized by zeaxanthin, α -carotenes, β -carotenes and lutein (Holdt and Kraan, 2011; Zubia *et al.*, 2014; Kavalappa *et al.*, 2019; Cotas *et al.*, 2020b; Ávila-Román *et al.*, 2021).

Gracilaria

Two of the most widely grown seaweeds in the world are the red algae *Gracilaria* and *Gracilariopsis*, which produce about 3.8 million tons a year and are valued at approximately US \$1 billion (FAO, 2017). China and Indonesia have been the primary producers of *Gracilaria/Gracilariopsis*, accounting for 70% and 28% of global production, respectively, whereas Chile is the most producing nation in the Americas (FAO, 2014). Presently, 185 species of *Gracilaria* and 24 species of

Gracilariopsis are taxonomically recognized (Guiry, 2014). Approximately 66% of all agar is produced by *Gracilaria/Gracilariopsis* (Pereira and Yarish 2008). Currently, 24 species of *Gracilariopsis* and 185 species of *Gracilaria* are recognized taxonomically (Guiry and Guiry, 2016). There are four primary methods for cultivating *Gracilaria/Gracilariopsis* including pond tank cultures, near-shore bottom cultivation, and open water rope cultivation (Oliveira *et al.*, 2000; Sahoo and Yarish 2005; Pereira and Yarish, 2008). *Gracilaria* is mostly utilized for

agar production and as feed for abalone, while *Kappaphycus/Eucheuma* is predominantly used for carrageenan extraction. Similar to alginate that is taken from brown seaweeds, seaweed-based hydrocolloids like agar and carrageenan are utilized extensively in both the food and non-food industries. The coastal populations where *Gracilaria* and *Kappaphycus/Eucheuma* are grown also eat them as human foods (such as pickles and salads). *Porphyra* are typically consumed by humans as ingredients in soups and sushi wraps (FAO, 2018) (Table 2).

Table 2: Brown seaweed.

Country	Laminaria/Saccharina cultivation	
	Tonnes	Share of world (%)
World	12273748	100.00
1 China	10978362	89.45
2 Japan	32600	0.27
3 Spain	0.14	0.00
4 Republic of Korea	662557	5.40
5 Norway	73	0.00
6 Faroe Island	156	0.00
7 Democratic people's republic of Korea	600000	4.89

FAO 2021 Fishery and Aquaculture Statistics.

Brown seaweed

Asia produced almost all of the kelp: South Korea 6.6%, North Korea 4.4%, and China 88.3% (FAO, 2017). Traditionally used primarily for human food, kelp has recently seen a rise in use as abalone feed because of its inexpensive production costs (Hwang *et al.*, 2013). 358 taxonomic species of brown seaweed, among all genres *Sargassum* is richest one (Guiry and Guiry, 2016). Beds of *Sargassum* are

essential habitat that provides nursery, spawning and grounds of feeding for different aquatic organisms (Komatsu *et al.*, 2014) play role an important ground for the support of aquatic life and whole ecosystem, good enough habitat for mangrove forests, coral reefs and seagrass beds. In addition *Sargassum* also has potential to use as an animal feed (Kim *et al.*, 2015) and fertilizer (Williams *et al.*, 2010).

In the cell wall of brown seaweed alginates are seen with varies structures of chemical, characteristics depends on the various genera of brown seaweed. *Durvillaea*, *Laminaria*, *Ecklonia*, *Macrocystis*, *Lessonia*, *Ascophyllum* and *Sargassum spp.* are the species of brown seaweed that have alginate. Use of alginates in cosmetic, food, textile, pharmaceutical and construction industries due to its capability to be used as thickeners, emulsifiers, gel forming agent, binding agent because of its condense aqueous nature (Wiltshire *et*

al., 2015).

From brown algae get fucoidans (*Costaria costalla*, *Undaria pinnatifida*, *Ecklonia cava*, *Sargassum horney*) that hinder the spread of colon cancer cells and melanoma in human, and used as an operative agent for anti-tumor (Suganthi *et al.*, 2010; Ermakova *et al.*, 2011). The strong antioxidant activity of various macroalgae are used for decreasing the oxidative stress on cell and treat different serious diseases. These macroalgae also contain hepatoprotective, wound curative properties and anthelmintic (Sharma *et al.*, 2016) (Table 3).

Table 3: Bioactivity of brown macro-algae.

Brown Macroalgae	Compound	Bioactivity	References
<i>Ascophyllum nodosum</i>	Sodium alginate	Prebiotic	Okolie <i>et al.</i> , 2020
<i>Fucus vesiculosus</i>	Fucoidan	Anti-angiogenesis	Oliveira <i>et al.</i> , 2019
<i>Sargassum polycystum</i>	Fucoidan fraction-2	Antibacterial	Palanisamy <i>et al.</i> , 2019
<i>Sargassum glaucescens</i>	Fucoidan	Hair growth promoting	Huang <i>et al.</i> , 2022
<i>Padina pavonioca</i>	Sulphated polysaccharides	Anticancer, antioxidant	Cao <i>et al.</i> , 2016
<i>Sargassum angustifolium</i>	Fucoidan	Wound healing	Amiri <i>et al.</i> , 2023
<i>L.japonica</i>	Polysaccharides	Antiviral	Cao <i>et al.</i> , 2016
<i>Sargassum ilicifolium</i>	Fucoidan	Antioxidant (bone regeneration)	Devi <i>et al.</i> , 2022
<i>Sargassum horneri</i>	Alginic acid	Anti-inflammatory	Fernando <i>et al.</i> , 2018
<i>Sargassum fulvellum</i>	Sulphated polysaccharide	Anti-inflammatory	Wang <i>et al.</i> , 2021

Extract and uses

Different chemicals were extracted out from macro-algae and utilized in the different products of biotechnology, cosmetics and food (Carneiro-daCunha *et al.*, 2011; Singh *et al.*, 2018; Hu *et al.*, 2021). Extract of seaweed used as addition to diet of fish which can increase the lipid metabolism, growth, stress response, physical activity, and

carcass quality, disease resistance of different species of fish (Soler *et al.*, 2009; Güroy *et al.*, 2011). Microalgae also show promise in carbon sequestration, biofuels, wastewater remediation, and algae meal and oils (Khan *et al.*, 2008). Recently seaweed farms are inhibited near coast and protected area (Kim *et al.*, 2017).

As reported, algae of marine are enough in biocompounds and nutrients (Vlaisavljevi *et al.*, 2021). Phyco-colloids are naturally polysaccharides that obtain from seaweeds and have miscellaneous physicochemical features (Los Ficocoloides en la Industria, 2022). The phycocolloids found in the species named *Sargassum* are laminarin, fucoidan and alginate (Marliana *et al.*, 2018). They are known to perform various biological functions e.g neuroprotective effects (Bálas *et al.*, 2020), anti-collagenase activity, antitumor potentials and antimicrobial effects (Kalasariya *et al.*, 2021), antioxidant activities (Liu *et al.*, 2020).

Fucoidans

Fucoidans are considering as the important bioactive compound originate in *Sargassum* specie, and fucose is chief monomer. The fucoidans composition depends on various factors i.e climate condition, species and geographical area of recovery etc. Antiviral, anti-bacterial properties seen in fucoidans also contain anti-cancer and antioxidant (Marliana *et al.*, 2018).

Alginates

Aginic acid salts from alginates are the derivatives. Their main function is give structural composition to the cell wall because of its viscous nature and gel formation physiochemical properties (Bálas *et al.*, 2020). It is demonstrated from study that in the body, alginic acid averts the preoccupation of heavy metals. It is examined that derivatives of

alginate play role as a curative compound against neurodegeneration. Alginic acid is advantageous for health, play role as a dietary fiber and decrease cholesterol level (Holdt, and Kraan, 2011).

Phycocolloid

From different brown seaweed species (Saccharina, Laminaria, Fucus, Laminariaceae and Eisenia) cell wall present linear polysaccharide which is non-toxic and biodegradable named phycocolloid laminarin. This compound has ability to play anti-inflammatory, antioxidant, as prebiotics and antitumor properties (Huang *et al.*, 2022).

The industry of food is the largest industry, consumable macroalgae have the ability to form functional food form various years. Researchers made enough effort from 15 years to know their novel way to use in meat products as bioactive compound to enhance its value (Gullón *et al.*, 2020). The industries of cosmetics contain biocompound of marine that used as a, gelling agents, viscosifiers and stabilizers (Pradhan *et al.*, 2022).

Distribution

In Pakistan 234 species of macro-algae are seen in it 110 seaweed genera are reported on the coast of Balochistan, distributed 57 families widely, 33 orders, 12 classes and 6 divisions (MFF Pakistan, 2016). Variety of marine benthic seaweeds seen on the different coastal waters e.g Manora, Pacha, Cape Monze, Sandpit, Buleji, Nathiagali and Paradise Point (Rizvi *et al.*, 2001).

It is explained that the occurrence of important seaweeds was seen in Buleji and Manora where commonly 20 species were seen e.g *Halymenia porphyroides*, *S.tytopodium zonale*, *P.adina pavonica*, *Iyengaria nizamudinii*, *S. boveanum*, *Colpomenia sinuosa*, *Lobophora variegata*, *D. indica*, *Colpomenia sinuosa*, *Jania adherence*, *Stokya indica*, *Spatoglossum variable*, *P. gymnospra*, *Cystoseira indica*, *S. vulgare*, *I. stellata*, *S. filifolium*, *D. hauckiana* and *Dictyota dichotoma* were seen abundantly (Bashir *et al.*, 2023).

14 species of macro algae seen in Hawksbay, Paradise and Sandspit areas e.g *Udotea indica*, *Colpomenia sinuosa*, *Iyengaria* sp., *Padina* spp., *Padina* spp., *Jania adherence*, *Laurencia pinnatifida*, *Caulerpa taxifolia*, *Cystoseira indica*, *Dictyota indica*, *Sargassum* spp., *Gelidium pusillum* while at Mubarak village and Sonehra point the most common found was *Dictyota indica* and *Codium iyengarii*, in the area Chach Jaan Khan K.T. Bandar and at Shahbandar the dominant specie seen was *Enteromorpha flexsousa* (Bashir *et al.*, 2023)

Hameed and Ahmed (1999) figured 85 species from Bulegi and explained different micro-habitat linked with algae. It is reported by Saifullah (1973) that 48 species seen in Buleji, Karachi. 36 species of phenophyta seen at the coast of Karachi (Abbas, 2010), it is reported 60 species of algae in Karachi at side of ocean Nathiliagali, 58 species of algae were reported by Nazim *et al.* (2012) at Bulegi, Karachi. Nearly, brown seaweed has 29 genera and 90

species which identified from the coast of Karachi in Pakistan (Shameel and Tanaka, 1992; Aisha and Shameel, 2013).

Cultivation status

In 2019 normally, different five types of seaweed consider 95% present in world among all seaweed cultivation. *Saccharina* and *Laminaria* consider 34.65% of overall world production for consumption purpose, mostly in sauces, condiments and salads. From tropical algae carrageen is taken *Eucheuma* and *Kappaphycus* nearly 32.62%. *Undaria*, *Gracilaria* and *Porphyra* accounted for 7.16%, 10.32% and 8.33% (Zhang *et al.*, 2022). It is reported that in 2018 nearly 50 countries appealingly doing seaweed cultivation with representing 32.4 million tons 97.1% in average collectively cultivated and wild (FAO, 2020; Chopin and Tacon, 2021).

It is reported by financial time that the rise in global population to nearly 10 billion by 2050 (Koyande *et al.*, 2021). from terrestrial plants algae grow 10 times faster, and contain less than one tenth of land for cultivation by producing same amount of biomass. Algae growth does not content with other plants of land. It inseminates more effectively than terrestrial plants, evades intensive use of water, wasting of fertilizers, eutrophication downstream linked with advanced agriculture (Tzachor, 2019). Sea space around 4 million square kilometers would be used in the culturing of seaweed that based for the formation of biofuels to balance the

liquid fuel and its consumption globally (Kite-Powell *et al.*, 2022).

Cultivation history

Undaria spp. and *Saccharina spp.*

From last 50 years different trails for the cultivation of kelp were applied all over the world to gain good results and new methods (Bak *et al.*, 2020). Production of *Saccharina spp.* and *Undaria spp.* are increased due to high demand in the feed of Korea (Hwang *et al.*, 2013).

Neopyropia/Pyropia/Porphyra

From hundred years in Japan cultivation of *Neopyropia/Pyropia/Porphyra* is performed and it is popular in the industry of aquaculture of China, Japan and Korea (Mumford *et al.*, 1988; Pereira and Yarish, 2008). *N. haitanensis*, *N. yezoensis* and *N. tenera* are the commercial species produced (commonly in Japan, China, and Korea) among 138 species taxonomically *Porphyra*, *Pyropia* and *Neopyropia* are accepted (Guiry and Guiry, 2016). Mainly three species (*Py. Haitanensis*, *Py. Tenera* and *Py. Yezoensis*) cultivated commercially, commonly in Japan, Korea and China (99.99% of total production) (FAO, 2017). The method of culturing *Porphyra* in all these countries with some productive changes (Sahoo and Yarish, 2005; Pereira and Yarish, 2008; Pereira *et al.*, 2015). It is reported that largest blooms of macroalgae in world were originated from grown *ulva* on the bundle of *Porphyra* farm in the china Southern yellow sea (Liu *et al.*, 2009; Hu *et al.*,

2010; Zhang *et al.*, 2016; Huo *et al.*, 2016).

Kappaphycus sp. and *Eucheuma sp.*

Eucheuma sp., and *Kappaphycus sp.* has been produced in Philippines and Indonesia (FAO, 2014) major source of carrageenan (over 80% of world's carrageenan production) (Hayashi *et al.*, 2010). *Eucheuma denticulatum* and *Kappaphycus alvarezii* are mostly farmed where 30 and 6 species are accepted taxonomically of each genus (Kim *et al.*, 2017).

Methods of seaweed cultivation

10,000 seaweed species identified in world but only 145 is being cultured by humans for their texture, culinary versatility and flavor also include *Monostroma*, *Laminaria*, *Caulerpa*, *Hizikia*, *Undaria*, *Porphyra* and *Palmaria* (Baweja *et al.*, 2016). Commonly substrates are required to green seaweeds in order to attach themselves that makes slight difference in the process of cultivation in between red and green seaweed. The green seaweed of genus *caulerpa* (sea grapes) requires sand or loamy substratum to attach themselves using rhizoids and elongates and they propagate by their stolon extension (Zubia *et al.*, 2020). Many *ulva* species are stored as vegetables and its extensive production of biomass makes it viable for the cultivation on large scale. The maximized rate of growth of *ulva sp.* per day was noticed and reported at 19.2% using offshore cages with fixed tumbling

and mixing of biomass with air and water exchange (Chemodanov *et al.*, 2019).

Onshore cultivation

In 1970-1980s cultivation of *Chondrus crispus* on-land or onshore started for the extraction of carrageenan (Craigie and Shacklock, 1995). Production take place by applying closed systems (e.g., in raceways, tanks, ponds or lagoons) in it water retained under control condition to provide suspended environment and exposed light for seaweed cultivation (Hafting *et al.*, 2012; Currie, 2018).

The key benefits of land based farming are to check the prospects of adjustments used in the cultivation. Input of nutrients can be arranged precisely to enhance the creation of bioactive compounds and reduce the discharge of harmful components (Hafting *et al.*, 2015). Monitoring of outflows and inflows done easily, water of sea is impelled and set it according to cultivation needs for seaweed. Addition of nutrients done from efficient methods by forming media under control conditions (Hafting *et al.*, 2012; Reid *et al.*, 2020).

Main shortcomings of land based farming are the high input cost for infrastructure and maintenance of whole condition of farm. Availability of land with watery area is quite expensive (Hafting *et al.*, 2012). Land based cultivation of feedstock biofuel have various drawbacks, it need arable land to be unfocused from production of food, use of different fertilizers with its release

footprint of carbon (DeCicco *et al.*, 2016), which frequently use in the irrigation which is threatened in various agricultural areas (Rathmann *et al.*, 2010; Besharat *et al.*, 2020).

Offshore cultivation

Agency-Energy (ARPA-E) of the U.S. Department of Energy for the production of chemicals, fuels, and feeds. Among the possible species mentioned are *Sargassum spp.* in the Gulf of Mexico and the Caribbean, *Saccharina* in the Northeast (Western Atlantic Ocean), and Northwest (eastern Pacific Ocean of Washington and Alaska) (Kim *et al.*, 2017).

The farming of macroalgae for use in the products commercially is not productive in system of pond due its cost above the normal range and the seaweed produced from all these systems used in the formation of high quality products (Hafting *et al.*, 2015). Offshore production of seaweed is applied in enough area, around the shore space containing farms for floating and flat cultivation of kelp (Bird, 1987), currently wild farm integrated system started and also practice ring cultivation (Buck and Buchholz, 2004).

Offshore cultivation is a challenging system consider for the growth of epiphytes (Fletcher 1995, Vairappan *et al.*, 2008). For all these reasons, species of seaweed selected to farm in open water to reduce the epiphyte growth in the whole season and prevention from local state. The change of climate with the subsequent change of water chemistry and temperature of water that

could be the reason of decreasing of the cultivation area in ocean (Troell *et al.*, 2017; Oyinlola *et al.*, 2018).

The previous offshore farming is not still suitable for doing in open area and in deep water, the important aquaculture techniques installed in the protected areas. Generally the recent offshore and onshore farming are not seem good for environmental conditions, they consider unstable economically, fluctuations seen in their production because of biotic and abiotic factors (Sulaiman *et al.*, 2012; Peteiro *et al.*, 2016; Buschmann *et al.*, 2017).

Nearshore cultivation

The cultivation near-shore is an important and mostly applied technique for seaweed cultivation, which were started in estuaries near shore areas (Soto and Wurmman, 2019). This system is also beneficial for not creating hurdles for arable area cultivation, from this it will be protected from the damages promoted from sea storms and agitation of sea. Advantage of this technique is the facilitation provided for bioremediation of river bowls that were polluted from human activities derived from agriculture activities (Zheng *et al.*, 2019). As compared to the onshore and offshore production it is not much costly and laborious (Grote, 2019).

IMTA Cultivation

The ancient way of singular cultivation of seaweed is now modified by a combined system named IMTA (integrated multitropic aquaculture) to

solve the environmental issues of aquaculture animal e.g, water eutrophication due to excretion and supplementary feed (Granada *et al.*, 2016). The integrated multitropic aquaculture is a system used for rearing and culturing different species of various tropic levels close to each other. The waste products inorganic and organic are used and reused in the system provide as a nutrient to other factors (Knowler *et al.*, 2020). In integrated multitropic aquaculture (IMTA) technique the excreta (as nutrient) taken from mollusk and fish contain phosphate and dissolved ammonia, from stabilizing the oxygen level, CO₂ and PH convert these waste into valuable biomass (Fernand *et al.*, 2017; Zheng *et al.*, 2019; Knowler *et al.*, 2020; Tanaka *et al.*, 2020).

Integrated farming

In the cultivation of shrimps, litter remains are the main problem seen that cause toxicity in aquatic environment. In intensive farming of shrimps, the level of nitrogen is high in water which causes somehow problems for the immunity of shrimps. So for balancing the conditions of capacity and yield of stock need to balance the waste assimilation in environment (Neori *et al.*, 2004).

Seaweed used as bioremediators in biofloc system to treat the waste formed from organisms. Different species of Gracilaria present naturally in seaside area of Brazil that used in the effluent of shrimps. Use seaweed as biomediator, the advanced eutrophic condition of farming ponds (Samocha *et al.*, 2015).

Shrimp are exposed to disease easily bacterial and viral, like *V. parahaemolyticus* and WSSV (White Spot Virus Syndrome). This combined farming might be suitable for controlling the pathogenic organisms (Brito *et al.*, 2016).

Rate of survival is a main issue seen in farming of shrimp. An experiment performed on diseased shrimp with bacterial pathogen, that culture with seaweed show good survival and immunity against disease control. These macroalgae contain antiviral and antibacterial capacity beside immuno-stimulating characteristics that can alleviate its survival chances (Thanigaivel *et al.*, 2016). In integrated culture of shrimp and seaweed, shrimp take seaweed as supplementary feed. From this enhance enzymatic activity and immunity. Due to this reduce oxidative stress because of antioxidant and bioactive compound taken from macroalgae and reduce pathogenic activities. The bioactive compounds are phenolic, sulfated, antioxidant and polysaccharides (Anaya *et al.*, 2019).

Role of women in culture

In seaweed agriculture and the value chain, women frequently hold important leadership positions (Msuya, 2013). Seaweed farming was first and primarily adopted by women in India because it provided them with a safe and secure source of income (Krishnan and Narayanakumar, 2013). Women in the United Republic of Tanzania are leaders in seaweed cultivation and value addition, having taken the initiative in

this field (Msuya, 2013). Women were heavily involved in seaweed farming in the Philippines, particularly in seeding and post-harvest treatments; they made up roughly 44% of the regular seaweed farming workforce and were the primary source of casual labor (Hurtado, 2013).

Growing seaweed significantly promotes women's empowerment and community cohesion (Valderrama *et al.*, 2015; Suyo *et al.*, 2020; Suyo *et al.*, 2021). Many homes with limited resources or vulnerable people can participate in seaweed production due to its labor-intensive, low-capital, and easy farming technology. This is especially true for the tropical species *Kappaphycus/Eucheuma* (Needham and Lentisco, 2013).

Maintainable culture of macroalgae to contribute in the creation of jobs and for the well-being of society, firstly i.e hatcheries, processing and grow-out operations secondly supplying goods from industries, provide services to mariculture such as equipment and feed, thirdly providing the jobs associated with it i.e directly or indirectly employment in the culture of seaweed (Pinfold, 2013).

Uses

In 20 years, the bioactive compounds from micro and macroalgae are commonly used in the industry of cosmetic. Algae as compared to terrestrial crops contain various new and exclusive components e.g terpenoid, polyphenols, sterol, halogen, polysaccharides and unsaturated fatty acids additionally protein, vitamin, trace

element and minerals (López-Hortas *et al.*, 2021). Seaweed is used to enhance beauty. Many countries in South East Asia, including Japan, China, Korea, Malaysia, Thailand, Indonesia, and the Philippines, use seaweed because of its high protein content. Soups, salads, and curries are prepared using seaweeds such as *Ulva* sp., *Enteromorpha* sp., *Caulerpa* sp., *Codium* sp., *Monostroma* sp., *Sargassum* sp., *Hydroclathrus* sp., *Laminaria* sp., *Undaria* sp., *Macrocystis* sp., *Porphyra* sp., *Gracilaria* sp., *Eucheuma* sp., *Laurencia* sp., and *Acanthophora* sp. (Kolanjinathan *et al.*, 2014).

Seaweed cultivation role in economics

Seaweed's upstream and downstream operations have benefited various sectors by fostering innovation and boosting economic development (Chen *et al.*, 2020; Felaco *et al.*, 2020; Padam and Chye, 2020; Wang *et al.*, 2020). Additionally, seaweed offers vital resources such as food, renewable energy, and raw materials, and contributes to cultural value through recreation, learning, heritage conservation, and scientific exploration (Hasselström *et al.*, 2018).

Around 221 species of seaweed in which Chlorophytes 32, Phaeophytes 64 and Rhodophytes 125 that are using in development at commercial level among all these 24 utilize in medicinal products, 25 species used in animal feed, compost and in agriculture, 145 species are used as a food (White, 2015). Macroalgae are used as compost to enhance fertility of soil and better production of plants.

Seaweed also used as an organic fertilizer in farming recompenses the shortage of plant nutrients e.g phosphorous, nitrogen and potassium (Soares *et al.*, 2020).

Seaweed also used as enhancer of beauty due to good amount of protein and used in various areas e.g China, Japan, Korea, Indonesia, Thailand, South east Asia and Philippines. Seaweeds like *Enteromorpha* sp., *Codium* sp., *Sargassum* sp., *Laminaria* sp., *Macrocystis* sp., *Gracilaria* sp., *Acanthophora* sp. *Ulva* sp., *Caulerpa* sp., *Monostroma* sp., *Hydroclathrus* sp., *Undaria* sp., *Porphyra* sp., *Laurencia* sp. and *Eucheuma* sp. are used in the formation of curry, salad and soup (Kolanjinathan *et al.*, 2014) (Fig. 4).

Used in animal food

Seaweed also used as a food to the animals of farm e.g poultry, cattle etc. Macro-algae provide disease resistance and contain balanced amount of micronutrients. They also help in reducing the cow fever and mastitis. It also enhances iodine content and fat amount in milk and their products. Seaweed enhances yolk color of eggs and aids in increasing fertility and rate births in animals (CMFRI, 2010). From species *Sargassum*, *Gelidiella*, *Gracilaria* and *Hypnea* feed were prepared and used in culturing of prawns and fishes. The feed was balanced amino acids, minerals, carbohydrates that can also maintain quality of water in aquaculture (Chapman 2012).

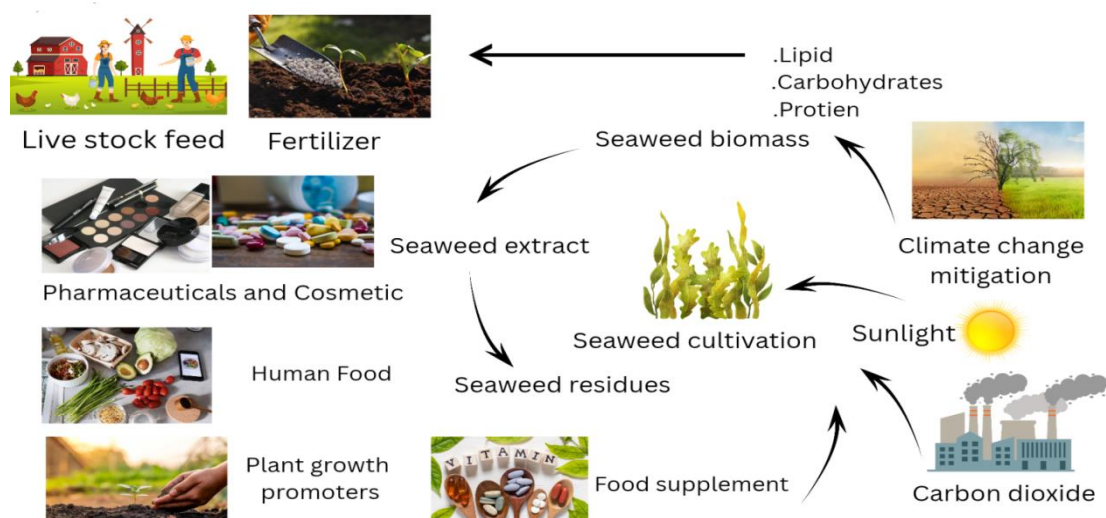


Figure 4: Various uses of seaweed.

Findings confirmed that from cattle decreasing emission of methane when give seaweed feed (Li *et al.*, 2016; Duarte *et al.*, 2017). Supplementation of seaweed in poultry, aquaculture feed and livestock were given from decades and show good results in meat quality, health of animal (Vijn *et al.*, 2020). In case of addition of brown seaweed (*Ascophyllum nodosum* and *Undaria pinnatifida*) in the feed of sheep and pigs to enhance the health of animal's intestine and with the use of some percent red seaweed (*Eucheuma denticulatum*) in fishmeal enhance quality of meat of Japanese flounders by extending the omega-3 fatty acid in muscles (Shimazu *et al.*, 2019).

del Olmo *et al.* (2018), examined a noteworthy development in physiochemical eminence of hard cheese when given as a supplementary with macroalgae e.g *Undaria pinnatifida*, *Laminaria ochroleuca*, *Porphyra umbilicalis*, *Ulva lactuca*, and *Himanthalia elongate* species. It is founded from research that microbiota

of intestine were enhance in mice because of the use of polysaccharides of seaweed (*Ulva prolifera* and *Porphyra haitanensis*) as oral supplementary feed (Zhang *et al.*, 2022)

Food additive

Due to high level of vitamins minerals and protein in seaweed it is highly used as food for long time mainly in East Asia (Sho, 2001). They are also good source of animal feed and as supplements. *A. nodosum* brown seaweed was used in the feed of animals by the Canadian company to enhance the microbial invasion and immunity (Allen *et al.*, 2001; Saker *et al.*, 2001).

More than hundred seaweed species are used by Japanese. *A. nodosum* is used in the diet of people who are overweight to decrease the energy intake after feasting (Hall *et al.*, 2012). From species of *porphyra* derived R-phycoerythrin (Suganya *et al.*, 2016) also used as a food of color (Dumay *et al.*, 2014).

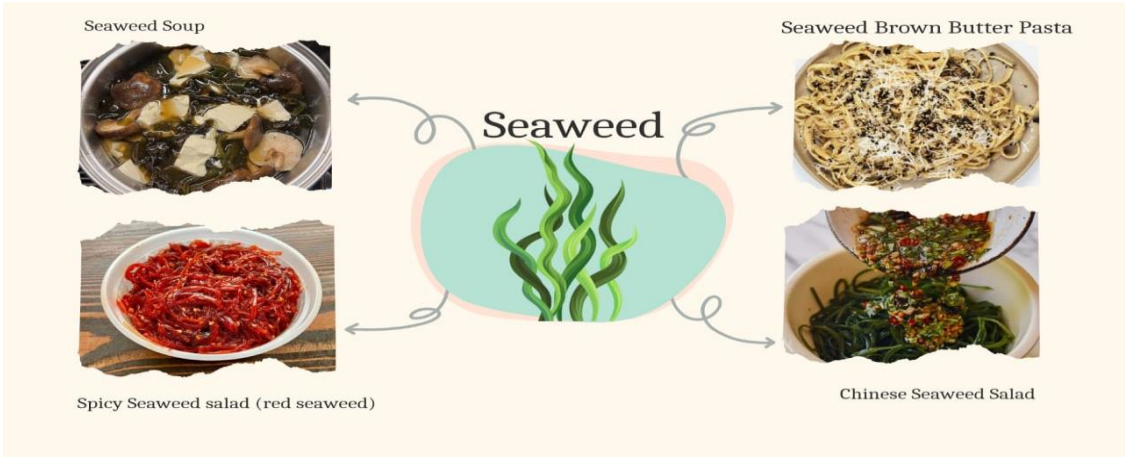


Figure 5: Uses in food.

High value macroalgae product is phycobiliproteins (approximately \$5000/g) (Suganya *et al.*, 2016). Although their lower constancy in light and heat, phycobiliproteins also play role in cosmetics (phycoerythrin and phycocyanin) and food natural colorant (Griffiths *et al.* 2016). Seaweed has enough fiber content, which can complicate the digestion phenomenon in humans, while ruminants have designed enzymes in their system that are helpful in their digestion and enhance the accessibility of nutrients (Holman and Malau, 2013) (Table 4).

Table 4: Edilble species of macro-algae.

Edible species of Seaweeds		
Brown Algae	Green Algae	Red Algae
<i>Durvillaea Antarctica</i>	<i>Caulerpa</i> spp	<i>Chondrus crispus</i>
<i>Alaria esculenta</i>	<i>Codium</i> spp	<i>Eucheuma denticulatum</i>
<i>Eisenia bicyetis</i>	<i>Enteromorpha</i> spp	<i>Gracitaria edutis</i>
<i>Ascophyllum nodosum</i>	<i>Ulva lactuca</i>	<i>Champia compressa</i>
<i>Fucus vesiculosus</i>	<i>Ulva</i> spp	<i>Gelidiella acerosa</i>
<i>Fucus serratus</i>	<i>Ulva pertusa</i>	<i>Porphyra</i> spp
<i>Laminaria hyperborean</i>	<i>Monostroma</i> spp	<i>Porphyra columbina</i>
<i>Laminaria digitata</i>	<i>Ulva australis</i>	<i>Porphyra umbiticitis</i>
<i>Himanthatia elongate</i>	<i>Entromorpha</i> spp	<i>Porphyra laciniata</i>
<i>Sargassum swartzii</i>		<i>Gracilariopsis tongissima</i>
<i>Sargassum fusiforme</i>		<i>Palmaria palmate</i>
<i>Sarrgassum vulgare</i>		<i>Osmundea pinnatifida</i>
<i>Sarrgassum muticum</i>		<i>Mastocarpus stellatus</i>
<i>Undaria undarioides</i>		<i>Solieria robusta</i>
<i>Undaria pinnatifida</i>		
<i>Stoechospermum marginatum</i>		

Fleurence, 2022; Pandey *et al.*, 2020

Heavy metals absorption

From seawater absorb heavy metals that depends on different factors e.g season, species, wave exposure, location, light intensity, temperature, PH, age of the plant, nitrogen availability and salinity (Griffiths *et al.*, 2016). Chen *et al.* (2018), explain the metalloids and metal Se, Mn, As, Hg, Al, Cr, Ni, Cu, Cd, and Pb that are absorb by red seaweeds. Mainly the concentrations of heavy metal in seaweed are low than preserving toxic limit. But the bioaccumulation level of lead, cadmium and arsenic are consider the culture of seaweed hazardous that can cause hyperpigmentation, allergies and cancer. Arsenic is consider as hazardous chemical compound carcinogenic in nature, it is taken as food by seaweeds and it is seen high level of arsenic present in seaweed as in organic form (Taylor *et al.*, 2017). In coastal defense aquatic seaweed play role by decreasing the hydrodynamics energy of waves by normalizing high bed tides, and protecting the area of tides from erosion (Christianen *et al.*, 2013, Ondiviela *et al.*, 2014).

The photosynthetic activity of seaweeds and microalgae can clean wastewater, lessen ocean acidification, reduce eutrophication, and capture carbon dioxide by removing nutrients (phosphorus and nitrogen) from nearby waters and absorbing carbon dioxide (Muraoka, 2004).

Pharmaceutical use

Natural commercially formed product named InSea2 available in different

countries which is used to control metabolism of carbohydrates and blood sugar. The product InSea2 is formed from two seaweeds natural extracts *A. nodosum* and *F. vesiculosus*. These extracts have inhibitory factors called α -glucosidase and α -amylase (Roy *et al.*, 2011).

Alginate is used to enhance the texture of paper. It is being used as a reactive foundation for textile reactive dye printing, a stabilizer for ice cream, and a possible additive for frozen foods. Alginates are also frequently employed as stabilizers in the pharmaceutical industry (Krishnamurthy 2005). Approximately 13000 years, marine macro algae were subjugated by anthropogenic activities in the field of food and medicine. In Chile at late Pleistocene settlement findings of seaweed based presence seen at Jomon period and Monte Verde in Japan (Ugent and Tindall, 1997), an archeological implication from earlier sites macro algae used in three different purpose of medical, from 400BC Ayurvedic medicine formed (Misra and Sinha, 1979) (Fig. 6).

Anti inflammatory activity

Methanol extract taken from *Ulva linza* and *Undaria pinnatifida* used due their anti-inflammation function, have actively participated in the resistant of inflammation while used against mouse erythema and ear edema. Edema was inactive by the species of macro-algae *Ulva linza* and *Undaria pinnatifida* (Mohammed *et al.*, 2008).

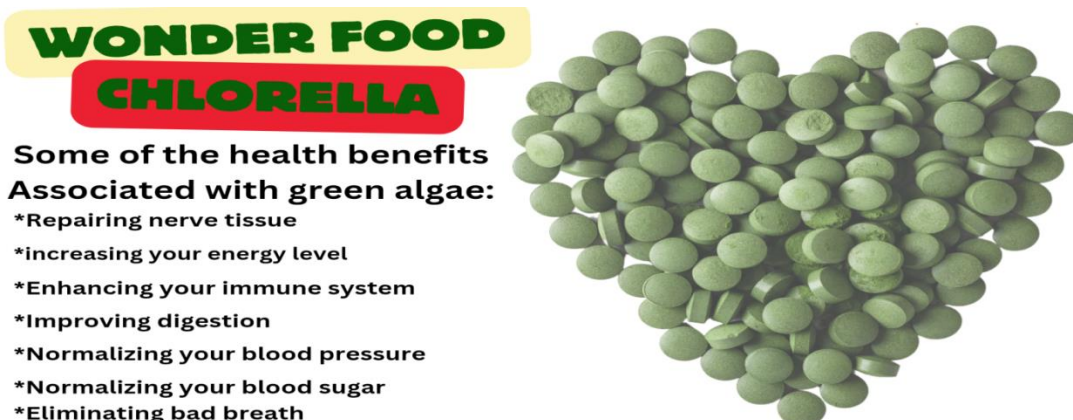


Figure 6: *Chlorella*.

During in-vivo study extract of methanol from *Ulva linza* and *Undaria Pinnatifida* seaweeds which play role of anti-inflammation (Khan *et al.*, 2008). Extract of methane from Dictyota dichotoma was consider as noteworthy against β -lactamases by constraining GES-22 which can be effective in controlling the emergence condition of antibiotic resistance of bacteria in humans (Houchi *et al.*, 2018)

Bioactive compounds

Seaweeds are known for their natural richness in especial sulfated polysaccharides, bioactive molecules such as polyunsaturated fatty acids, proteins, lipids, natural pigments, carbohydrates, and minerals such as iron, potassium, sulfur, iodine, vitamins (Plaza *et al.*, 2008; Kraan, 2013). Different studies noticed that various seaweed contain good amount of polyphenols that are antioxidant in nature (Gómez *et al.*, 2018).

In red seaweed found carrageenans which are composed of D-series of 4-linked α -galactose residues. Polysaccharides called agarans are

extracted from red seaweed, which have L-series of 4-linked α -galactose remains (Knutsen *et al.*, 1994). These are the molecules contain broad spectrum of antiviral, anticancer activities and anticoagulant (Jiao *et al.*, 2011; Mayer *et al.*, 2011).

Effect on climate and soil

The autotrophic seaweed mass production can help in enhancing the effect in change of climate. Seaweeds are considered as a good carbon sinks which help in absorbing the carbon dioxide from the environment to avert the atmosphere from acidification (Fernández *et al.*, 2019). *Sargassum*, *Ascomphyllum*, *Laminaria* are the species used as organic manure, which is by nature non-toxic, non-hazardous, biodegradable and non-polluting to birds animals and humans. Apart from this it enhance moisture holding ability and fertility of soil (Pati *et al.*, 2016).

Seaweed enhance the soil nutritional profile e.g K, P, N and all the nutrients necessary for growth of plant (Nabti *et al.*, 2017). In seaweed the elements composition seen in the species of

Karachi coast are Zn, Pb, Na, Cd, Ca, Cu, Cr, Mg, Fe, k and Co. Most abundant elements are K, Mg, Ca, Na and Fe (Rizvi *et al.*, 2001).

Use in Cosmetics

Pakistan is known for different emporium of seaweed that is used in cosmetics. Shameel and Tanaka (1992) reported 177 genera and 475 species of benthic algae and marine planktonic present the aquatic environment. Among all seaweed, various used in the products of cosmetics formation. Commonly seen macro alga at the coast of Pakistan are *Gracilaria gracilis*, *Hypnea musciformis*, *Laurencia obtusa*, *Gelidium usmanghanii*, *Scinaia saifullahii*, *Gracilaria corticata*, *Botryocladia leptopoda* that used in the formation cosmetics products by exploitation of agar (Afaq-Husain *et al.*, 2001).

Biofuel production

By Using and applying seaweed in the industries of agriculture and energy. Convert seaweed into biofuel e.g biodiesel and bio-alcohols (butanol and ethanol) by adopting the process of microbial fermentation and the breakdown of phytochemical components (Ashokkumar *et al.*, 2017; Ra *et al.*, 2019). Biofuel yeild from different seaweeds species, after the fermentation of various microorganisms, by using different technology which can help in the efficient conversion that would be more effective for the production of biofuels on commercial scale (Tabassum *et al.*, 2017; Ra *et al.*,

2019). *Ulva sp.* the green seaweed used to produce polylactic acid from feedstock (Helmes *et al.*, 2018).

Use to cure Cancer

The species of seaweed *Fucus spp* in brown algae has shown resistant against breast cancer and colorectal. Seaweed has various anticancer effects on breast and colon of human (Moussavou *et al.*, 2014). In past, Chinese used *Laminaria sp.* for cancer treatment (Loeser, 1956). It is documented that various seaweed has anticancer effect against breast and colon cancer in human beings. Different macroalgae play defensive role from cancer by decreasing the formation of cancer cells. Due to good anti-oxidant ability of macroalgae as a defensive agent against heart diseases (Sharma *et al.*, 2016). Macroalgae also used against diabetes. Administration of extract of *Ulva faciata* orally that decrease the glycosylated hemoglobin level and blood glucose as compared to other in-vivo standard of medicines (Abirami and Kowsalya, 2013).

Reproductive function

Macro-algae raise the amount of fat and iodine in milk and milk products and it improves the yolk color of eggs and increases animal fertility and birth rates. Fish and prawn culture are fed the food made from the species of *Sargassum*, *Gracilaria*, *Gelidiella*, and *Hypnea* (Gómez *et al.*, 2018). The feed's mineral, amino acid, and carbohydrate enrichment helps to maintain aquaculture's water quality (Kaladharan *et al.*, 1998). In aquaculture system it is

used to prevent the waste from the culture of shrimp, which decrease the chances of eutrophication. The *Gracilaria verrucosa* specie of red seaweed have good efficacy to eliminate level of BOD and COD whereas *Ulva fasciata* a green macroalgae have higher efficacy for the ammonia removal (Sasikumar and Rengsam, 1994).

Difficulties and challenges

Apart from benefits, many of difficulties seen in the propagation of seaweed that includes the lack of significant protocols for the gaining of callus induction, axenic cultures and the whole plant regeneration by using regulators for plant growth. For commercialization, hybridization is another technique used in the production and growth enhancement of macroalgae (Kopprio *et al.*, 2021). New hybrid strain *K. alvarezii* and *E. denticulatum* with new carrageenan composition and good growth rates was formed by using cell-cell fusion technique and fusion of protoplast (Cheney *et al.*, 1998; Bindu and Levine 2011).

External challenges

The problems of environment seen due to the evolvement of algae's processing and breeding. Halogenated hydrocarbons released from various algae that can cause effect on ultraviolet rays and on ozone. The mass of growing area is a suitable platform for the addition of new species. The equipment of breeding is the base of attachment for green macroalgae. Within the rise of

temperature the production rate of green seaweed increases that will be added in ocean and cause the green tides (Yu *et al.*, 2020).

Internal challenges

Macroalgae processing and production technology does not meet to its cultivation need. Recently the production of macroalgae is mostly established in low developed areas. Some counties, mainly China know the importance of large-scale offshore production, industrial seed raising, automated harvesting and maintain industrial chain for sale different products. Most of the countries, done basic productions and their practices for processing and harvesting are common. From the value addition in the products are squat due to the use of low techniques for purification and extraction (Pérez *et al.*, 2016). In the open ocean the barriers seen for seaweed culture include insufficient technology that ropes the operational purpose and cultivation in open environment and professed possibility for aquatic (marine) mammal predicaments. Forming structures of aquaculture that can be viable economically and bear-up the hurdles of open sea area and the engineering difficulties faced for the advancement of tools, analysis and demonstration of complete system (Moscicki *et al.*, 2024).

Conclusion

Seaweeds are an increasingly significant and highly valued marine asset, with

expanding and varied uses across a wide range of industries, particularly in aquaculture. This is largely attributed to their rich nutritional content and the presence of numerous biologically active compounds. The integration of seaweed into aquaculture diets has yielded significant benefits, such as faster fish growth, strengthened immune systems, and a natural source of critical minerals and pigments. The escalating global demand for seaweed and its by-products is driving the advancement of sustainable, environmentally responsible, and efficient farming techniques. To fully tap into seaweed's potential while ensuring both ecological and economic resilience, it is imperative to focus on maximizing nutrient yield, breeding durable and climate-resilient strains, and promoting the development of integrated multi-trophic aquaculture systems.

References

- Abbas, A., 2010.** *Anatomical studies on the Phaeophycota of Karachi Coast* (Doctoral dissertation, federal urdu university of arts, science & technology, gulshan-e-iqbal campus, karachi-75300, Pakistan) <http://localhost:80/xmlui/handle/123456789/9637>
- Abirami, R.G. and Kowsalya, S., 2013.** Antidiabetic activity of *Ulva fasciata* and its impact on carbohydrate enzymes in alloxan-induced diabetic rats. *International Journal of Research in Phytochemistry and Pharmacology*, 3, 136–141. <http://pharmascope.org/ijrpp/jdownl>
- Afaq-Husain, S., Saeed, V.A. and Masood, A., 2001.** Economic seaweeds of Pakistan coast. *Pakistan Journal of Marine Biology*, 7(1and2), 281–290.
- Aisha, K. and Shameel, M., 2013.** Comparative study on four species of the order Scytosiphonales (Phaeophycota) from Karachi coast. *International Journal of Phycology and Phycochemistry*, 8(2), 149–158.
- Allen, V.G., Pond, K.R., Saker, K.E., Fontenot, J.P., Bagley, C.P., Ivy, R.L., Evans, R.R., Brown, C.P., Miller, M.F. and Montgomery, J.L., 2001.** Tasco-Forage: III. Influence of a seaweed extract on performance, monocyte immune cell response, and carcass characteristics in feedlot-finished steers. *Journal of Animal Science*, 79, 1032–1040 <https://doi.org/10.2527/2001.7941032x>
- Ambati, R. R., Phang, S. M., Ravi, S. and Aswathanarayana, R. G., 2014.** Astaxanthin: Sources, extraction, stability, biological activities and its commercial applications – a review. *Marine Drugs*, 12(1), 128–152. <https://doi.org/10.3390/md12010128>
- Amiri Goushki, M., Sabahi, H., & Kabiri, M., 2023.** In vitro evaluation of the wound healing properties and safety assessment of fucoidan extracted from *Sargassum angustifolium*. *Current Applied Science and Technology*, 23, 10-55003

- <https://doi.org/10.2527/2001.7941032x>
- Anaya-Rosas, R. E., Rivas-Vega, M. E., Miranda-Baeza, A., Piña-Valdez, P., & Nieves-Soto, M., 2019.** Effects of a co-culture of marine algae and shrimp (*Litopenaeus vannamei*) on the growth, survival and immune response of shrimp infected with *Vibrio parahaemolyticus* and white spot virus (WSSV). *Fish & Shellfish Immunology*, 87, 136–143 <https://doi.org/10.1016/j.fsi.2018.12.071>
- Ashokkumar, V., Salim, M. R., Salam, Z., Sivakumar, P., Chong, C. T., Elumalai, S., Suresh, V., & Ani, F. N., 2017.** Production of liquid biofuels (biodiesel and bioethanol) from brown marine macroalgae *Padina tetrastrum*. *Energy Conversion and Management*, 135, 351–361 <https://doi.org/10.1016/j.enconman.2016.12.054>
- Astorga-España, M. S., Rodríguez-Galdón, B., Rodríguez-Rodríguez, E. M., & Díaz-Romero, C. (2016).** Amino acid content in seaweeds from the Magellan Straits (Chile). *Journal of Food Composition and Analysis*, 53, 77–84 <https://doi.org/10.1016/j.jfca.2016.09.004>
- Ávila-Román, J., García-Gil, S., Rodríguez-Luna, A., Motilva, V., & Talero, E., 2021.** Anti-inflammatory and anticancer effects of microalgal carotenoids. *Marine Drugs*, 19(10), 531. <https://doi.org/10.3390/md19100531>
- Bak, U. G., Gregersen, Ó., & Infante, J., 2020.** Technical challenges for offshore cultivation of kelp species: Lessons learned and future directions. *Botanica Marina*, 63, 341–353 <https://doi.org/10.1515/bot-2019-0005>
- Barbier, M., Charrier, B., Araujo, R., Holdt, S., Jacquemin, B., Rebours, C., & Wichard, T. (2018). PEGASUS: Phycomorph european guidelines for sustainable aquaculture of seaweeds. *Centro Oceanográfico de Santander*.
- Bashir, F., Shaukat, S. S., Abbas, A., SIDDIQUI, M. F., & Qureshi, I. A., 2025.** Multivariate analysis of marine algal assemblages and their distribution pattern along the Sindh coastal area of Pakistan. *Pak. J. Bot*, 57(4), 1515–1528
- Baweja, P., Kumar, S., Sahoo, D., & Levine, I., 2016.** Biology of seaweeds. In J. Fleurence & I. Levine (Eds.), *Seaweed in health and disease prevention* (pp. 41–106). Academic Press. <https://doi.org/10.1016/B978-0-12-802772-1.00003-3>
- Besharat, S., Barão, L., & Cruz, C., 2020.** New strategies to overcome water limitation in cultivated maize: Results from sub-surface irrigation and silicon fertilization. *Journal of Environmental Management*, 263, 110398. <https://doi.org/10.1016/j.jenvman.2020.110398>

- Bikker, P., Stokvis, L., Van Krimpen, M. M., Van Wikselaar, P. G., & Cone, J. W., 2020.** Evaluation of seaweeds from marine waters in Northwestern Europe for application in animal nutrition. *Animal Feed Science and Technology*, 263, 114460.
<https://doi.org/10.1016/j.anifeedsci.2020.114460>
- Bindu, M. S., & Levine, I. A., 2011.** The commercial red seaweed *Kappaphycus alvarezii*—An overview on farming and environment. *Journal of Applied Phycology*, 23, 789–796.
<https://doi.org/10.1007/s10811-010-9568-4>
- Bird, K. T., 1987.** Cost analyses of energy from marine biomass. *Developments in Aquaculture and Fisheries Science*, 16, 327–350.
[https://doi.org/10.1016/S0167-9309\(08\)70162-5](https://doi.org/10.1016/S0167-9309(08)70162-5)
- Bixler, H. J., & Porse, H. (2011).** A decade of change in the seaweed hydrocolloids industry. *Journal of Applied Phycology*, 23, 321–335.
<https://doi.org/10.1007/s10811-010-9529-3>
- Bleakley, S., & Hayes, M., 2017.** Algal proteins: Extraction, application, and challenges concerning production. *Foods*, 6(5), 33.
<https://doi.org/10.3390/foods6050033>
- Botkin, D. B., Saxe, H., Araujo, M. B., Betts, R., Bradshaw, R. H., Cedhagen, T., Chesson, P., Dawson, T. P., Etterson, J. R., & Faith, D. P., 2007.** Forecasting the effects of global warming on biodiversity. *Bioscience*, 57(3), 227–236.
<https://doi.org/10.1641/B570306>
- Brito, L. O., Chagas, A. M., da Silva, E. P., Soares, R. B., Severi, W., & Gálvez, A. O., 2016.** Water quality, *Vibrio* density and growth of Pacific white shrimp *Litopenaeus vannamei* (Boone) in an integrated biofloc system with red seaweed *Gracilaria birdiae* (Greville). *Aquaculture Research*, 47(3), 940–950.
<https://doi.org/10.1111/are.12551>
- Buck, B. H., & Buchholz, C. M., 2004.** The offshore-ring: A new system design for the open ocean aquaculture of macroalgae. *Journal of Applied Phycology*, 16(4), 355–368.
<https://doi.org/10.1023/B:JAPH.0000047941.96231.ea>
- Burrows, M. T., Smale, D., O'Connor, N., Van Rein, H., & Moore, P., 2014.** *Marine Strategy Framework Directive indicators for UK kelp habitats Part 1: Developing proposals for potential indicators.* Joint Nature Conservation Committee
<http://jncc.defra.gov.uk/page-6820>
- Buschmann, A. H., Camus, C., Infante, J., Neori, A., Israel, A., Hernández-González, M. C., Pereda, S. V., Gomez-Pinchetti, J. L., Golberg, A., Tadmor-Shalev, N., 2017.** Seaweed production: Overview of the global state of exploitation, farming and emerging research activity. *European Journal of Phycology*, 52(4), 391–406.

- <https://doi.org/10.1080/09670262.2017.1365175>
- Cao, S., Yang, Y., Liu, S., Shao, Z., Chu, X., & Mao, W., 2022.** Immunomodulatory activity *in vitro* and *in vivo* of a sulfated polysaccharide with novel structure from the green alga *Ulva conglobata* Kjellman. *Marine Drugs*, 20(7), 447. <https://doi.org/10.3390/md20070447>
- Cao, Y. G., Hao, Y., Li, Z. H., Liu, S. T., & Wang, L. X., 2016.** Antiviral activity of polysaccharide extract from *Laminaria japonica* against respiratory syncytial virus. *Biomedicine & Pharmacotherapy*, 84, 1705–1710. <https://doi.org/10.1016/j.biopha.2016.10.014>
- Carl, C., de Nys, R., Lawton, R. J., & Paul, N. A., 2014.** Methods for the induction of reproduction in a tropical species of filamentous *Ulva*. *PLoS ONE*, 9(5), e97396. <https://doi.org/10.1371/journal.pone.0097396>
- Carneiro-da-Cunha, M. G., Cerqueira, M. A., Souza, B. W., Teixeira, J. A., & Vicente, A. A., 2011.** Influence of concentration, ionic strength and pH on zeta potential and mean hydrodynamic diameter of edible polysaccharide solutions envisaged for multi-nano-layered films production. *Carbohydrate Polymers*, 85(3), 522–528. <https://doi.org/10.1016/j.carbpol.2011.03.026>
- Chakraborty, S., & Santra, S. C., 2008.** Biochemical composition of eight benthic algae collected from Sunderban. *Indian Journal of Marine Sciences*, 37, 329–332. <http://jncc.defra.gov.uk/page-6820>
- Chapman, V., 2012.** *Seaweeds and their uses*. Springer Science & Business Media. 85P.
- Chemodanov, A., Robin, A., Jinjikhashvily, G., Yitzhak, D., Liberzon, A., Israel, A., & Golberg, A., 2019.** Feasibility study of *Ulva* sp. (Chlorophyta) intensive cultivation in a coastal area of the Eastern Mediterranean Sea. *Biofuels, Bioproducts and Biorefining*, 13(4), 864–877. <https://doi.org/10.1002/bbb.1999>
- Chen, Q., Pan, X. D., Huang, B. F., & Han, J. L., 2018.** Distribution of metals and metalloids in dried seaweeds and health risk to population in southeastern China. *Scientific Reports*, 8, 3578. <https://doi.org/10.1038/s41598-018-21918-6>
- Chen, Y., Li, J., Huang, Z., Su, G., Li, X., Sun, Z., & Qin, Y., 2020.** Impact of short-term application of seaweed fertilizer on bacterial diversity and community structure, soil nitrogen contents, and plant growth in maize rhizosphere soil. *Folia Microbiologica*, 65, 591–603. <https://doi.org/10.1007/s12223-020-00768-5>
- Chen, Y., Ouyang, Y., Chen, X., Chen, R., Ruan, Q., Farag, M. A., Chen, X., & Zhao, C., 2022.**

- Hypoglycaemic and anti-ageing activities of green alga *Ulva lactuca* polysaccharide via gut microbiota in ageing-associated diabetic mice. *International Journal of Biological Macromolecules*, 212, 97–110. <https://doi.org/10.1016/j.ijbiomac.2022.05.109>
- Cheney, D., Rudolph, B., Wang, L. Z., Metz, B., Watson, K., Roberts, K., & Levine, I., 1998.** Genetic manipulation and strain improvement in commercially valuable red seaweeds. In Y. Le Gal & H. O. Halvorson (Eds.), *New developments in marine biotechnology* (pp. 101–104). Springer. https://doi.org/10.1007/978-1-4615-5705-2_10
- Choi, Y. K., Ye, B. R., Kim, E. A., Kim, J., Kim, M. S., Lee, W. W., Ahn, G. N., Kang, N., Jung, W. K., & Heo, S. J., 2018.** Bis (3-bromo-4,5-dihydroxybenzyl) ether, a novel bromophenol from the marine red alga *Polysiphonia morrowii* that suppresses LPS-induced inflammatory response by inhibiting ROS-mediated ERK signaling pathway in RAW 264.7 macrophages. *Biomedicine & Pharmacotherapy*, 103, 1170–1177. <https://doi.org/10.1016/j.biopha.2018.04.121>
- Chopin, T., & Tacon, A. G. J., 2021.** Importance of seaweeds and extractive species in global aquaculture production. *Reviews in Fisheries Science & Aquaculture*, 29, 139–148. <https://doi.org/10.1080/23308249.2020.1810626>
- Christianen, M. J. A., Van Belzen, J., Herman, P. M. J., Van Katwijk, M. M., Lamers, L. P. M., Van Leent, P. J. M., & Bouma, T. J., 2013.** Low-canopy seagrass beds still provide important coastal protection services. *PLoS ONE*, 8(5), e62413. <https://doi.org/10.1371/journal.pone.0062413>
- Christie, H., Jørgensen, N. M., Norderhaug, K. M., & Waage-Nielsen, E., 2003.** Species distribution and habitat exploitation of fauna associated with kelp (*Laminaria hyperborea*) along the Norwegian coast. *Journal of the Marine Biological Association of the United Kingdom*, 83(4), 687–699. <https://doi.org/10.1017/S0025315403007658h>
- Cian, R. E., Caballero, M. S., Sabbag, N., González, R. J., & Drago, S. R., 2014.** Bio-accessibility of bioactive compounds (ACE inhibitors and antioxidants) from extruded maize products added with a red seaweed *Porphyra columbina*. *LWT - Food Science and Technology*, 55, 51–58. <https://doi.org/10.1016/j.lwt.2013.09.010>
- Ciaccia, M., Quintana, I., Vizcargüénaga, M. I., Cerezo, A. S., & Damonte, E. B., 2007.** Polysaccharides from the green seaweeds *Codium fragile* and *C. vermilara* with controversial effects on hemostasis. *International Journal of Biological Macromolecules*, 41(5), 641–649.

- <https://doi.org/10.1016/j.ijbiomac.2007.08.007>
- CMFRI., 2010.** *Socio-economic dimensions of seaweed farming in India* (Special Publication, p. 104). Central Marine Fisheries Research Institute
<https://doi.org/10.3390/md18080384>
- Cotas, J., Leandro, A., Monteiro, P., Pacheco, D., Figueirinha, A., Gonçalves, A., da Silva, G. J., & Pereira, L., 2020a.** Seaweed phenolics: From extraction to applications. *Marine Drugs*, 18(8), 384. <https://doi.org/10.3390/md18080384>
- Cotas, J., Leandro, A., Pacheco, D., Gonçalves, A. and Pereira, L., 2020b.** A comprehensive review of the nutraceutical and therapeutic applications of red seaweeds (*Rhodophyta*. *Life*, 10(3), 19. <https://doi.org/10.3390/life10030019>
- Craigie, J.S. and Shacklock, P.F., 1995.** *Culture of Irish moss* (2nd ed. The Canadian Institute for Research on Regional Development.
- Currie, M. E., 2018.** The growing sustainable seaweed industry: A comparison of Australian state governance directing current and future seaweed cultivation. https://digitalcollections.sit.edu/isp_collection/2956
- Dawes, C., 2016.** Macroalgae systematics. In J. Fleurence and I. Levine (Eds.), *Seaweed in health and disease prevention* (pp. 107–148.
- De Schryver, A. M., Brakkee, K. W., Goedkoop, M.J. and Huijbregts, M.A., 2009.** *Characterization factors for global warming in life cycle assessment based on damages to humans and ecosystems*. ACS Publications.
- DeCicco, J. M., Liu, D. Y., Heo, J., Krishnan, R., Kurthen, A. and Wang, L., 2016.** Carbon balance effects of US biofuel production and use. *Climatic Change*, 138, 667–680. <https://doi.org/10.1007/s10584-016-1764-4>
- del Olmo, A., Lopez-Perez, O., Picon, A., Gaya, P. and Nunez, M., 2019.** Cheese supplementation with five species of edible seaweeds: Effect on proteolysis, lipolysis and volatile compounds. *International Dairy Journal*, 90, 104–113. <https://doi.org/10.1016/j.idairyj.2018.11.012>
- Dembitsky, V. M. and Maoka, T., 2007.** Allenic and cumulenenic lipids. *Progress in Lipid Research*, 46(6), 328–375. <https://doi.org/10.1016/j.plipres.2007.07.001>
- Devi, G.V.Y., Nagendra, A.H., Shenoy, P.S., Chatterjee, K. and Venkatesan, J., 2022.** Isolation and purification of fucoidan from *Sargassum ilicifolium*: Osteogenic differentiation potential in mesenchymal stem cells for bone tissue engineering. *Journal of the Taiwan Institute of Chemical Engineers*, 136, 104418. <https://doi.org/10.1016/j.jtice.2022.104418>

- Dong, H., Liu, M., Wang, L., Liu, Y., Lu, X., Stagos, D., Lin, X. and Liu, M., 2021.** Bromophenol bis (2,3,6-tribromo-4,5-dihydroxybenzyl) ether protects HaCaT skin cells from oxidative damage via Nrf2-mediated pathways. *Antioxidants*, 10(9), 1436. <https://doi.org/10.3390/antiox10091436>
- Duarte, C. M., Wu, J., Xiao, X., Bruhn, A. and Krause-Jensen, D., 2017.** Can seaweed farming play a role in climate change mitigation and adaptation? *Frontiers in Marine Science*, 4, 100. <https://doi.org/10.3389/fmars.2017.01000>
- Dumay, J., Morançais, M., Munier, M., Le Guillard, C. and Fleurence, J., 2014.** Chapter Eleven—Phycoerythrins: Valuable proteinic pigments in red seaweeds. In F. Bourgoignon (Ed.), *Advances in Botanical Research*, 71, pp. 321–343. Academic Press. <https://doi.org/10.1016/B978-0-12-408062-1.00011-1>
- El-Baz, F.K., Hussein, R.A., Mahmoud, K. and Abdo, S.M., 2018.** Cytotoxic activity of carotenoid rich fractions from *Haematococcus pluvialis* and *Dunaliella salina* microalgae and the identification of the phytoconstituents using LC-DAD/ESI-MS. *Phytotherapy Research*, 32(2), 298–304. <https://doi.org/10.1002/ptr.5976>
- Ermakova, S., Sokolova, R., Kim, S.M., Um, B.H., Isakov, B. and Zvyagintseva, T., 2011.** Fucoidans from brown seaweeds *Sargassum horneri*, *Ecklonia cava*, *Costaria costata*: Structural characteristics and anticancer activity. *Applied Biochemistry and Biotechnology*, 164, 841–850. <https://doi.org/10.1007/s12010-011-9178-2>
- FAO., 2014.** *State of World Fisheries and Aquaculture: 2014*. Food & Agriculture Organization of the UN.
- FAO., 2017.** *The state of world fisheries and aquaculture*. <http://www.fao.org/fishery/en>
- FAO., 2018.** *The global status of seaweed production, trade and utilization* (Globefish Research Program, No. 124. FAO.
- FAO., 2020.** *The state of world fisheries and aquaculture 2020: Sustainability in action*. FAO. <https://doi.org/10.4060/ca9229en>
- FAO., 2021.** Global Production Statistics 1950–2019. *FAO Fisheries Division. FishStaJ—Software for Fishery and Aquaculture Statistical Time Series*.
- Felaco, L., Olvera-Novoa, M.A. and Robledo, D., 2020.** Multitrophic integration of the tropical red seaweed *Solieria filiformis* with sea cucumbers and fish. *Aquaculture*, 527, 735475. <https://doi.org/10.1016/j.aquaculture.2020.735475>
- Fernand, F., Israel, A., Skjermo, J., Wichard, T., Timmermans, K.R. and Golberg, A., 2017.** Offshore macroalgae biomass for bioenergy production: Environmental aspects, technological achievements and

- challenges. *Renewable and Sustainable Energy Reviews*, 75, 35–45.
<https://doi.org/10.1016/j.rser.2016.10.060>
- Fernández-Segovia, I., Lerma-García, M.J., Fuentes, A. and Barat, J.M., 2018.** Characterization of Spanish powdered seaweeds: Composition, antioxidant capacity and technological properties. *Food Research International*, 111, 212–219.
<https://doi.org/10.1016/j.foodres.2018.05.054>
- Fernández, P.A., Leal, P.P. and Henríquez, L.A., 2019.** Co-culture in marine farms: Macroalgae can act as chemical refuge for shell-forming molluscs under an ocean acidification scenario. *Phycologia*, 58(6), 542–551.
<https://doi.org/10.1080/00318884.2019.1644080>
- Fernando, I.P.S., Sanjeeva, K.K.A., Samarakoon, K.W., Lee, W.W., Kim, H.S. and Jeon, Y.J., 2018.** Squalene isolated from marine macroalgae *Caulerpa racemosa* and its potent antioxidant and anti-inflammatory activities. *Journal of Food Biochemistry*, 42(5), e12628.
<https://doi.org/10.1111/jfbc.12628>
- Fletcher, R.L., 1995.** Epiphytism and fouling in *Gracilaria* cultivation: An overview. *Journal of Applied Phycology*, 7(4), 325–333.
<https://doi.org/10.1007/BF00004473>
- Fleurence, J., 1999.** Seaweed proteins: Biochemical, nutritional aspects and potential uses. *Trends in Food Science and Technology*, 10(1), 25–28.
[https://doi.org/10.1016/S0924-2244\(99\)00005-3](https://doi.org/10.1016/S0924-2244(99)00005-3)
- Fleurence, J., 2022.** Biotechnological processes applied to edible seaweeds: What perspectives? *Trends in Food Science and Technology*, 129, 617–620.
<https://doi.org/10.1016/j.tifs.2022.02.001>
- Fujiwara-Arasaki, T., Mino, N. and Kuroda, M., 1984.** The protein value in human nutrition of edible marine algae in Japan. In C. J. Bird and M. A. Ragan (Eds.), *Proceedings of the Eleventh International Seaweed Symposium, Qingdao, China*, 19–25 June 1983 (pp. 513–516). Springer.
https://doi.org/10.1007/978-94-017-1131-0_84
- Ganesan, P., Matsubara, K., Ohkubo, T., Tanaka, Y., Noda, K., Sugawara, T., & Hirata, T., 2010.** Anti-angiogenic effect of siphonaxanthin from green alga, *Codium fragile*. *Phytomedicine*, 17(14), 1140–1144.
<https://doi.org/10.1016/j.phymed.2010.05.005>
- Ganesan, P., Noda, K., Manabe, Y., Ohkubo, T., Tanaka, Y., Maoka, T., & Hirata, T., 2011.** Siphonaxanthin, a marine carotenoid from green algae, effectively induces apoptosis in human leukemia (HL-60) cells. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1810(5), 497–503.

- <https://doi.org/10.1016/j.bbagen.2011.02.008>
- Pinfold, G., 2013.** Socio-economic impact of aquaculture in Canada. *Fisheries and Oceans Canada Aquaculture Management Directorate. Gardner Pinfold Consultants Inc., Nova Scotia.*
- Moussavou, G., Kwak, D. H., Obiang-Obonou, B. W., Ogandaga Maranguy, C. A., Dinzouna-Boutamba, S. D., Lee, D. H., ... & Choo, Y. K., 2014.** Anticancer effects of different seaweeds on human colon and breast cancers. *Marine drugs*, 12(9), 4898-4911. <https://doi.org/10.3390/md12124898>
- <https://doi.org/10.3390/md12124898>
- Gómez-Guzmán, M., Rodríguez-Nogales, A., Algieri, F. and Gálvez, J., 2018.** Potential role of seaweed polyphenols in cardiovascular-associated disorders. *Marine Drugs*, 16(8), 250. <https://doi.org/10.3390/md16080250>
- Granada, L., Sousa, N., Lopes, S. and Lemos, M.F.L., 2016.** Is integrated multitrophic aquaculture the solution to the sector's major challenges?—A review. *Reviews in Aquaculture*, 8(3), 283–300. <https://doi.org/10.1111/raq.12093>
- Griffiths, M., Harrison, S.T.L., Smit, M. and Maharajh, D., 2016.** Major commercial products from micro- and macroalgae. In F. Bux and Y. Chisti (Eds.), *Algae biotechnology: Products and processes* (pp. 269–300). Springer.
- https://doi.org/10.1007/978-3-319-12334-9_12
- Grote, B., 2019.** Recent developments in aquaculture of *Palmaria palmata* (Linnaeus) (Weber and Mohr 1805): Cultivation and uses. *Reviews in Aquaculture*, 11(1), 25–41. <https://doi.org/10.1111/raq.12199>
- Guiry, M. D., Guiry, G. M., Morrison, L., Rindi, F., Miranda, S. V., Mathieson, A. C., & Garbary, D. J., 2014.** AlgaeBase: an on-line resource for algae. *Cryptogamie, Algologie*, 35(2), 105-115. <http://www.algaebase.org>
- Guiry, M.D. and Guiry, G.M., 2016.** *AlgaeBase*. National University of Ireland, Galway. <http://www.algaebase.org>
- Gullón, B., Gagaoua, M., Barba, F.J., Gullón, P., Zhang, W. and Lorenzo, J.M., 2020.** Seaweeds as promising resource of bioactive compounds: Overview of novel extraction strategies and design of tailored meat products. *Trends in Food Science and Technology*, 100, 1–18. <https://doi.org/10.1016/j.tifs.2020.03.039>
- Gurgel, C.F.D. and Lopez-Bautista, J.M., 2007.** Red algae. In *Encyclopedia of Life Sciences* (pp. 1–5). Wiley. <https://doi.org/10.1002/9780470015902.a0020368>
- Güroy, D., Güroy, B., Merrifield, D.L., Ergün, S., Tekinay, A.A. and Yigit, M., 2011.** Effect of dietary *Ulva* and *Spirulina* on weight loss and body composition of rainbow trout, *Oncorhynchus mykiss* (Walbaum),

- during a starvation period. *Journal of Animal Physiology and Animal Nutrition*, 95(3), 320–327. <https://doi.org/10.1111/j.1439-0396.2010.01060.x>
- Hafting, J.T., Critchley, A.T., Cornish, M.L., Hubley, S.A. and Archibald, A.F., 2012.** On-land cultivation of functional seaweed products for human usage. *Journal of Applied Phycology*, 24(3), 385–392. <https://doi.org/10.1007/s10811-011-9720-1>
- Hafting, J.T., Craigie, J.S., Stengel, D.B., Loureiro, R.R., Buschmann, A.H., Yarish, C., Edwards, M.D. and Critchley, A.T., 2015.** Prospects and challenges for industrial production of seaweed bioactives. *Journal of Phycology*, 51(5), 821–837. <https://doi.org/10.1111/jpy.12326>
- Hall, A.C., Fairclough, A.C., Mahadevan, K. and Paxman, J.R., 2012.** *Ascomyllum nodosum* enriched bread reduces subsequent energy intake with no effect on post-prandial glucose and cholesterol in healthy, overweight males: A pilot study. *Appetite*, 58(1), 379–386. <https://doi.org/10.1016/j.appet.2011.11.020>
- Hameed, S. and Ahmed, M., 1999.** Distribution and seasonal biomass of seaweeds on the rocky shore of Buleji, Karachi, Pakistan. *Pakistan Journal of Botany*, 31(1), 199–210.
- Hasselström, L., Visch, W., Gröndahl, F., Nylund, G.M. and Pavia, H., 2018.** The impact of seaweed cultivation on ecosystem services—A case study from the west coast of Sweden. *Marine Pollution Bulletin*, 133, 53–64. <https://doi.org/10.1016/j.marpolbul.2018.05.005>
- Hayashi, L., Hurtado, A. Q., Msuya, F.E., Bleicher-Lhonneur, G. and Critchley, A.T., 2010.** A review of *Kappaphycus* farming: Prospects and constraints. In A. Israel, R. Einav and J. Seckbach (Eds.), *Seaweeds and their role in globally changing environments* (pp. 251–283). Springer. https://doi.org/10.1007/978-90-481-8569-6_13
- Helmes, R.J.K., López-Contreras, A.M., Benoit, M., Abreu, H., Maguire, J., Moejes, F. and Burg, S.W.K., 2018.** Environmental impacts of experimental production of lactic acid for bioplastics from *Ulva* spp. *Sustainability*, 10(7), 2462. <https://doi.org/10.3390/su10072462>
- Hiraoka, M., Kinoshita, Y., Higa, M., Tsubaki, S., Monotilla, A. P., Onda, A., & Dan, A., 2020.** Fourfold daily growth rate in multicellular marine alga *Ulva meridionalis*. *Scientific reports*, 10(1), 12606. <https://doi.org/10.1038/s41598-020-69536-4>
- Holdt, S.L. and Kraan, S., 2011.** Bioactive compounds in seaweed: Functional food applications and legislation. *Journal of Applied Phycology*, 23(3), 543–597. <https://doi.org/10.1007/s10811-010-9632-5>

- Holman, B.W.B. and Malau-Aduli, A.E.O., 2013.** Spirulina as a livestock supplement and animal feed. *Journal of Animal Physiology and Animal Nutrition*, 97(4), 615–623. <https://doi.org/10.1111/jpn.12058>
- Houchi, S., Mahdadi, R., Khenchouche, A., Song, J., Zhang, W., Pang, X., Zhang, L., Sandalli, C. and Du, G., 2018.** Investigation of common chemical components and inhibitory effect on GES-type β -lactamase (GES-22) in methanolic extracts of Algerian seaweeds. *Microbial Pathogenesis*. <https://doi.org/10.1016/j.micpath.2018.10.034>
- Hu, C., Li, D., Chen, C., Ge, J., Muller-Karger, F. E., Liu, J., Yu, F. and He, M.X., 2010.** On the recurrent *Ulva prolifera* blooms in the Yellow Sea and East China Sea. *Journal of Geophysical Research: Oceans*, 115(C5), C05017. <https://doi.org/10.1029/2009JC005561>
- Hu, Y., Guangbo, K., Lina, W., Mengxue, G., Ping, W., Dong, Y. and He, H., 2021.** Current status of mining, modification, and application of cellulases in bioactive substance extraction. *Current Issues in Molecular Biology*, 43(2), 687–703. <https://doi.org/10.3390/cimb43020050>
- Huang, C.Y., Huang, C.Y., Yang, C.C., Lee, T.M. and Chang, J.S., 2022.** Hair growth-promoting effects of *Sargassum glaucescens* oligosaccharides extracts. *Journal of the Taiwan Institute of Chemical Engineers*, 134, 104307. <https://doi.org/10.1016/j.jtice.2022.104307>
- Huo, Y., Han, H., Hua, L., Wei, Z., Yu, K., Shi, H., Kim, J. K., Yarish, C. and He, P., 2016.** Tracing the origin of green macroalgal blooms based on the large-scale spatio-temporal distribution of *Ulva* microscopic propagules and settled mature *Ulva* vegetative thalli in coastal regions of the Yellow Sea, China. *Harmful Algae*, 59, 91–99. <https://doi.org/10.1016/j.hal.2016.09.004>
- Hurtado, A. Q., 2013.** Social and economic dimensions of carrageenan seaweed farming in the Philippines. In D. Valderrama, J. Cai, N. Hishamunda, & N. Ridler (Eds.), *Social and economic dimensions of carrageenan seaweed farming* (pp. 91–113). FAO Fisheries and Aquaculture Technical Paper No. 580. Rome: FAO. <http://hdl.handle.net/1969.1/1581938>
- Hwang, E.K., Gong, Y.G., Hwang, I.K., Park, E.J. and Park, C.S., 2013.** Cultivation of the two perennial brown algae *Ecklonia cava* and *E. stolonifera* for abalone feeds in Korea. *Journal of Applied Phycology*, 25(3), 825–829.
- Imeson, A., 2009.** *Food stabilisers, thickeners and gelling agents*. Wiley-Blackwell.
- Jiao, G., Yu, G., Zhang, J. and Ewart, H.S., 2011.** Chemical structures and bioactivities of sulfated polysaccharides from marine algae.

- Marine Drugs*, 9(2), 196–223.
<https://doi.org/10.3390/md9020196>
- Jones, L.A., Hiscock, K. and Connor, D.W., 2000.** *Marine habitat reviews: A summary of ecological requirements and sensitivity characteristics for the conservation and management of marine SACs.* Joint Nature Conservation Committee.
- Kadam, S.U., Álvarez, C., Tiwari, B.K. and O'Donnell, C.P., 2017.** Extraction and characterization of protein from Irish brown seaweed *Ascophyllum nodosum*. *Food Research International*, 99, 1021–1027.
<https://doi.org/10.1016/j.foodres.2016.05.019>
- Kaladharan, P., Kaliaperumal, N. and Ramalingam, J.R., 1998.** *Marine fishery information series. Marine Fisheries Information Service*, 157, 1–10.
- Kalasariya, H.S., Yadav, V.K., Yadav, K.K., Tirth, V., Algahtani, A., Islam, S., Gupta, N. and Jeon, B.H., 2021.** Seaweed-based molecules and their potential biological activities: An eco-sustainable cosmetic. *Molecules*, 26(17), 5313.
<https://doi.org/10.3390/molecules26175313>
- Kavalappa, Y. P., Rudresh, D. U., Gopal, S. S., Shivarudrappa, A. H., Stephen, N. M., Rangiah, K. and Ponesakki, G., 2019.** β -carotene isolated from the marine red alga, *Gracilaria* sp. potently attenuates the growth of human hepatocellular carcinoma (HepG2) cells by modulating multiple molecular pathways. *Journal of Functional Foods*, 52, 165–176.
<https://doi.org/10.1016/j.jff.2018.11.015>
- Kazir, M., Abuhassira, Y., Robin, A., Nahor, O., Luo, J., Israel, A., Golberg, A. and Livney, Y.D., 2019.** Extraction of proteins from two marine macroalgae, *Ulva* sp. and *Gracilaria* sp., for food application, and evaluating digestibility, amino acid composition and antioxidant properties of the protein concentrates. *Food Hydrocolloids*, 87, 194–203.
<https://doi.org/10.1016/j.foodhyd.2018.07.029>
- Kazłowski, B., Chiu, Y. H., Kazłowska, K., Pan, C.L. and Wu, C.J., 2012.** Prevention of Japanese encephalitis virus infections by low-degree polymerisation sulfated saccharides from *Gracilaria* sp. and *Monostroma nitidum*. *Food Chemistry*, 133(3), 866–874.
<https://doi.org/10.1016/j.foodchem.2012.01.106>
- Kendel, M., Wielgosz-Collin, G., Bertrand, S., Roussakis, C., Bourgougnon, N. and Bedoux, G., 2015.** Lipid composition, fatty acids and sterols in the seaweeds *Ulva armoricana* and *Solieria chordalis* from Brittany (France): An analysis from nutritional, chemotaxonomic, and antiproliferative activity perspectives. *Marine Drugs*, 13(9), 5606–5628.
<https://doi.org/10.3390/md13095606>

- Khan, M.N.A., Choi, J.S., Lee, M.C., Kim, E., Nam, T.J. and Fujii, H., 2008.** Anti-inflammatory activities of methanol extracts from various seaweed species. *Journal of Environmental Biology*, 29, 465–469. Retrieved from http://www.jeb.co.in/journal_issues/200807_jul08_spl/paper_09.pdf
- Khan, W., Rayirath, U. P., Subramanian, S., Jithesh, M. N., Rayorath, P., Hodges, D. M., & Prithiviraj, B., 2009.** Seaweed extracts as biostimulants of plant growth and development. *Journal of plant growth regulation*, 28(4), 386–399. <https://doi.org/10.1007/s00344-009-9103-x>
- Kim, J.K., Cho, M.L., Karnjanapratum, S., Shin, I.S. and You, S.G., 2011.** *In vitro* and *in vivo* immunomodulatory activity of sulfated polysaccharides from *Enteromorpha prolifera*. *International Journal of Biological Macromolecules*, 49(5), 1051–1058. <https://doi.org/10.1016/j.ijbiomac.2011.08.032>
- Kim, J.K., Kraemer, G.P. and Yarish, C., 2015.** Use of sugar kelp aquaculture in Long Island Sound and the Bronx River Estuary for nutrient extraction. *Marine Ecology Progress Series*, 531, 155–166. <https://doi.org/10.3354/meps11305>
- Kim, J.K., Yarish, C., Hwang, E.K., Park, M. and Kim, Y., 2017.** Seaweed aquaculture: cultivation technologies, challenges and its ecosystem services. *Algae*, 32(1), 1–13. <https://doi.org/10.4490/algae.2017.32.3.3>
- Kite-Powell, H.L., Ask, E., Augyte, S., Bailey, D., Decker, J., Goudey, C.A., Grebe, G., 2022.** Estimating production cost for large-scale seaweed farms. *Applied Phycology*, 3(1), 435–445. <https://doi.org/10.1080/26388081.2022.2111271>
- Knowler, D., Chopin, T., Martínez-Espíñeira, R., Neori, A., Nobre, A., Noce, A. and Reid, G., 2020.** The economics of integrated multi-trophic aquaculture: Where are we now and where do we need to go? *Reviews in Aquaculture*, 8, 1–21. <https://doi.org/10.1111/raq.12345>
- Knutsen, S., Myslabodski, D., Larsen, B. and Usov, A.I., 1994.** A modified system of nomenclature for red algal galactans. *Botanica Marina*, 37(2), 163–170. <https://doi.org/10.1515/botm.1994.37.2.163>
- Kolanjinathan, K., Ganesh, P. and Saranraj, P., 2014.** Pharmacological importance of seaweeds: A review. *World Journal of Fish and Marine Sciences*, 6(1), 1–15. <https://doi.org/10.5829/idosi.wjfm.2014.6.1.7420>
- Komatsu, T., Mizuno, S., Natheer, A., Kantachumpoo, A., Tanaka, K., Morimoto, A., Ajisaka, T., 2014.** Unusual distribution of floating seaweeds in the East China Sea in the early spring of 2012. *Journal of Applied Phycology*, 26, 1169–1179. <https://doi.org/10.1007/s10811-013-0236-5>

- Kopprio, G. A., Luyen, N. D., Cuong, L. H., Fricke, A., Kunzmann, A., Huong, L. M., & Gärdes, A. (2021). Bacterial community composition of the sea grape *Caulerpa lentillifera*: a comparison between healthy and diseased states. *BioRxiv*, 2021-06. <https://doi.org/10.1101/2021.06.30.450479>
- Koyande, A.K., Chew, K.W., Manickam, S., Chang, J.S. and Show, P.L., 2021. Emerging algal nanotechnology for high-value compounds: A direction to future food production. *Trends in Food Science and Technology*, 116, 290–302. <https://doi.org/10.1016/j.tifs.2021.07.026>
- Kraan, S., 2013. Pigments and minor compounds in algae. In H. Dominguez (Ed.), *Functional ingredients from algae for foods and nutraceuticals* (pp. 205–251) <https://doi.org/10.1533/9780857098689.1.205>.
- Krishnamurthy, V., 2005. *Seaweeds: Wonder plants of the sea* (30 P.).
- Krishnan, M. and Narayanakumar, R., 2013. Social and economic dimensions of carrageenan seaweed farming. *FAO Fisheries and Aquaculture Technical Paper*, 580, 163–184. <http://www.fao.org/docrep/019/i3344e/i3344e.pdf>
- Leiro, J.M., Castro, R., Arranz, J.A. and Lamas, J., 2007. Immunomodulating activities of acidic sulphated polysaccharides obtained from the seaweed *Ulva rigida* C. Agardh. *International Immunopharmacology*, 7(7), 879–888. <https://doi.org/10.1016/j.intimp.2007.02.007>
- Li, X., Norman, H.C., Kinley, R., Laurence, M., Wilmot, M., Bender, H., de Nys, R. and Tomkins, N., 2016. *Asparagopsis taxiformis* decreases enteric methane production from sheep. *Animal Production Science*, 58(4), 681–688. <https://doi.org/10.1071/AN15883>
- Li, X., Chen, Y., Gao, X., Wu, Y., El-Seedi, H.R., Cao, Y. and Zhao, C., 2021a. Antihyperuricemic effect of green alga *Ulva lactuca* ulvan through regulating urate transporters. *Journal of Agricultural and Food Chemistry*, 69(38), 11225–11235. <https://doi.org/10.1021/acs.jafc.1c03607>
- Liu, D., Keesing, J.K., Xing, Q. and Shi, P., 2009. World's largest macroalgal bloom caused by expansion of seaweed aquaculture in China. *Marine Pollution Bulletin*, 58(6), 888–895. <https://doi.org/10.1016/j.marpolbul.2009.01.013>
- Liu, J., Wu, S. Y., Chen, L., Li, Q. J., Shen, Y. Z., Jin, L., & Tong, H. B., 2020. Different extraction methods bring about distinct physicochemical properties and antioxidant activities of *Sargassum fusiforme* fucoidans. *International journal of biological macromolecules*, 155, 1385-1392.

- <https://doi.org/10.1016/j.ijbiomac.2020.03.084>
- Loeser, A.A., 1956.** Hormones and breast cancer (Letter to the Editor). *The Lancet*, ii, 961.
- López-Hortas, L., Flórez-Fernández, N., Torres, M.D., Ferreira-Anta, T., Casas, M.P., Balboa, E.M., Falqué, E. and Domínguez, H., 2021.** Applying seaweed compounds in cosmetics, cosmeceuticals and nutricosmetics. *Marine Drugs*, 19(10), 552. <https://doi.org/10.3390/md19100552>
- Los Ficocoloides en la Industria., 2022, December 9.** *Repositorio USIL*. Retrieved December 9, 2022, from <https://repositorio.usil.edu.pe/items/a25f3eba-1057-4142-a378-b1e8bf5b9ef3>
- Mabeau, S. and Fleurence, J., 1993.** Seaweed in food products: Biochemical and nutritional aspects. *Trends in Food Science and Technology*, 4(4), 103–107. [https://doi.org/10.1016/0924-2244\(93\)90011-N](https://doi.org/10.1016/0924-2244(93)90011-N)
- Macartain, P., Gill, C.I.R., Brooks, M., Campbell, R. and Rowland, I.R., 2007.** Nutritional value of edible seaweeds. *Nutrition Reviews*, 65(7), 535–543. <https://doi.org/10.1111/j.1753-4887.2007.tb00365.x>
- Marliana, Baba, B., Wan Mustapha, W.A. and Seng Joe, L., 2018.** Effect of extraction methods on the yield, fucose content and purity of fucoidan from *Sargassum* sp. obtained from Pulau Langkawi, Malaysia (Kesan kaedah pengekstrakan fukoidan terhadap hasil, kandungan fukosa dan ketulenan fukoidan daripada *Sargassum* sp. dari Pulau Langkawi, Malaysia. *Malaysian Journal of Analytical Sciences*, 22(1), 87–94. <https://doi.org/10.17576/mjas-2018-2201-11>
- Mayer, A.M., Rodríguez, A.D., Berlinck, R.G.S. and Fusetani, N., 2011.** Marine pharmacology in 2007–2008: Marine compounds with antibacterial, anticoagulant, antifungal, anti-inflammatory, antimalarial, antiprotozoal, antituberculosis, and antiviral activities; affecting the immune and nervous system, and other miscellaneous mechanisms of action. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology*, 153(2), 191–222. <https://doi.org/10.1016/j.cbpc.2010.11.008>
- MFF Pakistan, 2016.** *A handbook on Pakistan's coastal and marine resources* (78 P.). IUCN Pakistan.
- Misra, A. and Sinha, R., 1979.** Algae as drug plants in India. In H. A. Hoppe-Levring and T. Tanaka (Eds.), *Marine algae in pharmaceutical science* (pp. 237–242). Walter de Gruyter.
- Miyashita, K., 2009.** Function of marine carotenoids. *Forum of Nutrition*, 61, 136–146. <https://doi.org/10.1159/000212746>
- Moscicki, Z., Swift, M.R., Dewhurst, T., MacNicoll, M., Chambers, M., Tsukrov, I., MacAdam, N., 2024.** Design, deployment, and operation of

- an experimental offshore seaweed cultivation structure. *Aquacultural Engineering*, 105, 102413. <https://doi.org/10.1016/j.aquaeng.2024.102413>
- Msuya, F. E., 2013.** Social and economic dimensions of carrageenan seaweed farming in the United Republic of Tanzania. *Social and economic dimensions of carrageenan seaweed farming*, 115-146. FAO Fisheries and Aquaculture Technical Paper No. 580. Rome: FAO.
- Mumford, T.F. and Miura, A., 1988.** *Porphyra* as food: Cultivation and economics. In C. A. Lembi and J. R. Waaland (Eds.), *Algae and human affairs* (pp. 87–117). Cambridge University Press.
- Muraoka, D., 2004.** Seaweed resources as a source of carbon fixation. *Bulletin of Fisheries Research Agency, Supplement*, 1, 59–63.
- Nabti, E., Jha, B. and Hartmann, A., 2017.** Impact of seaweeds on agricultural crop production as biofertilizer. *International Journal of Environmental Science and Technology*, 14(5), 1119–1134. <https://doi.org/10.1007/s13762-016-1202-1>
- Nazim, K., Ahmed, M., Abbas, A. and Khan, M.U., 2012.** Quantitative description and multivariate analysis of flora and fauna of Buleji area of Karachi coast. *FUUAST Journal of Biology*, 2(1), 117–123.
- Needham, S. and Lentisco, A., 2013.** *Case study: Seaweed for a better life.* <https://openknowledge.fao.org/handle/20.500.14283/ar486e>
- Neori, A., Chopin, T., Troell, M., Buschmann, A.H., Kraemer, G.P., Halling, C. and Yarish, C., 2004.** Integrated aquaculture: Rationale, evolution and state of the art emphasizing seaweed biofiltration in modern mariculture. *Aquaculture*, 231(1–4), 361–391. <https://doi.org/10.1016/j.aquaculture.2003.11.015>
- Ohta, Y., Lee, J.B., Hayashi, K. and Hayashi, T., 2009.** Isolation of sulfated galactan from *Codium fragile* and its antiviral effect. *Biological and Pharmaceutical Bulletin*, 32(5), 892–898. <https://doi.org/10.1248/bpb.32.892>
- Okolie, C.L., Mason, B., Mohan, A., Pitts, N. and Udenigwe, C.C., 2020.** Extraction technology impacts on the structure-function relationship between sodium alginate extracts and their in vitro prebiotic activity. *Food Bioscience*, 37, 100672. <https://doi.org/10.1016/j.fbio.2020.100672>
- Oliveira, E.C., Alveal, K. and Anderson, R.J., 2000.** Mariculture of the agar-producing Gracilarioid red algae. *Reviews in Fishery Science*, 8(4), 345–377. <https://doi.org/10.1080/1064126009129188>
- Oliveira, C., Granja, S., Neves, N.M., Reis, R.L., Baltazar, F., Silva, T.H. and Martins, A., 2019.** Fucoidan from *Fucus vesiculosus* inhibits new blood vessel formation and breast

- tumor growth in vivo. *Carbohydrate Polymers*, 223, 115034. <https://doi.org/10.1016/j.carbpol.2019.115034>
- Olsen, E.K., Hansen, E., Isaksson, J. and Andersen, J.H., 2013.** Cellular antioxidant effect of four bromophenols from the red algae *Vertebrata lanosa*. *Marine Drugs*, 11(8), 2769–2784. <https://doi.org/10.3390/md11082769>
- Ondiviela, B., Losada, I.J., Lara, J.L., Maza, M., Galván, C., Bouma, T.J. and van Belzen, J., 2014.** The role of seagrasses in coastal protection in a changing climate. *Coastal Engineering*, 87, 158–168. <https://doi.org/10.1016/j.coastaleng.2013.11.005>
- Oyinlola, M.A., Reygondeau, G., Wabnitz, C.C.C., Troell, M. and Cheung, W.W.L., 2018.** Global estimation of areas with suitable environmental conditions for mariculture species. *PLoS ONE*, 13(1), e0191086. <https://doi.org/10.1371/journal.pone.0191086>
- Padam, B.S. and Chye, F.Y., 2020.** Chapter 2: Seaweed components, properties, and applications. In M. D. Torres, S. Kraan and H. Dominguez (Eds.), *Sustainable seaweed technologies: Cultivation, biorefinery, and applications* (pp. 33–87). <https://doi.org/10.1016/B978-0-12-817943-7.00002-0>
- Palanisamy, S., Vinosha, M., Rajasekar, P., Anjali, R., Sathiyaraj, G., Marudhupandi, T., Selvam, S., Prabhu, N.M. and You, S.G., 2019.** Antibacterial efficacy of a fucoidan fraction (Fu-F2) extracted from *Sargassum polycystum*. *International Journal of Biological Macromolecules*, 125, 485–495. <https://doi.org/10.1016/j.ijbiomac.2018.12.060>
- Pandey, A.K., Chauhan, O.P. and Semwal, A.D., 2020.** Seaweeds—A potential source for functional foods. *Defence Life Science Journal*, 5(4), 315–322. <https://doi.org/10.14429/dlsj.5.15503>
- Parthiban, C., Saranya, C., Girija, K., Hemalatha, A., Suresh, M. and Anantharaman, P., 2013.** Biochemical composition of some selected seaweeds from Tuticorin coast. *Advances in Applied Science Research*, 4(1), 362–366.
- Pati, M.P., Sharma, S.D., Nayak, L.A.K.S.H.M.A.N. and Panda, C.R., 2016.** Uses of seaweed and its application to human welfare: A review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 8(10), 12–20. <http://dx.doi.org/10.22159/ijpps.2016v8i10.12740>
- Paudel, P., Seong, S.H., Zhou, Y., Park, H.J., Jung, H.A. and Choi, J.S., 2019.** Anti-Alzheimer's disease activity of bromophenols from a red alga, *Symphyocladia latiuscula* (Harvey) Yamada. *ACS Omega*, 4(7), 12259–12270. <https://doi.org/10.1021/acsomega.9b01557>
- Paul, M.A., Gill, C.I.R., Campbell, R. and Rowland, I.R., 2007.** Nutritional

- value of edible seaweeds. *International Life Sciences Institute Special Paper*, 1, 535–543.
- Pereira, L., & Mesquita, J. F., 2003.** Carrageenophytes of occidental Portuguese coast: Spectroscopic analysis in eight carrageenophytes from Buarcos Bay. *Biomolecular Engineering*, 20(3), 217–222. [https://doi.org/10.1016/S1389-0344\(03\)00056-X](https://doi.org/10.1016/S1389-0344(03)00056-X)
- Pereira, R. and Yarish, C., 2008.** Mass production of marine macroalgae. In S. E. Jørgensen and B. D. Fath (Eds.), *Encyclopedia of ecology*, 3, pp. 2236–2247. <https://doi.org/10.1080/09670260701763393>
- Pereira, L., Meireles, F., Abreu, H. T., & Ribeiro-Claro, P. J., 2015.** A comparative analysis of carrageenans produced by underutilized versus industrially utilized macroalgae (Gigartinales, Rhodophyta). *Marine algae extracts: processes, products, and applications*, 277–294. <https://doi.org/10.1002/9783527679577.ch16>.
- Pereira, L., 2016.** *Edible seaweeds of the world*. <https://doi.org/10.3390/md14030052>
- Pérez, M.J., Falqué, E. and Domínguez, H., 2016.** Antimicrobial action of compounds from marine seaweed. *Marine Drugs*, 14(3), 52. <https://doi.org/10.3390/md14030052>
- Peteiro, C., Sánchez, N. and Martínez, B., 2016.** Mariculture of the Asian kelp *Undaria pinnatifida* and the native kelp *Saccharina latissima* along the Atlantic coast of Southern Europe: An overview. *Algal Research*, 15, 9–23. <https://doi.org/10.1016/j.algal.2016.01.012>
- Plaza, M., Cifuentes, A. and Ibáñez, E., 2008.** In the search of new functional food ingredients from algae. *Trends in Food Science and Technology*, 19(1), 31–39. <https://doi.org/10.1016/j.tifs.2007.07.012>
- Pradhan, B., Bhuyan, P.P., Patra, S., Nayak, R., Behera, P.K., Behera, C., Behera, A.K., Ki, J.S. and Jena, M., 2022.** Beneficial effects of seaweeds and seaweed-derived bioactive compounds: Current evidence and future prospective. *Biocatalysis and Agricultural Biotechnology*, 39, 102242. <https://doi.org/10.1016/j.bcab.2022.102242>
- Qi, H., Zhang, Q., Zhao, T., Li, Z. and Zhang, H., 2005.** Antioxidant activity of different sulfate content derivatives of polysaccharide extracted from *Ulva pertusa* (Chlorophyta) *in vitro*. *International Journal of Biological Macromolecules*, 37(4), 195–199. <https://doi.org/10.1016/j.ijbiomac.2005.10.008>
- Ra, C.H., Sunwoo, I.Y., Nguyen, T.H., Sukwang, P., Sirisuk, P., Jeong, G.T. and Kim, S.K., 2019.** Butanol and butyric acid production from *Saccharina japonica* by *Clostridium acetobutylicum* and *Clostridium*

- tyrobutyricum* with adaptive evolution. *Bioprocess and Biosystems Engineering*, 42(9), 1559–1567.
<https://doi.org/10.1007/s00449-019-02176-5>
- Rajasulochana, P., Krishnamoorthy, P. and Dhamotharan, R., 2012.** Biochemical investigation on red algae family of *Kappahycus* sp. *Journal of Chemical and Pharmaceutical Research*, 4(11), 4637–4641. <http://jocpr.com/vol4-iss1-2012/JCPR-2012-4-1-33-37.pdf>
- Rathmann, R., Szklo, A. and Schaeffer, R., 2010.** Land use competition for production of food and liquid biofuels: An analysis of the arguments in the current debate. *Renewable Energy*, 35(1), 14–22. <https://doi.org/10.1016/j.renene.2009.02.025>
- Reid, G.K., Lefebvre, S., Filgueira, R., Robinson, S.M.C., Broch, O.J., Dumas, A. and Chopin, T.B.R., 2020.** Performance measures and models for open-water integrated multi-trophic aquaculture. *Reviews in Aquaculture*, 12(1), 47–75. <https://doi.org/10.1111/raq.12304>
- Rey, F., Cartaxana, P., Melo, T., da Costa, E., Domingues, M.R. and Calado, R., 2020.** Domesticated populations of *Codium tomentosum* display lipid extracts with lower seasonal shifts than conspecifics from the wild—Relevance for biotechnological applications of this green seaweed. *Marine Drugs*, 18(4), 188. <https://doi.org/10.3390/md18040188>
- Rizvi, M., Farooqui, S., Khan, M. and Shameel, M., 2001.** Elemental composition and bioactivity of seaweeds from coastal areas of Karachi, Pakistan. *Journal of King Abdulaziz University: Science*, 12(1), 209–215. <https://doi.org/10.4197/mar.12-1.15>
- Roy, M.-C., Anguenot, R., Fillion, C., Beaulieu, M., Bérubé, J. and Richard, D., 2011.** Effect of a commercially-available algal phlorotannins extract on digestive enzymes and carbohydrate absorption *in vivo*. *Food Research International*, 44(10), 3026–3029. <https://doi.org/10.1016/j.foodres.2011.07.020>
- Sahoo, D. and Yarish, C., 2005.** Mariculture of seaweeds. In R. A. Andersen (Ed.), *Phycological methods: Algal culturing techniques* (pp. 219–237). Academic Press.
- Saifullah, S.M., 1973.** A preliminary survey of the standing crop of seaweeds from Karachi coast. *Botanica Marina*, 16(3), 139–144. <https://doi.org/10.1515/botm.1973.16.3.139>
- Saker, K. E., Allen, V.G., Fontenot, J.P., Bagley, C.P., Ivy, R.L., Evans, R.R. and Wester, D.B., 2001.** Tasco-Forage: II. Monocyte immune cell response and performance of beef steers grazing tall fescue treated with a seaweed extract. *Journal of Animal Science*, 79(5), 1022–1031. <https://doi.org/10.2527/2001.7951022x>
- Samocha, T.M., Fricker, J., Ali, A.M., Shpigel, M. and Neori, A., 2015.**

- Growth and nutrient uptake of the macroalga *Gracilaria tikvahiae* cultured with the shrimp *Litopenaeus vannamei* in an Integrated Multi-Trophic Aquaculture (IMTA) system. *Aquaculture*, 446, 263–271. <https://doi.org/10.1016/j.aquaculture.2015.05.041>
- Sasikumar, C. and Rengsam, R., 1994.** Role of red alga *Hypnea valentiae* (Gigartinales, Rhodophyta) in domestic effluent treatment at different light intensity and quality. *Indian Journal of Marine Sciences*, 23, 162–164. <http://pascal-francis.inist.fr/vibad/index.php?action=getRecordDetail&idt=3344400>
- Satpati, G. and Pal, R., 2011.** Biochemical composition and lipid characterization of marine green algae. *Journal of Algal Biomass Utilization*, 2(4), 10–13.
- Sekar, S. and Chandramohan, M., 2008.** Phycobiliproteins as a commodity: Trends in applied research, patents and commercialization. *Journal of Applied Phycology*, 20(2), 113–136. <https://doi.org/10.1007/s10811-007-9188-1>
- Shameel, M. and Tanaka, J., 1992.** A preliminary check-list of marine algae from the coast and inshore waters of Pakistan. In T. Nakaike and S. Malik (Eds.), *Cryptogamic flora of Pakistan*, 1, pp. 1–64. National Science Museum, Tokyo.
- Sharma, S.D., Pati, M.P., Nayak, L. and Panda, C.R., 2016.** Uses of seaweed and its application to human welfare: A review. *International Journal of Pharmacy and Pharmaceutical Sciences*, 8(10), 12–20. <https://doi.org/10.22159/ijpps.2016v8i10.12740>
- Shimazu, T., Borjigin, L., Katoh, K., Roh, S. G., Kitazawa, H., Abe, K., & Suzuki, K., 2019.** Addition of Wakame seaweed (*Undaria pinnatifida*) stalk to animal feed enhances immune response and improves intestinal microflora in pigs. *Animal Science Journal*, 90(9), 1248–1260. <https://doi.org/10.1111/asj.13262>
- Sho, H., 2001.** History and characteristics of Okinawan longevity food. *Asia Pacific Journal of Clinical Nutrition*, 10(2), 159–164. <https://doi.org/10.1111/j.1440-6047.2001.00235.x>
- Siddhanta, A.K., Shanmugam, M., Mody, K.H., Goswami, A.M. and Ramavat, B.K., 1999.** Sulphated polysaccharides of *Codium dwarkense* Boergs. from the west coast of India: Chemical composition and blood anticoagulant activity. *International Journal of Biological Macromolecules*, 26(2–3), 151–154. [https://doi.org/10.1016/S0141-8130\(99\)00079-3](https://doi.org/10.1016/S0141-8130(99)00079-3)
- Singh, S., Singh, G. and Arya, S.K., 2018.** Mannans: An overview of properties and application in food products. *International Journal of Biological Macromolecules*, 119, 79–95.

- <https://doi.org/10.1016/j.ijbiomac.2018.07.130>
- Soares, C., Švarc-Gajić, J., Oliva-Teles, M.T., Pinto, E., Nastić, N., Savić, S., Almeida, A. and Delerue-Matos, C., 2020.** Mineral composition of subcritical water extracts of *Saccorhiza polyschides*, a brown seaweed used as fertilizer in the north of Portugal. *Journal of Marine Science and Engineering*, 8(4), 244. <https://doi.org/10.3390/jmse8040244>
- Soler-Vila, A., Coughlan, S., Guiry, M. D. and Kraan, S., 2009.** The red alga *Porphyra dioica* as a fish-feed ingredient for rainbow trout (*Oncorhynchus mykiss*): Effects on growth, feed efficiency, and carcass composition. *Journal of Applied Phycology*, 21, 617–624. <https://doi.org/10.1007/s10811-008-9400-5>
- Soto, D. and Wurmman, C., 2019.** Offshore aquaculture: A needed new frontier for farmed fish at sea. In *The future of ocean governance and capacity development* (pp. 379–384. Brill | Nijhoff.
- Spolaore, P., Joannis-Cassan, C., Duran, E. and Isambert, A., 2006.** Commercial applications of microalgae. *Journal of Bioscience and Bioengineering*, 101(2), 87–96. <https://doi.org/10.1263/jbb.101.87>
- Suárez, Y., González, L., Cuadrado, A., Berciano, M., Lafarga, M. and Muñoz, A., 2003.** A new marine-derived compound induces oncosis in human prostate and breast cancer cells. *Molecular Cancer Therapeutics*, 2(9), 863–872.
- Suganthi, N., Pandian, S. K. and Davi, K.P., 2010.** Neuroprotective effect of seaweeds inhabiting South India coastal area (Hare Island, Gulf of Mannar Marine Biosphere Reserve): Cholinesterase inhibitory effect of *Hypnea valentiae* and *Ulva reticulata*. *Neuroscience Letters*, 468(3), 216–219. <https://doi.org/10.1016/j.neulet.2009.11.001>
- Suganya, T., Varman, M., Masjuki, H.H. and Renganathan, S., 2016.** Macroalgae and microalgae as a potential source for commercial applications along with biofuels production: A biorefinery approach. *Renewable and Sustainable Energy Reviews*, 55, 909–941. <https://doi.org/10.1016/j.rser.2015.11.026>
- Sulaiman, O.O., Magee, A., Nik, W.B.W., Saharuddin, A.H. and Kader, A.S.A., 2012.** Design and model testing of offshore aquaculture floating structure for seaweed oceanic plantation. *Biosciences Biotechnology Research Asia*, 9(2), 477–494.
- Suyo, J.G.B., Le Masson, V., Shaxson, L., Luhan, M.R.J. and Hurtado, A.Q., 2020.** A social network analysis of the Philippine seaweed farming industry: Unravelling the web. *Marine Policy*, 118, 104007. <https://doi.org/10.1016/j.marpol.2020.104007>
- Suyo, J. G. B., Le Masson, V., Shaxson, L., Luhan, M. R. J. and**

- Hurtado, A. Q., 2021.** Navigating risks and uncertainties: Risk perceptions and risk management strategies in the Philippine seaweed industry. *Marine Policy*, 126, 104408.
<https://doi.org/10.1016/j.marpol.2020.104408>
- Tabassum, M.R., Xia, A. and Murphy, J.D., 2017.** Potential of seaweed as a feedstock for renewable gaseous fuel production in Ireland. *Renewable and Sustainable Energy Reviews*, 68, 136–146.
<https://doi.org/10.1016/j.rser.2016.09.044>
- Tanaka, Y., Ashaari, A., Mohamad, F.S. and Lamit, N., 2020.** Bioremediation potential of tropical seaweeds in aquaculture: Low-salinity tolerance, phosphorus content, and production of UV-absorbing compounds. *Aquaculture*, 518, 734853.
<https://doi.org/10.1016/j.aquaculture.2019.734853>
- Taylor, V.F., Li, Z., Sayarath, V., Palys, T.J., Morse, K.R., Scholzbright, R.A. and Karagas, M.R., 2017.** Distinct arsenic metabolites following seaweed consumption in humans. *Scientific Reports*, 7, 3920.
<https://doi.org/10.1038/s41598-017-04220-0>
- Thanigaivel, S., Chandrasekaran, N., Mukherjee, A. and Thomas, J., 2016.** Seaweeds as an alternative therapeutic source for aquatic disease management. *Aquaculture*, 464, 529–536.
<https://doi.org/10.1016/j.aquaculture.2016.08.015>
- Tibbetts, S.M., Milley, J.E. and Lall, S.P., 2016.** Nutritional quality of some wild and cultivated seaweeds: Nutrient composition, total phenolic content and in vitro digestibility. *Journal of Applied Phycology*, 28, 3575–3585.
<https://doi.org/10.1007/s10811-016-0835-7>
- Troell, M., Jonell, M., John, P. and Henriksson, G., 2017.** Ocean space for seafood. *Nature Ecology and Evolution*, 1, 1224–1225.
<https://doi.org/10.1038/s41559-017-0257-2>
- Tzachor, A., 2019.** The future of feed: Integrating technologies to decouple feed production from environmental impacts. *Industrial Biotechnology*, 15(2), 52–62.
<https://doi.org/10.1089/ind.2019.29162.atz>
- Ugent, D. and Tindall, D.R., 1997.** *Sargassum: An edible seaweed*. In T. D. Dillehay (Ed.), *Monte Verde: A Late Pleistocene settlement in Chile. Volume 2: The archaeological context and interpretation*, 2, pp. 911–914. Smithsonian Institution Press.
- Vairappan, C.S., Chung, C. S., Hurtado, A.Q., Soya, F.E., Lhonneur, G.B. and Critchley, A., 2008.** Distribution and symptoms of epiphyte infection in major carrageenophyte-producing farms. *Journal of Applied Phycology*, 20(5), 477–483.

- <https://doi.org/10.1007/s10811-007-9291-3>
- Valderrama, D., Cai, J., Hishamunda, N., Ridler, N., Neish, I.C., Hurtado, A. Q., Msuya, F.E., Krishnan, M., Narayanakumar, R., Kronen, M. and others., 2015.** The economics of *Kappaphycus* seaweed cultivation in developing countries: A comparative analysis of farming systems. *Aquaculture Economics and Management*, 19(2), 251–277. <https://doi.org/10.1080/13657305.2015.1024340>
- Vijn, S., Compart, D. P., Dutta, N., Foukis, A., Hess, M., Hristov, A. N. and Kurt, T. D., 2020.** Key considerations for the use of seaweed to reduce enteric methane emissions from cattle. *Frontiers in Veterinary Science*, 7, 597430. <https://doi.org/10.3389/fvets.2020.597430>
- Vlaisavljević, S., Rašeta, M., Berežni, S., Passamonti, S. and Tramer, F., 2021.** Four selected commercial seaweeds: Biologically active compounds, antioxidant and cytotoxic properties. *International Journal of Food Sciences and Nutrition*, 72(6), 757-766. <https://doi.org/10.1080/09637486.2020.1866503>
- Wang, M., Chen, L. and Zhang, Z., 2021.** Potential applications of alginate oligosaccharides for biomedicine—a mini review. *Carbohydrate Polymers*, 271, 118408. <https://doi.org/10.1016/j.carbpol.2021.118408>
- Wang, S., Wang, W., Hou, L., Qin, L., He, M., Li, W. and Mao, W., 2020.** A sulfated glucuronorhamnan from the green seaweed *Monostroma nitidum*: Characteristics of its structure and antiviral activity. *Carbohydrate Polymers*, 227, 115280. <https://doi.org/10.1016/j.carbpol.2019.115280>
- Wang, X., He, L., Ma, Y., Huan, L., Wang, Y., Xia, B. and Wang, G., 2020b.** Economically important red algae resources along the Chinese coast: History, status, and prospects for their utilization. *Algal Research*, 46, 101817. <https://doi.org/10.1016/j.algal.2020.101817>
- Wassie, T., Niu, K., Xie, C., Wang, H. and Xin, W., 2021.** Extraction techniques, biological activities and health benefits of marine algae *Enteromorpha prolifera* polysaccharide. *Frontiers in Nutrition*, 8, 747928. <https://doi.org/10.3389/fnut.2021.747928>
- White, W. L., & Wilson, P., 2015.** World seaweed utilization. In *Seaweed sustainability* (pp. 7-25). <https://doi.org/10.1016/B978-0-12-418697-2.00002-7>
- Williams, P.J.L.B. and Laurens, L.M., 2010.** Microalgae as biodiesel and biomass feedstocks: Review and analysis of the biochemistry, energetics and economics. *Energy and environmental science*, 3(5), 554-590. <https://doi.org/10.1039/B924978H>

- Wiltshire, K.H., Tanner, J.E., Gurgel, C.F.D. and Deveney, M.R., 2015.** Feasibility study for integrated multitrophic aquaculture in southern Australia. *Report to the Fisheries Research and Development Corporation; SARDI: Adelaide, Australia*, 883, F2015. <http://www.pir.sa.gov.au/research>
- Wu, D., Chen, Y., Wan, X., Liu, D., Wen, Y., Chen, X. and Zhao, C., 2020.** Structural characterization and hypoglycemic effect of green alga *Ulva lactuca* oligosaccharide by regulating microRNAs in *Caenorhabditis elegans*. *Algal Research*, 51, 102083. <https://doi.org/10.1016/j.algal.2020.102083>
- Yu, B., Bi, D., Yao, L., Li, T., Gu, L., Xu, H., & Xu, X. (2020).** The inhibitory activity of alginate against allergic reactions in an ovalbumin-induced mouse model. *Food & function*, 11(3), 2704-2713. <https://doi.org/10.1039/D0FO00170H>
- Zhang, J., Kim, J. K., Yarish, C., & He, P., 2016.** The expansion of *Ulva prolifera* OF Müller macroalgal blooms in the Yellow Sea, PR China, through asexual reproduction. *Marine pollution bulletin*, 104(1-2), 101-106. <https://doi.org/10.1016/j.marpolbul.2016.01.056>
- Zhang, L., Liao, W., Huang, Y., Wen, Y., Chu, Y. and Zhao, C., 2022.** Global seaweed farming and processing in the past 20 years. *Food Production, Processing and Nutrition*, 4(1), 23. <https://doi.org/10.1186/s43014-022-00103-2>
- Zheng, Y., Jin, R., Zhang, X., Wang, Q. and Wu, J., 2019.** The considerable environmental benefits of seaweed aquaculture in China. *Stochastic Environmental Research and Risk Assessment*, 33(4), 1203-1221. <https://doi.org/10.1007/s00477-019-01685-z>
- Zubia, M., Freile-Pelegrín, Y. and Robledo, D., 2014.** Photosynthesis, pigment composition and antioxidant defences in the red alga *Gracilariopsis tenuifrons* (Gracilariaceae, Rhodophyta) under environmental stress. *Journal of Applied Phycology*, 26(5), 2001–2010. <https://doi.org/10.1007/s10811-014-0325-3>
- Zubia, M., Draisma, S.G., Morrissey, K.L., Varela-Álvarez, E. and De Clerck, O., 2020.** Concise review of the genus *Caulerpa* JV Lamouroux. *Journal of Applied Phycology*, 32(1), 23-39. <https://doi.org/10.1007/s10811-019-01868-9>